

# Incorporation of Tetrahydrocannabinol into a Mixed Drug Substances Panel - Application to Three Commercially Available Oral Fluid Collection Devices

Rob Lee and Michelle Wood Waters Corporation, Wilmslow, UK



#### GOAL

To develop a single sample preparation method, using Waters® Oasis® MCX 96-well µElution™ Plate, for the analysis of a mixed drug panel containing Tetrahydrocannabinol (THC) that can be applied to different oral fluid collection devices.

### BACKGROUND

The requirement to analyze drugs at low levels in oral fluid has become an important requirement for many forensic laboratories around the world. Many different collection devices are commercially available which offer a simple way of collecting oral fluid samples in a non-invasive, yet supervised, manner. These devices provide the laboratory with either neat oral fluid or oral fluid that has been diluted in a variety of different preservative buffers. The lack of standardization for collection devices highlights the need for a sample preparation strategy suitable for all analytes that can be used with all the commonly available devices. The most commonly detected drug in these schemes is THC, however many other drug classes such as opiates, opioids, amphetamines, and benzodiazepines must also be measured, and as sample volume

## Inclusion of THC into an oral fluid mixed analyte panel using a single sample preparation method.

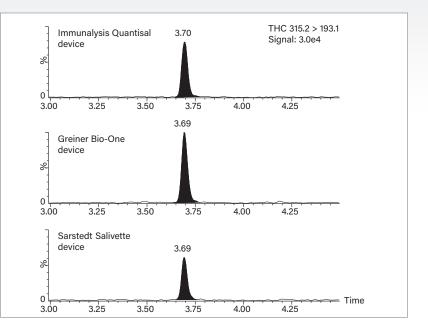


Figure 1. Chromatograms showing the quantifier MRM transition for THC spiked into oral fluid collected by the three different collection devices. The samples were prepared to give a concentration equivalent to 1 ng/mL in neat oral fluid.

can be limited, this has to be performed using a single sample preparation method. Oasis MCX µElution offers the ability to extract all of these analytes from limited volumes of this complex matrix with sufficient efficiency to meet current guidelines such as those applied by the European Workplace Drug Testing Society (EWDTS).<sup>1</sup>

### THE SOLUTION

The oral fluid collection devices tested in this study were the Salivette® saliva collection device from Sarstedt, the Quantisal<sup>™</sup> Oral Fluid Collection Device from Immunalysis, and the Saliva Collection System from Greiner Bio-One. Control oral fluid samples were collected as per the manufacturer's instructions and spiked with a mixture of 27 illicit or prescription substances commonly measured in oral fluid.

# [TECHNOLOGY BRIEF]

The analytes were extracted from the matrix using a simple Oasis MCX 96-well µElution Plate (P/N 186001830BA) protocol, based on previously reported methods.<sup>2,3</sup> The volume of sample loaded onto the plate for each collection device equated to the same volume (<75 µL) of neat oral fluid. The data was collected using a dual transition MRM method (quantifier and qualifier ions for each analyte) and processed using the TargetLynx<sup>™</sup> Application Manager.

The chemical properties of THC are very different to the other analytes in the panel, and as such, THC requires a different chromatographic gradient to be able to meet the 2 ng/mL cut-off for confirmation tests recommended by the EWDTS guidelines for oral fluid analysis, as seen in Figure 1.

The eluant from the Oasis MCX  $\mu$ Elution Plate was split into two equal aliquots in a 96-well Sample Collection Plate; 700  $\mu$ L Round well (P/N 186005837). Following evaporation under nitrogen the separate aliquots were reconstituted in one of two alternative solvents. One aliquot was reconstituted in 50% acetonitrile and analyzed specifically for THC; the other aliquot was reconstituted in 5% acetonitrile containing 0.1% blank human plasma and used to determine all the other compounds. Figure 2 shows the chromatographic separation of all the analytes (except THC) scaled to the most intense.

To avoid the need for column switching, the two analytical methods were run on the same ACQUITY<sup>®</sup> UPLC<sup>®</sup> BEH  $C_{18}$  Column (<u>P/N 186002352</u>) using the same mobile phases, allowing for the methods to be run consecutively.

The high sensitivity Xevo® TQ-S micro mass spectrometer in conjunction with the ACQUITY UPLC I-Class System (FTN) is ideally suited to this application as the analyte concentrations in oral fluid can be relatively low in comparison to other biological matrices.

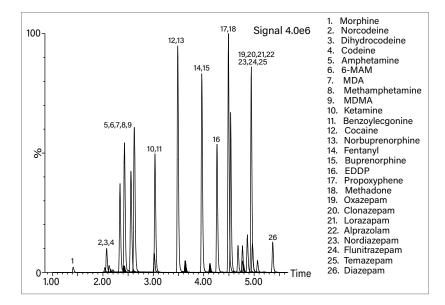


Figure 2. Chromatogram showing the quantifier MRM transitions for all analytes (except THC) spiked into oral fluid collected using the Immunalysis Quantisal Collection Device. The sample was prepared to give a concentration equivalent to 2 ng/mL in neat oral fluid. 6-MAM (peak #6), norbuprenorphine (peak #13), and buprenorphine (peak #15) are highlighted as compounds with EWDTS cut-off concentrations ( $\leq 2$  ng/mL).

#### SUMMARY

The presence of multiple chemical classes with very different chemical properties in a drug panel, in combination with the limited sample volume that is often provided by oral fluid collection devices, creates a very challenging application. The use of a single Oasis MCX µElution sample preparation method and two UPLC-MS/MS methods allow for the analysis of a mixed drug panel, which includes THC, in less than 15 minutes and at concentrations which meet current guidelines such as those applied by the EWDTS. This sample preparation method can be easily automated to increase sample throughput.



### References

- 1. European Workplace Drug Testing Society Guidelines. http://www.ewdts.org (accessed 29 June 2017).
- Danaceau et al. Direct Analysis of Opioids and Metabolites in Oral Fluid by Mixed-Mode µElution SPE Combined with UPLC-MS/MS for Forensic Toxicology. 2013. Waters Application Note. <u>720004838EN</u>.
- 3. Lee et al. Using UPLC-MS/MS for the Quantitation of Illicit or Prescription Drugs in Preserved Oral Fluid. 2016. Waters Application Note. 720005584EN.

For Research Use Only. Not for use in diagnostic procedures.



Waters, The Science of What's Possible, Oasis, ACQUITY, UPLC, and Xevo are registered trademarks of Waters Corporation. µElution and TargetLynx are trademarks of Waters Corporation. All other trademarks are the property of their respective owners.

©2017 Waters Corporation. Produced in the U.S.A. August 2017 720006081EN TC-PDF

#### Waters Corporation

34 Maple Street Milford, MA 01757 U.S.A. T: 1 508 478 2000 F: 1 508 872 1990 www.waters.com