

Oxygen Attachment Dissociation MS/MS for Differentiation between Cis and Trans Fatty Acids

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1. Overview

We have developed novel MS/MS techniques:

HAD-MS/MS

Hydrogen Attachment/Abstraction Dissociation

*Takahashi, H. et al. Anal. Chem., 2016, 88, 3810-

OAD-MS/MS

Oxygen Attachment **D**issociation

*Takahashi, H. et al. Anal. Chem., 2018, 56, 7230-

2. Introduction

Instead of using the collision gas (Ar/N_2) for CID-MS/MS, neutral reactive radical gas is introduced into an ion trap.



3. Methods and Materials

HAD-MS/MS and OAD-MS/MS were performed using a LC-ESI-MS and a prototype MALDI-IT-TOF-MS. H• was generated by passing H_2 gas through a heated tungsten capillary. OH • and O were generated by a microwave discharge of water vapor.



Fig.1. Schematic diagram for HAD-MS/MS and OAD-MS/MS using the LC-ESI-MS and MADLI-IT-TOF-MS.

Lipid name (Phosphatidylcholine:PC)	Exact Mass
PC(18:0/18:0)	789.625
PC (18:1(9Z)/16:0)	759.578
PC (18:0/18:1(9Z))	787.609
PC (14:1(9Z)/14:1(9Z))	673.468
PC (16:1(9E)/16:1(9E))	729.531
PC (18:1(6Z)/18:1(6Z))	785.593
PC (18:3(9Z,12Z,15Z)/18:3(9Z,12Z,15Z))	777.531
PC (16:0/20:4(5Z,8Z,11Z,14Z))	781.562

Table 1. List of model phospholipids used in this study.

4. Result

4-1. <u>HAD-MS/MS</u> for C=C and *sn*-position determination

The HAD-MS/MS spectrum of the model phospholipid shows a continuous series of fragment ions with the mass difference of 14 Da, which represents a CH_2 group. Meanwhile, the fragment ions corresponding to the C=C position shows a characteristic mass difference of 12 Da. These diagnostic product ions enables the structural analysis of C=C isomers.



4-2. <u>OAD-MS/MS</u> for C=C position determination The OAD-MS/MS spectrum provides the C=C specific fragmentation. The methylene bridges adjacent to C=C positions were selectively dissociated, accompanied by oxidation of the double bonds.



et al. Anal. Chem., 2016, 88, 3810





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4-3. Mixture of C=C position isomers

Since the OAD-MS/MS provides straightforward double-bond specific fragmentation, OAD can be applied to the analysis for mixture of C=C position isomers, based on the product ion intensity



Fig.4. OAD-MS/MS spectrum for the mixture of C=C position isomers of PC 18:1(9Z) and PC 18:1 (6Z).

4-4. LC-OAD-MS/MS for C=C position assignment

Auto-MS/MS mode with a cycle time of 1 s was used for LC-OAD-MS/MS. Double bond position assignment in a mixture of eight standard samples with multiple saturated fatty acyl chains was carried out successfully.







Fig. 5. (A) TIC of phospholipid mixture and (B)-(I) OAD-MS/MS spectra of each phospholipid obtained with the acquisition time of 0.5 s.

Fig.3. OAD-MS/MS spectrum of PC 16:0/20:4(5Z,8Z,11Z,14Z).

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5. Differentiation between Cis and Trans fatty acids

Focusing on the intensity of non-dissociative oxidized ion [M+H+O]⁺, cis and trans fatty acids can be differentiated. The intensity of [M+H+O]⁺ of *trans* fatty acid is higher than that of *cis* fatty acid by a factor of around 2. Since the reproducibility of the peak intensity is quite good, this technique can be applied to cis/trans mixture (Fig.7.).

OAD-MS/MS spectrum

Fig.6. Comparison between the intensity of [M+H+O]⁺ of *trans* fatty acid and that of *cis* fatty acid.

6. Conclusions

We have demonstrated OAD-MS/MS, in combination with LC, for the detailed structural analyses of several phospholipids. OAD-MS/MS is a promising analytical technique for the assignment of C=C positions and the discrimination of cis/trans fatty acids.

Fig.7. Differentiation between cis and trans fatty acids for cis/trans mixture.

d Group (PC, PS, PE, PI, PG)、Chain length、Number of C=C	CID	HAD OAD
$ \begin{array}{c} PC \\ \hline 0 \\ \hline 0 \\ P \\ \hline 0 \\ \hline 0 \\ P \\ \hline 0 \\ \hline$	¥	
/sn-2/sn-3 position		~
Position, Acyl chain modification	×	<u>AI</u>
ans		

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