

Achieve broad lipid quantitation using a high-throughput targeted lipidomics method

LC-based approach for lipid class separation and quantitation on high sensitivity SCIEX Triple Quad™ and QTRAP® Systems

Mackenzie Pearson, Santosh Kumar Kapil, Paul Norris, Christie Hunter
SCIEX, USA

Direct infusion ‘shotgun’ lipidomics is an established approach for broad-based lipidomic analysis¹. It is fast and simple, but it can suffer from inherent ion suppression effects, and due to the complexity of the lipidome and the extensive isobaric overlap, there is potential for ambiguous identification². Reverse and normal phase LC strategies coupled with MS are also frequently used for lipidomics analyses. These strategies separate lipids based on their physico-chemical properties, but the huge diversity of lipid molecular species makes development of standardized “all-inclusive” methods challenging, especially when quantitation is desired³. Furthermore, discovery-based approaches such as information dependent acquisition (IDA) suffer from poor reproducibility, making quantitation unreliable⁴.

An efficient way to maximize sensitivity and specificity is through targeted lipidomics using HPLC and Multiple Reaction Monitoring (MRM) with a triple quadrupole-based mass spectrometer. A LC



separation that can separate lipids into classes and subclasses is key to such an assay⁵.

Here, a targeted method is described using qualified MRM transitions and commercially available lipid internal standard mixtures, which provides quantitative measurement of ~1900 different lipid molecular species (Figure 1). The target list is can be shortened for a class-specific study or expandable to include new lipid classes.

Benefits of LC-MRM and SCIEX Triple Quad or QTRAP Systems for Targeted Lipidomics

- Optimized chromatography separates lipids by class, which reduces inter-class isobaric interferences (17 mins per sample)
- Individual lipid classes elute within a narrow RT window, making it simple to assign the RTs and implement time scheduled MRM to achieve the detection of ~ 1900 lipids
- SCIEX Systems offer wide dynamic range, fast polarity switching scan (< 5msec), as well as high sensitivity even at high acquisition rates (3-5 msec dwell times)
- The MRM scan mode maximizes the sensitivity and the specificity, which enables rapid and facile quantitation
- This method utilizes internal standards for developing time scheduled MRM methods as well as for downstream relative quantitative analysis
- SCIEX OS Software automatically processes data using a pre-built processing method

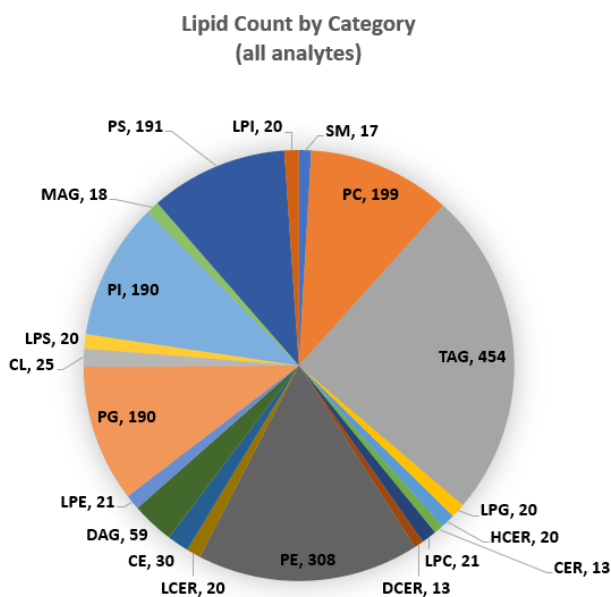


Figure 1. Overview of the LC-Based Targeted MRM Assay. The targeted lipidomics analysis via LC-MS/MS provides for rapid identification and quantification of a broad spectrum of lipid molecular species in a variety of matrices.

Methods

Sample Preparation: Plasma extract (NIST, 1g/100mL) is a commercially available lipid mixture that contains diverse lipid species. Available internal standards for retention time determination and for quantitation include the Avanti SPLASH standard mixture and the SCIEX Lipidzyer™ Platform kits⁶.

LC-MS Analysis: The LC separation was performed using a ExionLC™ System consisting of a binary high pressure mixing gradient pump with degasser, a thermostated autosampler, and a column oven. Data can be acquired using either the QTRAP® 6500+ System, equipped with an IonDrive™ Turbo V Ion Source, was operated in low mass mode with Analyst® Software or the SCIEX Triple Quad 7500 System equipped with an OptiFlow® Pro Ion Source and operated using SCIEX OS Software. Retention times are first roughly determined using either the NIST plasma sample spiked with internal standards or a representative matrix sample and the unscheduled MRM method. Analysis of this preliminary data enables the development of a time-scheduled MRM method leveraging the enhanced features in the Scheduled MRM™ Algorithm. Details on the chromatographic and MS methods can be obtained from the Comprehensive Targeted Method for Global Lipidomics Screening⁷.

Data Processing: All data was processed using SCIEX OS Software using AutoPeak algorithm. Automated computation of the time scheduled MRM methods was performed using the sMRM Pro Builder v1.3.⁸

Positive ion mode strategy: (M+H)	PC 34:2	(758.5/184.1)
Negative ion mode strategy: (M+CH3COO)	PC (14:0_20:2)	(816.6/307.3)
	PC (14:0_20:2)	(816.6/309.3)
	PC (14:0_20:2)	(816.6/279.2)
	PC (14:0_20:2)	(816.6/281.2)

Figure 2. Negative Mode Strategy for More Complete Phospholipid Identification. In a typical MRM assay, MRMs that denote the phospholipid head groups in positive mode are used. Here, negative mode MRMs are used that provide information on fatty acid composition to provide information at the molecular species level.

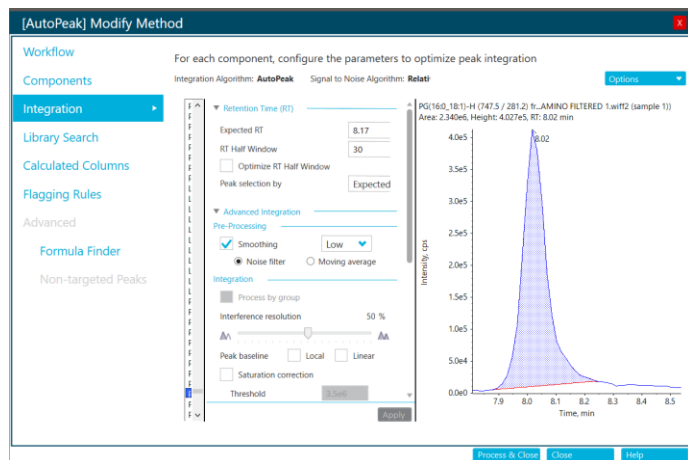


Figure 3. SCIEX OS Software for Data Processing of Lipid MRM Data. Data acquired using either the unscheduled or time scheduled MRM methods can be quickly processed using SCIEX OS Software. Export of retention times, peak areas and peak widths are then used to fully optimize the final method.

MRM Approach in Lipid Identification

Here a large panel of MRM transitions have been developed to provide a targeted assay for a broad range of lipid species. The targeted list is composed of ~1900 lipid molecular species, including PC/LPC, PE/LPE, PS/LPS, PI/LPI, PG/LPG, MAG, DAG, TAG, CL, CE, CER, DCER, HCER, LHCER and SM. Lipid classes currently included in assay provide good coverage of lipids commonly found in mammalian plasma, tissues and cells. Additional lipids can be added to the assay depending on matrix as workflow is easily extensible.

Most lipid classes and categories are acquired in the positive ion mode. However, for the targeting of phospholipids (PL), rather than using the more common approach via the head group loss in positive ion mode, this approach uses negative ion mode via the loss of fatty acid to identify the PL (PC, PE, PS, PI, PG) at the fatty acid composition molecular species level. As shown in Figure 2, the PC 34:2 can only be identified at the class level using the positive ion mode while the specific molecular species can be figured using the negative ion mode. Use of fast polarity switching on the SCIEX Systems allows monitoring of both positive and negative MRMs providing a key advantage for this method. This level of molecular species specificity is often key to understanding the specific biology.

Streamlined Method Development

As the number of lipids covered in this method is large, it is important to use the Scheduled MRM Algorithm approach during acquisition of biological dataset for highest quality quantitation results. To streamline the development of this final assay, an Excel based sMRM Pro Builder⁸ has been developed to assist the user during the assay refinement steps.

First the user collects MRM data on a representative matrix (with or without internal standards) for the upcoming biological study (3-5 replicates). Results from the first pass unscheduled MRM acquisitions are processed with SCIEX OS Software (Figure 3) and then entered into the sMRM Pro Builder and an initial rough approximation of the retention times is determined. Next 5-10 replicate injections are performed on the matrix using a preliminary time scheduled MRM method, ideally a pooled sample from the biological samples to be run. Data analysis here provides information on peak width, RT variance, and lipid signal and the sMRM Pro Builder computes a final optimized Scheduled MRM Acquisition method (Figure 4).

After method construction, the Scheduled MRM Algorithm Concurrency tool⁹ for Analyst Software or the sMRM Summary build into SCIEX OS Software¹⁰ can be used to check the concurrency and hence resulting dwell time for the final assay.

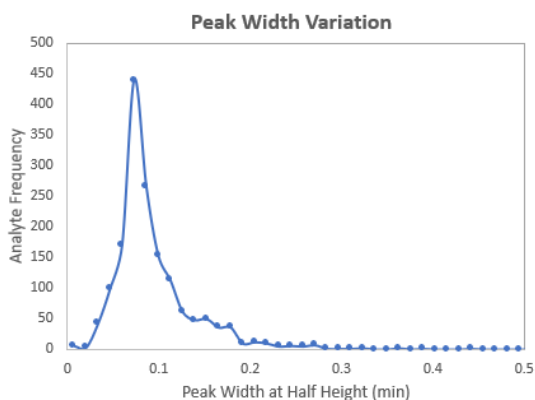


Figure 4. Optimization of Scheduled MRM Algorithm Assay using sMRM Pro Builder Template. To obtain better quality data, the assay can be optimized further using the advanced features in the Scheduled MRM Algorithm. Using the peak width and RT variance computed, an optimized MRM detection window can be included in assay for each lipid species. The types of enhancements improve the assay by maximizing dwell times while maintaining cycle time of the method.

Chromatographic Reproducibility

The LC-MRM reproducibility was evaluated by injecting the NIST plasma lipid extract (10 replicates) using the final Scheduled MRM Pro assay. Excellent reproducibility was observed with majority of lipid species showing retention time standard deviations below 0.05 minutes (Figure 5).

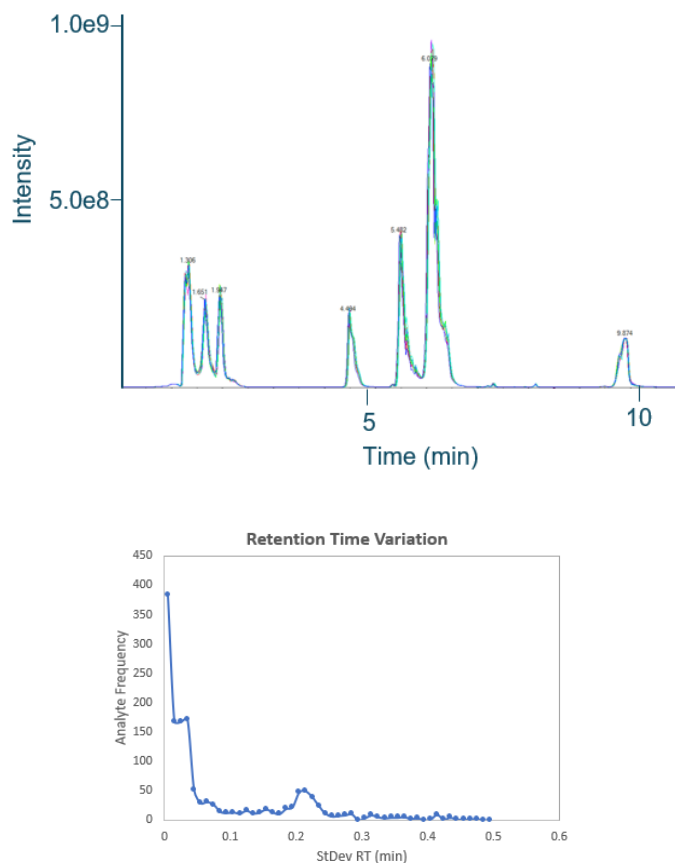


Figure 5. Reproducibility of the Chromatography for Plasma Replicates. (Top) Very reproducible chromatography was achieved with majority of lipid molecular species having retention deviations well below 0.05 minutes (bottom).

Quantitative Reproducibility

Using the optimized Scheduled MRM Algorithm acquisition method, 5 replicate injections were performed on plasma using the SCIEX Triple Quad™ 7500 System. Reproducibility of quantitation was assessed by computing the %CV of the peak area replicates for the lipids within each class. Figure 6 (middle) shows the cumulative fraction of lipids in selected classes at each % CV level for all lipids from the optimized method. Nine HCER lipids were measurable in plasma and HCER was the 5th most abundant class as measured by sum of peak areas (top) and showed highest reproducibility (middle). Other classes (SM, PC, CER, DCER, HCER and LPC) were also measured with very good reproducibility with 80% of lipids quantified with 20% CV or better. The TAGs are a large class (454 tested) but by filtering the assay to the 201 TAGs measurable in this sample, good reproducibility of this class was obtained (Figure 6 bottom).

Improved Results Quality with Scheduled MRM Pro Algorithm

Advanced method features like Retention Time Tolerance (Flexible Window Width) in Scheduled MRM Algorithm allow for additional method optimization to further fine tune an MRM method with many transitions such as this. Using the sMRM Pro Builder template and performing LC-MRM replicates, optimized values can be determined. sMRM Pro Builder template can also automatically reject low intensity peaks (uncertain detection) from final assay to further improve results quality by reducing MRM concurrency. Results obtained using the algorithm after filtering can be compared for 10 replicates of plasma matrix and filtering can be optimized.

Using Internal Standards for Quantitation

It is important to note that quantitation of lipids via global lipid profiling is challenging. In this method, several options are available to quantitate lipid levels in samples. For relative quantitation, a single lipid molecular species for each lipid class is needed. This strategy corrects for extraction efficiency as well as systematic errors; however, this strategy should only be used for comparative analysis between two samples. Using this strategy, it is not correct to report the concentrations of individual lipid molecular species, because the fragmentation efficiency of different molecular species within the same class is not equal and depends on the length and the number(s) of double bonds present in the fatty acid that fragments in the negative ion mode. Using this strategy, it is possible to conclude certain lipids increase or decrease when two different samples are compared. SPLASH mix from Avanti Polar Lipids is a commercially available mixture of lipid internal standards that can be used for this quantitation strategy.

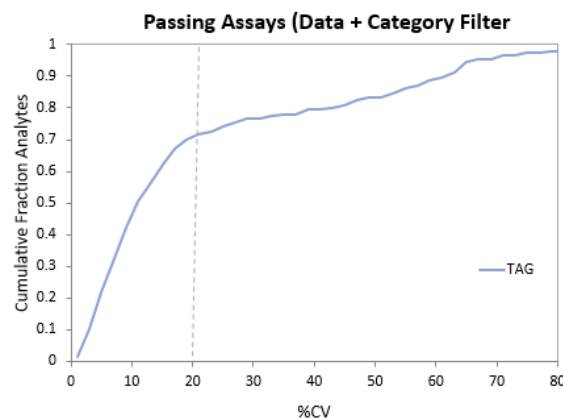
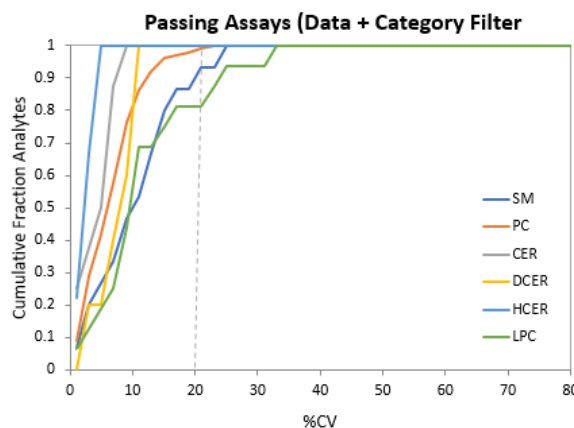
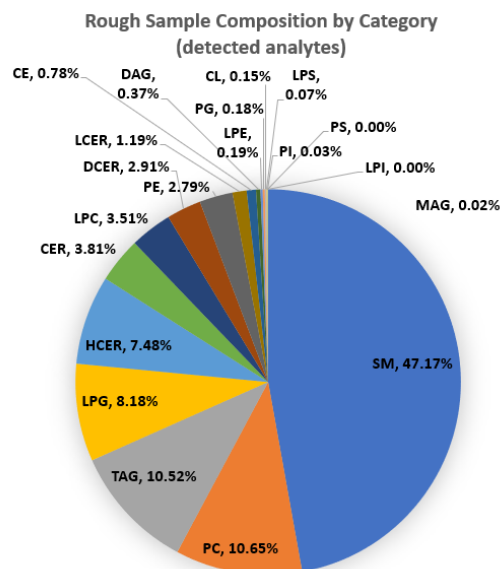


Figure 6. Increased Area Reproducibility of Assay using Scheduled MRM Pro Algorithm. Rough composition of lipids in plasma was estimated by summing the peak areas of detected lipids in each class from NIST plasma (top). Peak area %CV were computed across the replicate injections and plotted for selected lipid classes, plotted as cumulative %CV plots (middle). During assay development, the assay was filtered for just the detectable lipid species in the sample based on test data. The TAG class is a very large class; by filtering out the undetectable TAGs, good data reproducibility were observed (bottom).

A second strategy involves using multiple labeled internal standards to enable accurate quantitation (quantitative bias <10%); absolute quantitation requires a labeled internal standard for every analyte and is not possible in these experiments. Accurate quantitation can be achieved by assigning a structurally similar (but not necessarily identical) internal standard to each analyte. Recently, a comprehensive mixture of labeled internal standards was developed for the Lipidyzer™ Platform. Using these standards, accurate quantitation can be achieved for many but not all of the lipid classes targeted in this method (SM / DAG / CE / CER / TAG / LPE / LPC / PC / PE / FFA).

The method reported here is flexible to either internal standard strategy through addition of the chosen internal standards to the sample and inclusion in the MRM method development.

Summary

In this study, a targeted global lipid profiling strategy is presented that enables a broad array of different lipids to be quantified at the molecular species level (~1900 molecular species). An optimized chromatography method was developed to minimize isobaric interference through the chromatographic separation of lipid class, which allows for a relatively rapid and specific lipid screening technique. The target list of lipids is comprehensive, covering most major lipid classes and categories, and MRMs were selected to cover lipids containing fatty acids with 12-26 carbons and 0-6 double bonds including odd chain lipids. The method is customizable, so new lipid categories, classes and molecular species can be added.

Results quality of the method has been demonstrated in plasma matrix to highlight both the robustness and the flexibility of the LC-MRM assay.

An easy to use tool (sMRM Pro Builder template) was also developed to streamline the optimization of the targeted MRM assay to maximize results quality when performing assay with very large numbers of time scheduled MRM transitions.

References

1. Han X, Yang K, Gross RW. Multi-Dimensional Mass Spectrometry-Based Shotgun Lipidomics And Novel Strategies For Lipidomic Analyses. [Mass Spectrometry Reviews. 2012; 31\(1\): 134-178.](#)
2. Bielow C, Mastrobuoni G, Orioli M, Kempa S. On Mass Ambiguities in High-Resolution Shotgun Lipidomics. [Anal Chem. 2017; 89: 2986-2994.](#)
3. Ovčáčíková M, Lísa M, Cífková E, Holčápek M. Retention behavior of lipids in reversed-phase ultrahigh-performance liquid chromatography-electrospray ionization mass spectrometry. [J. Chromatogr A. 2016; 10\(1450\), 76-85.](#)
4. Koelmel JP, Kroeger NM, Gill EL, Ulmer CZ, Bowden JA, Patterson RE, Yost RA, Garrett TJ. Expanding Lipidome Coverage Using LC-MS/MS Data-Dependent Acquisition with Automated Exclusion List Generation. [J Am Soc Mass Spectrom. 2017. 28\(5\). 908-917.](#)
5. Hines KM, Herron J, Xu L., Assessment of Altered Lipid Homeostasis by HILIC-Ion Mobility-Mass Spectrometry-Based Lipidomics. [J Lipid Res. 2017. 58\(4\), 809-819.](#)
6. Novel Chemical Standards Kits Enable Facile Lipid Quantitation. [SCIEX Technical note RUO-MKT-02-3879-A.](#)
7. [Download method information.](#)
8. [Download the sMRM Pro Builder Template.](#)
9. [Download sMRM Concurrency Calculator](#) for use with Analyst Software.
10. Using Scheduled MRM™ Algorithm in SCIEX OS Software. [SCIEX community post RUO-MKT-18-11941-A.](#)

The SCIEX clinical diagnostic portfolio is For In Vitro Diagnostic Use. Rx Only. Product(s) not available in all countries. For information on availability, please contact your local sales representative or refer to <https://sciex.com/diagnostics>. All other products are For Research Use Only. Not for use in Diagnostic Procedures.

Trademarks and/or registered trademarks mentioned herein, including associated logos, are the property of AB Sciex Pte. Ltd. or their respective owners in the United States and/or certain other countries.

© 2020 DH Tech. Dev. Pte. Ltd. RUO-MKT-02-8482-B. AB SCIEX™ is being used under license.