Trouble-free Food Testing

Technologies to improve uptime and productivity

Chemistries & Supplies

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Overview

- Needs in Food Testing Today
 - Considerations and Tips for LC and LCMS analysis
 - A Quick Look at Sample Prep Options
- Important Method Development Fundamentals
- A Food Analysis Workflow using LC/MS
 - Botanical insecticide in fish
- Tools to Improve Your Food Analyses



Where is Food Testing Today?

- Increasing complexity in food sourcing and supply
 - Globalization of the food supply has made it necessary for detailed analysis of nutritional value, composition, and contaminants
 - Concerns over food safety and the effects on human health have increased regulation and requirements associated with food testing
 - Older methodologies for food testing were time consuming and not amenable to modern needs for higher throughput food analyses
- Samples come in all types
- Appropriate food analysis cannot be achieved without the proper LC and sample prep methodology



Chromatographic Techniques for Food testing

- Gas Chromatography
 - Typically used for:
 - Non-polar, volatile, oils, etc
 - Can require derivatization of compounds



- Typically used for:
- Polar, thermally labile
- No need for derivatization of many compounds
- Often has less time consuming sample prep







Challenges in Food Testing

- New method development
- Finding ways to cut costs
- Doing more with less increasing lab productivity
- Keeping up with the latest food safety changes





LC Method Development on Food Testing

What's different in LC analysis for food analysis?

- Sample matrix complexity
- Multiple residues of interest
- Often very polar analytes in the sample of interest

Modern column technologies make food testing faster and easier

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New Column Technologies for Method Development

Columns for high resolution and high speed analysis

- Sub-2 µm columns for ultra-high pressure operation
- Sub-3 µm superficially porous columns
- Considerations when developing methods on new column technologies
- Particle size (< 3 µm)
- Column pressure limits >400 bar (600-1200 bar typical)
- Other factors remain same as for legacy, 5 µm columns

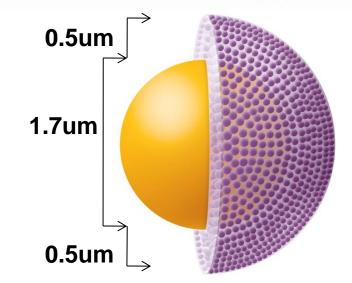




New Superficially Porous Column Technologies

Poroshell 120 columns:

- Efficiency ≈ 90% of sub-2 µm
- Pressure ≈ 40-50% of sub-2 µm
- N \approx 2X 3.5 μ m (totally porous)
- d_p = 2.7µm
- 2 µm frit to reduce clogging
- P_{limit} = 600 bar for HPLC or UHPLC
- Particles
 - 1.7 µm solid core
 - 0.5 µm diffusion path
 - 2.7 µm total diameter







Comparing Efficiency and Pressure with Different Types of Columns

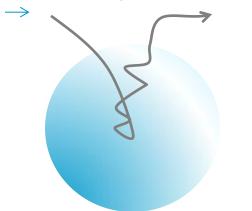
Particle Size/Type	Pressure	Efficiency	LC Compatibility
5µm Totally Porous	80 bar	5,000	All 400 bar instruments
3.5µm Totally Porous	123 bar	7,800	All 400 bar instruments
2.7µm Poroshell 120	180 bar	12,000	All LCs/UHPLCs (up to 600 bar)
1.8µm Totally Porous	285 bar	12,500	All LCs/UHPLCs (up to 1200 bar)

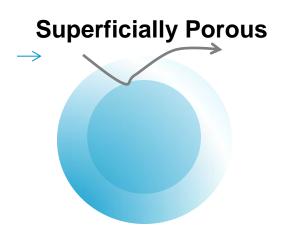
Columns: 4.6 x 50mm, Mobile Phase: 60% ACN:40% Water Flow Rate: 2 mL/min



Analyte Mass Transfer Improvements through Lower Diffusion

Totally Porous





The Measure of Confidence

- Totally porous particles
 - diffusion throughout particle
- Poroshell 120
 - diffusion limited to outer shell

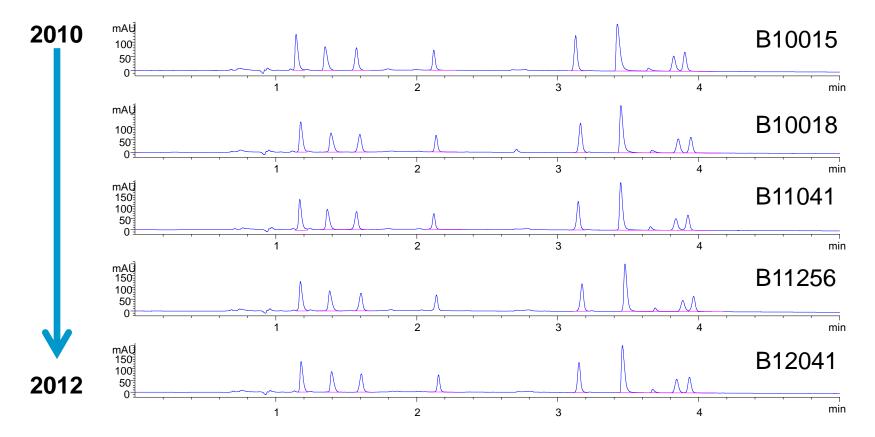
van Deemter equation: $h = A + B / v + C \cdot v$

- Results:
 - Lower C term
 - Higher efficiency
- And
 - Higher flow rate with
 - Minimal impact on efficiency



Other Considerations when Selecting a Column

 Robustness and batch-to-batch reproducibility of Poroshell 120 columns



Beverage Additives



General Steps to a robust LC method for Food Analysis

- Choose the type of chromatography often Reversed Phase (C18, polar embedded, etc...) or HILIC phase
- Choose the appropriate starting and ending gradient conditions
- Adjust pH to shift analyte retention
- Change the bonded phase to adjust for proper peak spacing and resolution





Change in Retention with pH for Ionizable Compounds is Key to Method Development

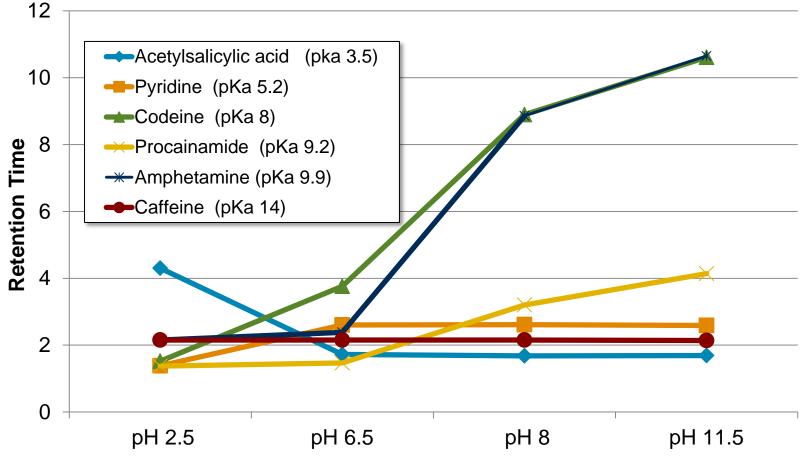
- Non-charged analytes have better retention (i.e. acids at low pH and bases at high pH)
- Silanols on silica ionize at mid-pH, increasing retention of basic analytes (i.e. possible ion-exchange interactions)
- Choose mobile phase pH to optimize retention and selectivity during method development
- Poroshell 120 EC-C18 can be used over a wide pH range
- Other choices exist for high pH





Change in Retention with pH for Ionizable Compounds is Compound-Dependent

More retention for non-charged analytes (i.e. acids at low pH and bases at high pH)



Mobile Phase: 45% MeOH, 55% 20 mM Phosphate Buffer

Why Change the Bonded Phase?

- Different interactions for polar and non-polar compounds.
- Exploit other interactions with bonded phase (e.g., pi-pi)
- These all change with bonded phase!
- Changing the bonded phase can improve selectivity/resolution, reduce analysis time
- When you use Poroshell 120 columns the comparison of bonded phases can be done quickly!
- Easy with multiple column choices plus high speed technologies





Poroshell 120 Column Chemistries

Poroshell 120 EC-C18 and C8

 Robust endcapped C18 for best peak shape at pH 2-9

Poroshell 120 StableBond C18 and C8

• Robust chemistries for pH<2

Poroshell 120 Phenyl-Hexyl

- Same Eclipse Plus bonding process as ZORBAX Eclipse Plus Phenyl-Hexyl
- Excellent choice for pi-pi interactions
- Alternative selectivity to EC-C18 or SB-C18
- Selectivity similar to phenyl, diphenyl, or other phenyl-hexyl columns

Poroshell 120 SB-Aq

 Proprietary bonding phase is an excellent choice for polar analytes

Poroshell 120 Bonus-RP

 Embedded polar group provides unique selectivity for polar compounds

Poroshell 120 EC-CN

 Flexible endcapped CN chemistry with Normal and Reversed Phase character

Poroshell 120 HILIC

 Bare silica HILIC for use in Hydrophilic interaction chromatography of polar molecules

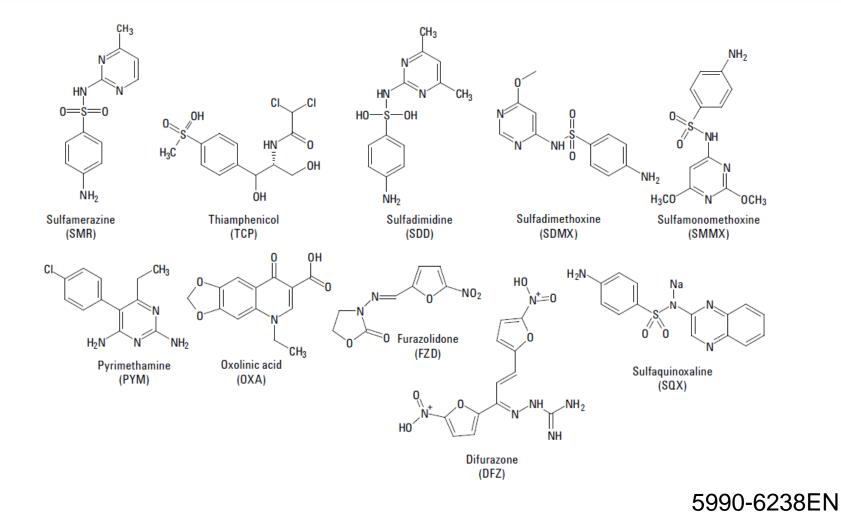


Considerations for LCMS Method Development

- Mobile Phase Selection
 - Gradients often preferred due to the number of analytes and differences in retention characteristics
- Buffer Choice
 - Choose a volatile buffer (formic acid, acetic acid, etc)
- Column dimensions for higher sensitivity and throughput
 - Often choose narrow columns for higher sensitivity and shorter columns for faster analysis

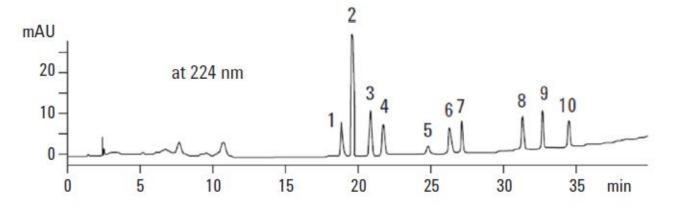






The Measure of Confidence





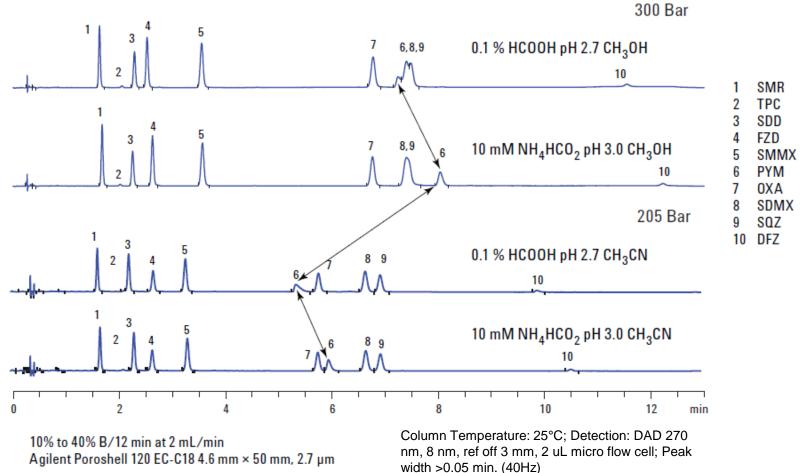
1	SMR	6	SMMX
2	PYM	7	DFZ
3	TCP	8	SDMX
4	SDD	9	SOX
5	FZD	10	OXA

Instrument:	Agilent 1100 Series HPLC
Column:	250 mm × 4 mm id, RP-18 Purospher, 5 μm, p/n 79925PU-584
Mobile phase:	$A = 0.7\%$ Phosphoric acid, $B = CH_3CN$
Gradient:	0.0 min 5% B; 10.0 min 5% B; 40.0 min 65% B; 45.0 min 65% B; Post Time 7.0 min 5% B
Flow rate:	1.0 mL/ min
Temperature:	40 °C
Injection volume:	20 µL

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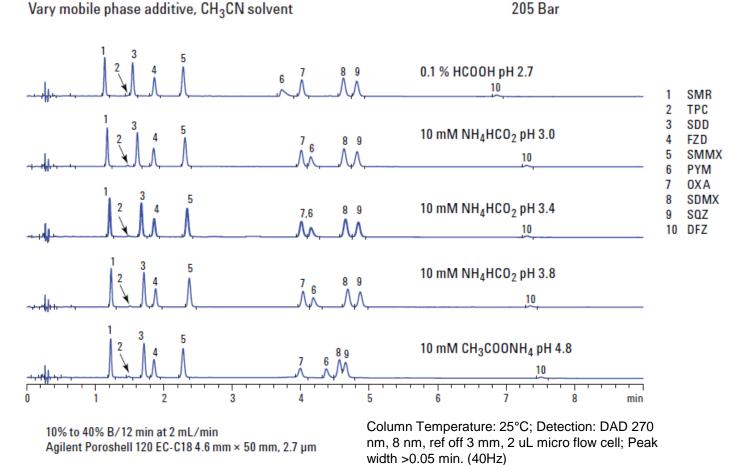




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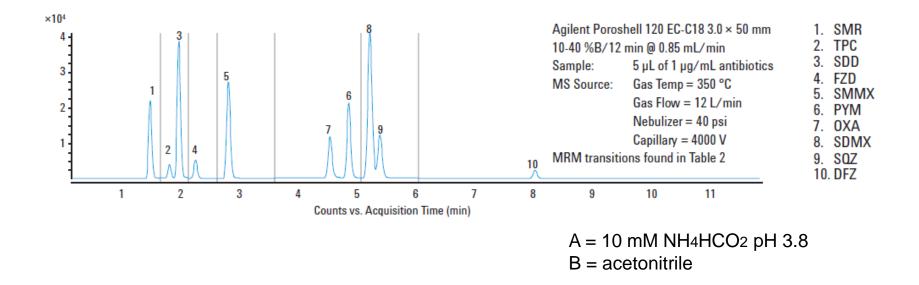




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•Conditions were scaled for a 3.0 x 50 mm column

•Shows that 3.0 mm can easily be used for conventional UV and MS detection

5990-6238EN





In the Food Analysis Workflow, what trips you up the most?

- Selecting the 'right' sample prep method for my food sample
- Sample preparation takes too long
- Column selection / refining methods
- Columns clog up







Sample Prep in Food Analysis



QuEChERS

•Easy-to-use

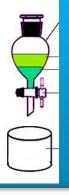
•Ideal for food samples



Liquid-liquid extraction

Phase separation

•Often time consuming



Filtration

Particulate removal

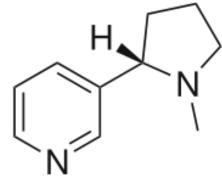
Lipid removal







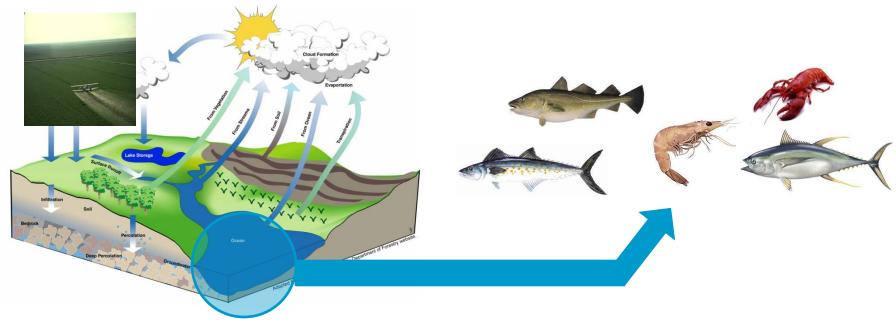
- Nicotine is used as a botanical insecticide in the US and Canada
- Banned as an insecticide in EU and will be banned in the US beginning 2014
- Oily liquid at room temperature
- Water miscible







- How nicotine end up in fish or other aquatic organisms?
 - Insecticides used on the farm \rightarrow Rain \rightarrow River \rightarrow Ocean







• How to prepare real fish sample?

QuEChERS

- If the sample matrix is solid, especially food, consider QuEChERS first.
- QuEChERS is the most common sample preparation method used in many food testing labs.
- <u>Qu</u>ick, <u>Easy</u>, <u>Ch</u>eap, <u>Effective</u>, <u>Rugged</u>, <u>Safe</u>.



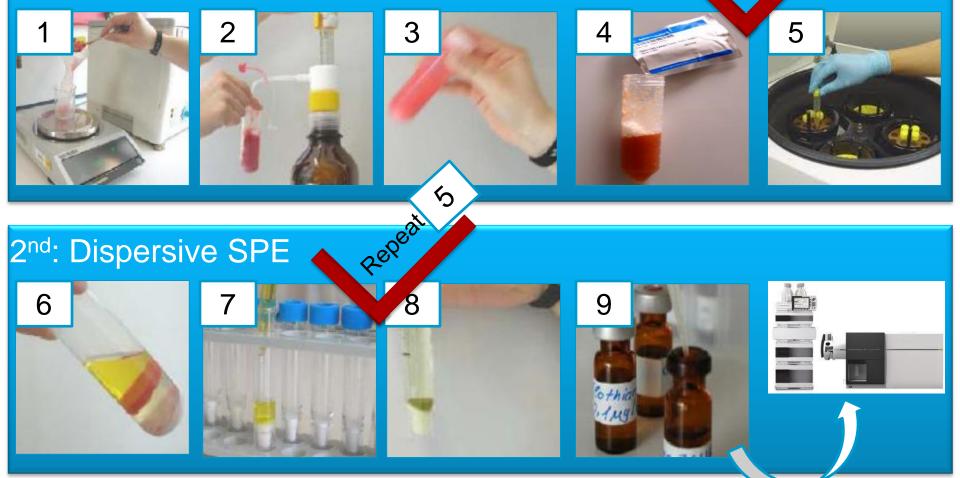


- Benefits of QuEChERS in nicotine analysis in fish
 - QuEChERS is a combination of two steps
 - 1st extraction, 2nd dispersive SPE
 - Little or no method development is required
 - 1st extraction: Simply choose AOAC, EN, or original method
 - 2nd dispersive SPE: Choose one of our dispersive packets
 - Customizable to your needs





1st: Extraction





Confidentiality Label September 26, 2013

• Which column to choose for analysis of nicotine? \rightarrow HILIC

Benefits of HILIC over Reversed Phase					
	HILIC	Reversed Phase			
Retention Time	Nicotine & metabolites will retain and separate	Most of nicotine and its metabolites will elute too early			
Sample Solvent	No evaporation & reconstitution	•Sample solvent switch or •Dilution is <i>REQUIRED</i>			
Initial Mobile Phase	90% ACN	Extremely low organic mobile phase or 100% aqueous			
MS Sensitivity	High	Lower			
Backpressure	Low \rightarrow Wider flow rate range	High \rightarrow Limited flow rate rang.			





- LC-MS/MS Conditions
 - Column:
 - LC:
 - MS: Agilent 6460 with JetStream and ESI+

ACN

0.7 mL/min

- Mobile phase A:
- Mobile phase B:
- Flow rate:
- Gradient:

Time (min)	%B
0	90
4	70
4.5	70
4.6	90
6	90

Agilent Infinity 1290 UHPLC



Poroshell 120 HILIC 2.7 µm, 2.1 X 100 mm

10 mM ammonium formate (pH=3.0)

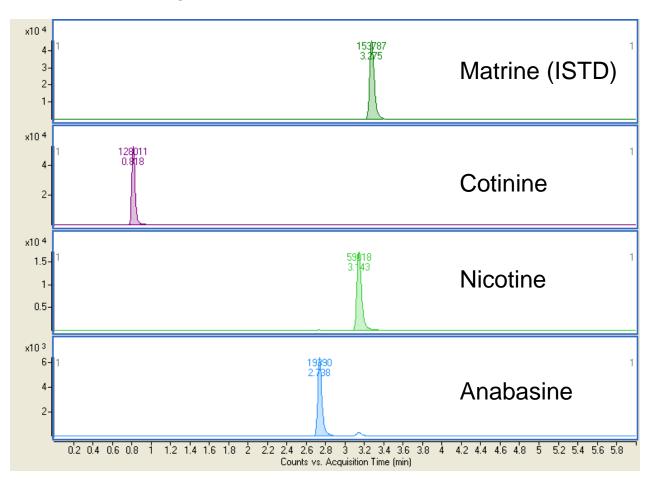
• Nicotine and its metabolites

	Nicotine	Cotinine	Anabasine		
	H N N	H ^N CH ₃ H			
log P	1.20	0.07	1.25		
рКа	8.02	8.8	11.0		
MRM	163.1→132.1	177.1→80.1	163.1→118.1		
Collision E	82	112	92		





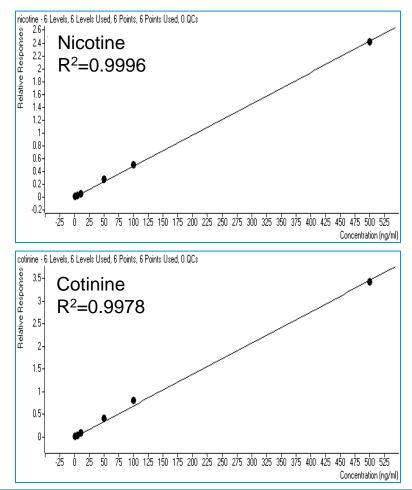
• Results – MS Chromatogram

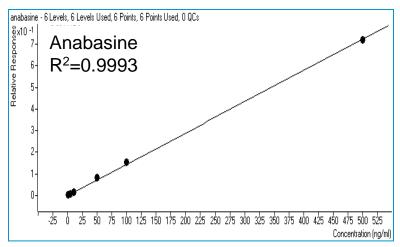






• Results - Calibration curves & detection limits





Calibration range 1 – 500 ng/g fish.

Great linearity.

LOD: 1 ng/g

LOQ: 5 ng/g





Results – Precision & accuracy (n=8)

	Nicotine		Anabasine			Cotinine			
	Low (10 ng/g)	Mid (100 ng/g)	High (500 ng/g)	Low (10 ng/g)	Mid (100 ng/g)	High (500 ng/g)	Low (10 ng/g)	Mid (100 ng/g)	High (500 ng/g)
Average	11.4	97.7	450.2	9.2	86.4	389.1	12.8	117.0	466.9
Recovery	113.7 %	97.7 %	90.0 %	91.8 %	86.4 %	77.8 %	127.9 %	117.0 %	93.4 %
%RSD	6.4 %	1.6 %	2.1 %	6.2 %	1.6 %	2.5 %	5.4 %	2.2 %	2.1 %



Where can we make improvements?

- Instrument maintenance tips
 - Add a guard column, in-line filter to protect your system.
 - Filter samples \rightarrow Guarantee removal of particulates
 - **Avoid** high salt concentration in HILIC

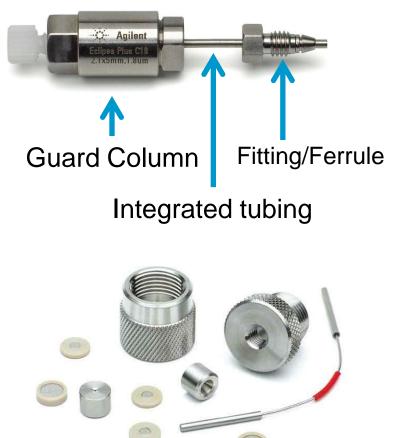
Start with 10 mM ammonium formate (pH=3.0)





Use of Guard Columns and Inline Filters

- Poroshell 120 Fast Guard for UHPLC
- Inline filters and guard columns extend the life of HPLC columns by preventing particulates and impurities from clogging and potentially irreversibly sticking to the analytical column
- Column lifetime is extended
- \$\$\$ savings from fewer analytical columns purchased
- Minimal, if any, impact to the chromatography

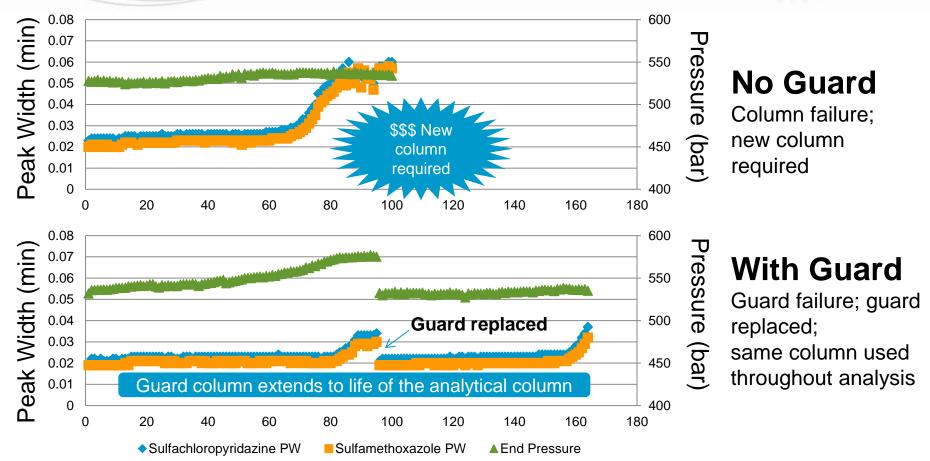


RRLC In-line filter



Benefits of Installing a Fast Guard for UHPLC

Method: Accelerated Lifetime Test - Similac sample (milk substitute diluted 300:1) containing 2 sulfa drugs; Peak width change indicating column failure



By installing a guard column when using dirtier samples, one can extend the life of their column, and utilize more inexpensive guard columns rather than column replacements



Summary—Getting to your trouble-free solution

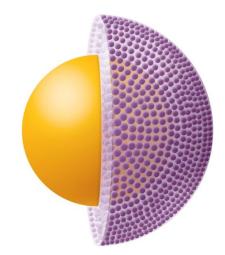
- Find the best tools for you
 - Choices include traditional SPE, liquid-liquid extraction or time & cost saving newer methods such as QuEChERS
 - QuEChERS is ideal for MRL (maximum residue level) detections for many food samples and traditional SPE is good for lower level detections when needed
 - Sample filtration and guard columns to protect your LC system





Benefits of Poroshell 120 Column technology

- Increased separation speeds (reduce your analysis time)
- Use with any instrument (HPLC or UHPLC)
- Reduced solvent & waste costs
- Poroshell 120
 - Scalability
 - Ease of method transfer
 - Similar phases to totally porous ZORBAX columns







ATool for Method Development

The Column and Sample Prep NAVIGATOR:

http://www.agilent.com/chem/navigator



Making Column and Sample Prep choices easier!





Agilent is Here to Help

See www.agilent.com/chem/cstechsupport

