

Waters

THE SCIENCE OF WHAT'S POSSIBLE.™

UPLC® Method Development and Validation



- Methods are developed throughout the drug development process
 - Samples vary in complexity
 - Redundancy across organization
- Method development is costly and time-consuming
 - Desire to streamline processes to bring products to market faster
 - Faster chromatographic methods will improve profitability

UPLC Technology Can Streamline Method Development

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UPLC Technology enables faster method development

Methods can be developed in a single work day!

- Systematic scouting protocol involving pH, organic and column chemistry
- High-resolution sub-2 μm column technology enables high resolution separations, faster
- Automated column and mobile phase selection



- ➔ ■ Approaches Towards Method Development
- Selectivity and Retention Tools
 - Stationary Phase and Particle Substrate
 - Solvent
 - pH
- Method Development Strategy
- Applications
- Conclusions

- Literature search, colleague, speculation?
- Stepwise iterative procedure
 - Next step experimental design based on results from previous experiment
- Systematic scouting
 - Evaluate pH, organic solvent and stationary phase
 - Select best combination of those three
 - Fine tuning/optimization (temperature, gradient slope)

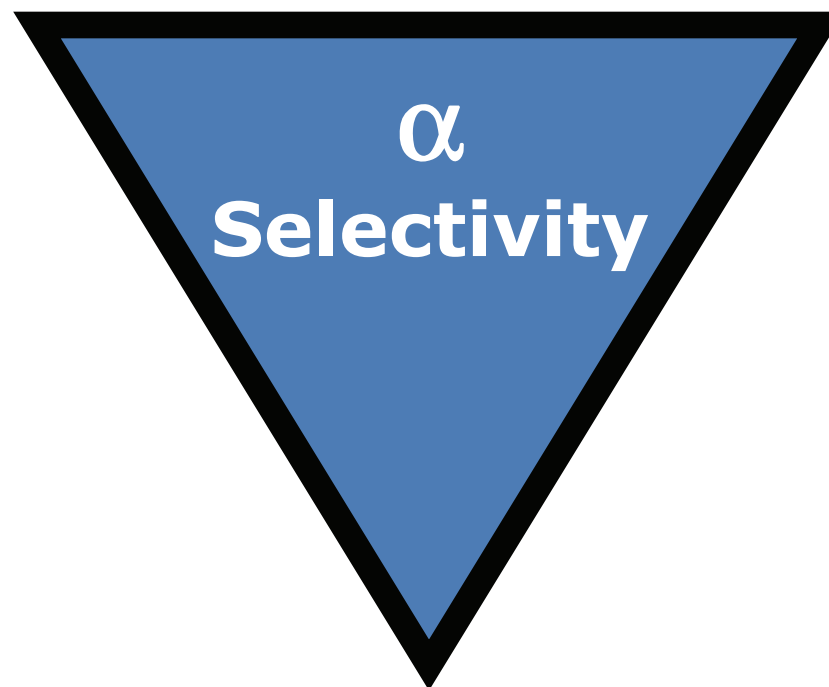
Desirable Information for Method Development

- Sample solubility
- Number of analytes
 - How many peaks of interest are you trying to separate?
- Chemical structure(s)
- Functional groups; how the analytes differ
 - Ionizable species? How will pH influence chromatography?
- Detection
 - Type of detection that is required or possible?
- Concentration range and quantitative requirements
- Sample matrix effects

- Approaches Towards Method Development
- ■ Selectivity and Retention Tools
 - Stationary Phase and Particle Substrate
 - Solvent
 - pH
- Method Development Strategy
- Applications
- Conclusions

Solvent

pH



Column Chemistry

- Ligand
- Base particle

Improving Resolution with Complementary Selectivities

$$R_s = \frac{\sqrt{N}}{4} \cdot \frac{\alpha - 1}{\alpha} \cdot \frac{k}{k + 1}$$

Physical

Chemical

Maximized in UPLC Separations by:

- Ultra-low dispersion system
- Small (< 2 μm) particles
- Higher pressure capability
- Well-designed columns

Maximized in UPLC Separations by:

- Range of chemistries
- Multiple particle substrates
- Wide usable pH range (BEH)
- Higher retentivity (HSS)

Impact on Resolution

Double **N**

Double **k**

Double **α**

% Improvement

20-40%

15-20%

> 400%

Different Ligands: Different Selectivity

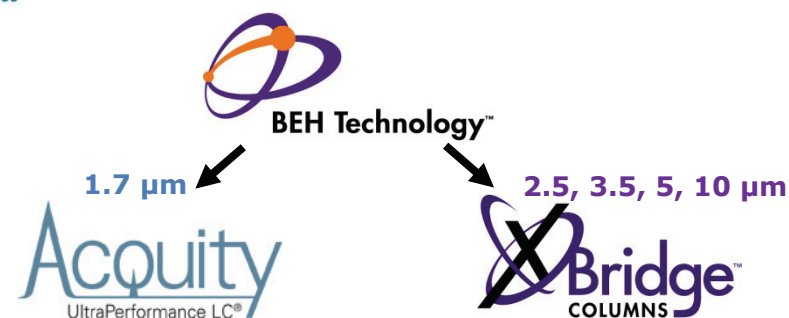
- Changes in hydrophobicity
 - Longer alkyl chain will provide greater retention
- Changes in silanol activity
 - Affect peak asymmetry and influences secondary interactions
- Changes in hydrolytic stability
 - Longer column lifetimes with greater number of ligand attachment points to the particle surface
- Changes in ligand density
 - Influences sample loadability

BEH Chemistries of UPLC Technology

- **BEH C₁₈**
 - Trifunctionally Bonded C₁₈
 - First UPLC® column choice
 - Superior peak shape & efficiencies
- **BEH C₈**
 - Trifunctionally Bonded C₈
 - Wide pH range
- **BEH Shield RP18**
 - Monofunctionally bonded
 - Embedded carbamate group
 - Alternate selectivities
- **BEH Phenyl**
 - Trifunctionally Bonded C₆ (Hexyl) Phenyl
 - Unique combination of chemistry & particle
 - Wide pH range
- **BEH HILIC**
 - Unbonded, rugged BEH particle
 - HILIC for very polar bases

| | BEH Particle | | | | |
|-----------------|-------------------------------|------------------------------|-------------------------------------|-------------------------------------|-------|
| | C ₁₈ | C ₈ | Shield RP18 | Phenyl | HILIC |
| Chemistry | | | | | |
| Ligand Type | Trifunctional C ₁₈ | Trifunctional C ₈ | Monofunctional Embedded Polar Group | Trifunctional C ₆ Phenyl | — |
| Ligand Density* | 3.1 μmol/m ² | 3.2 μmol/m ² | 3.3 μmol/m ² | 3.0 μmol/m ² | — |
| Carbon Load* | 18% | 13% | 17% | 15% | — |
| Endcap Style | Proprietary | Proprietary | TMS | Proprietary | — |
| pH Range | 1-12 | 1-12 | 2-11 | 1-12 | 1-8 |

*Expected or Approximate Values

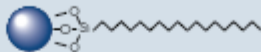


Ease of Migration from HPLC to UPLC

Simplified Purification and Isolation

HSS Chemistries of UPLC Technology

- **HSS T3**
 - T3: Polar compound retention
 - Aqueous-compatible C₁₈ chemistry
 - Designed for maximum retentivity
- **HSS C₁₈**
 - High coverage, trifunctionally bonded C₁₈ chemistry
 - Universal, high performance C₁₈ chemistry
 - Proprietary endcapping for superior peak shape
 - Silica particle performance
- **HSS C₁₈ SB**
 - SB: **S**electivity for **B**ases
 - Non-endcapped: optimum silanophilic selectivities

| | HSS Particle | | |
|-----------------|-------------------------------------------------------------------------------------|-------------------------------|-------------------------------|
| | C ₁₈ | C ₁₈ SB | T3 |
| Chemistry |  | | |
| Ligand Type | Trifunctional C ₁₈ | Trifunctional C ₁₈ | Trifunctional C ₁₈ |
| Ligand Density* | 3.2 μmol/m ² | 1.6 μmol/m ² | 1.6 μmol/m ² |
| Carbon Load* | 15% | 8% | 11% |
| Endcap Style | Proprietary | None | Proprietary |
| pH Range | 1-8 | 2-8 | 2-8 |

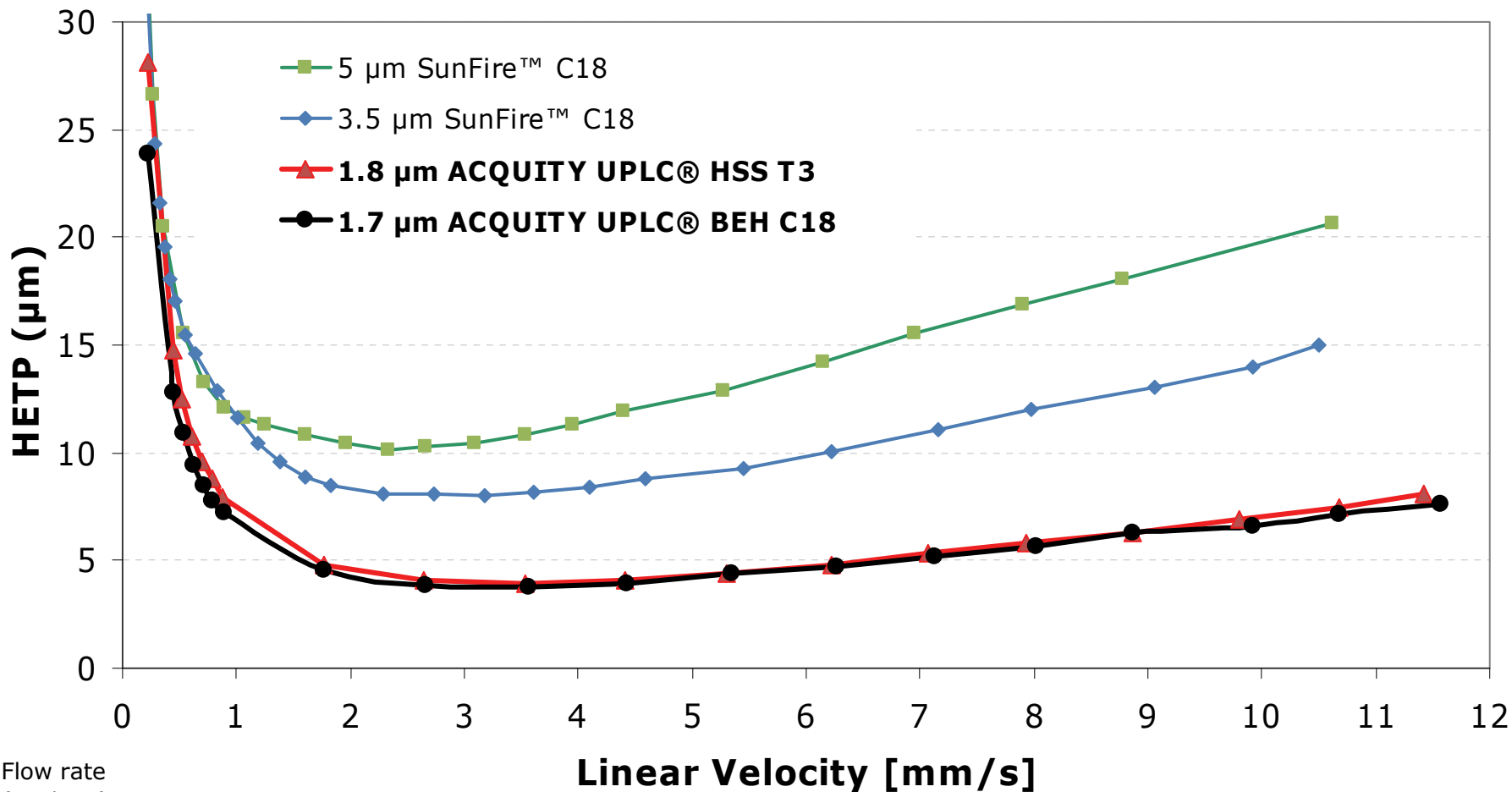
Challenges of Developing and Manufacturing UPLC Particles

- Developing UPLC particles requires:
 - R&D scientists capable of synthesizing, sizing and evaluating prototype sub-2 μm chromatographic media
 - Cutting-edge synthesis facilities capable of reliably and reproducibly producing commercial quantities of bulk materials

- UPLC particle requirements:
 - Pressure tolerance
 - Proper morphology
 - High efficiency/mass transfer
 - Advanced bonding and endcapping processes

Additional requirements as compared to HPLC particles

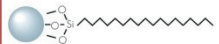
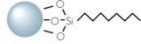

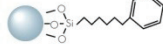

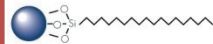
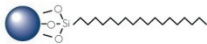
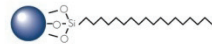
Two Rugged, Efficient UPLC Particles: van Deemter Curves (<500 MW)



Flow rate
(mL/min)

| | | | | | | | | | | | | |
|------------------|------|------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| 1.0 mm ID | 0.04 | 0.07 | 0.11 | 0.14 | 0.18 | 0.21 | 0.25 | 0.28 | 0.32 | 0.35 | 0.39 | 0.42 |
| 2.1 mm ID | 0.15 | 0.30 | 0.45 | 0.60 | 0.75 | 0.90 | 1.05 | 1.20 | 1.35 | 1.50 | 1.65 | 1.80 |
| 4.6 mm ID | 0.70 | 1.40 | 2.10 | 2.80 | 3.50 | 4.20 | 4.90 | 5.60 | 6.30 | 7.00 | 7.70 | 8.40 |

UPLC Particles and Chemistries Summary

| | | | BEH Particle | | | HSS Particle | | |
|-----------------|------------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------|
| Chemistry | C ₁₈  | C ₈  | Shield RP18  | Phenyl  | HILIC  | T3  | C ₁₈  | C ₁₈ SB  |
| Ligand Type | Trifunctional C ₁₈ | Trifunctional C ₈ | Monofunctional Embedded Polar Group | Trifunctional C ₆ Phenyl | — | Trifunctional C ₁₈ | Trifunctional C ₁₈ | Trifunctional C ₁₈ |
| Ligand Density* | 3.1 μmol/m ² | 3.2 μmol/m ² | 3.3 μmol/m ² | 3.0 μmol/m ² | — | 1.6 μmol/m ² | 3.2 μmol/m ² | 1.6 μmol/m ² |
| Carbon Load* | 18% | 13% | 17% | 15% | — | 11% | 15% | 8% |
| Endcap Style | Proprietary | Proprietary | TMS | Proprietary | — | Proprietary | Proprietary | None |
| pH Range | 1-12 | 1-12 | 2-11 | 1-12 | 1-8 | 2-8 | 1-8 | 2-8 |

* Expected or Approximate Values

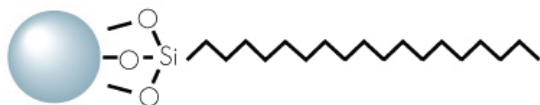
| Launch Date | Mar 2004 | Mar 2005 | Mar 2005 | Mar 2005 | Dec 2005 | Sep 2006 | Jun 2007 | Jan 2008 |
|-------------|----------|----------|----------|----------|----------|----------|----------|----------|
|-------------|----------|----------|----------|----------|----------|----------|----------|----------|



- Used in following method development protocol

ACQUITY UPLC Column Selection: Method Development Approach

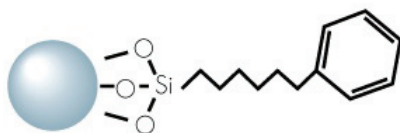
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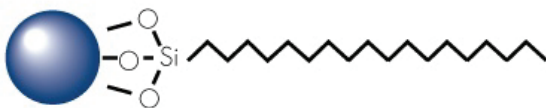
- **BEH C₁₈**
 - Widest pH range for maximum selectivity



- **BEH Shield RP18**
 - Embedded polar group offers complementary selectivity vs. alkyl C₁₈ chemistries



- **BEH Phenyl**
 - C₆ (Hexyl) Phenyl chemistry offers complementary selectivity to C₁₈ and Shield RP18 chemistries, especially for analytes with aromatic rings



- **HSS T3**
 - Polar compound retention and aqueous mobile phase compatibility

Selectivity Scouting Protocol

2.1 x 50 mm, <2 μm

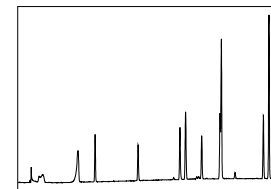
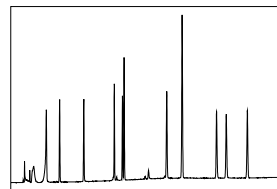
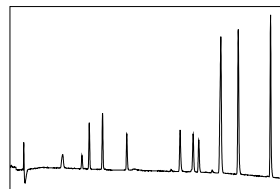
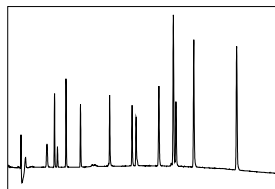
pH 3, ACN

pH 3, MeOH

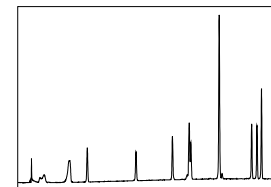
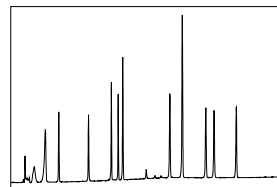
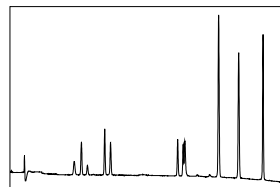
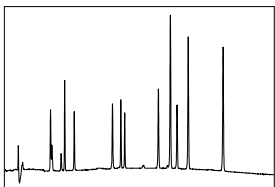
pH 10, ACN

pH 10, MeOH

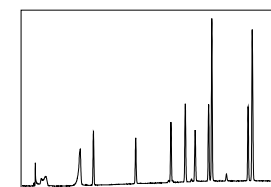
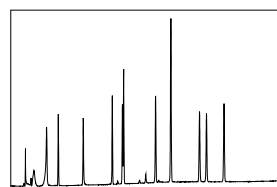
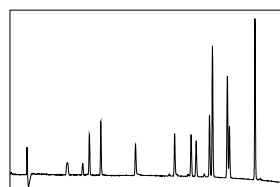
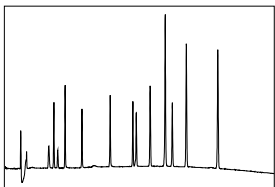
**ACQUITY UPLC
BEH C₁₈**



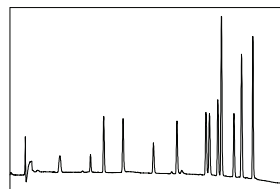
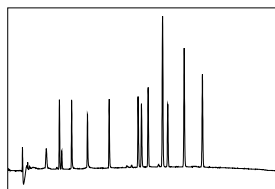
**ACQUITY UPLC
BEH Shield RP₁₈**



**ACQUITY UPLC
BEH Phenyl**



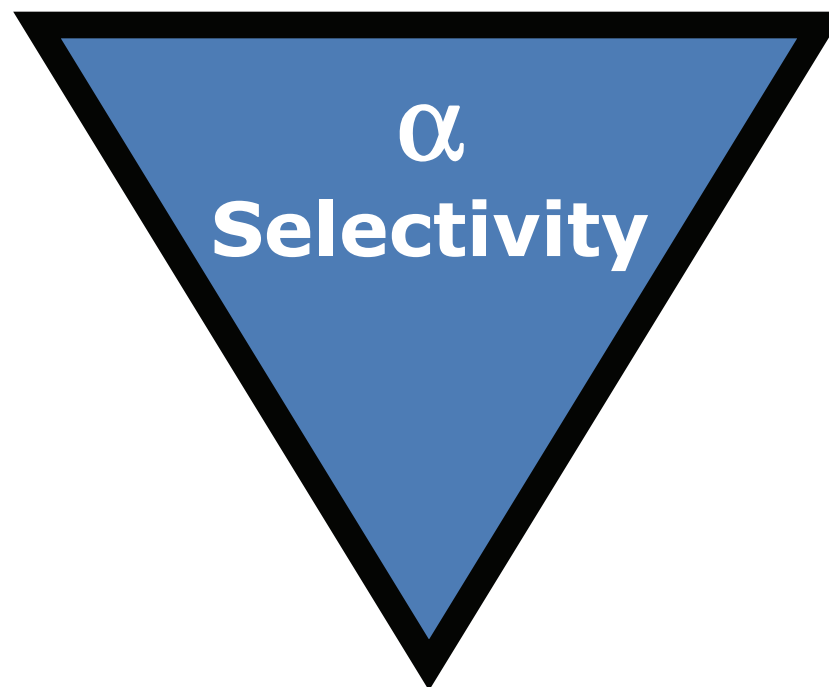
**ACQUITY UPLC
HSS T3**



Optimization

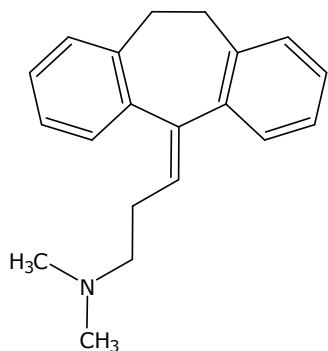
Solvent

pH

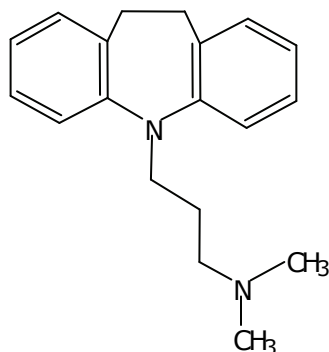


Column Chemistry

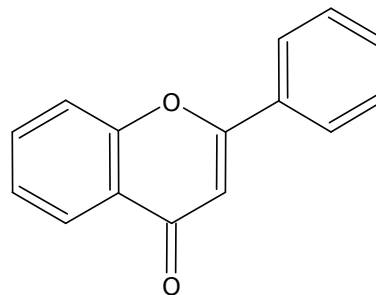
Selectivity Observations: Chemical Structures



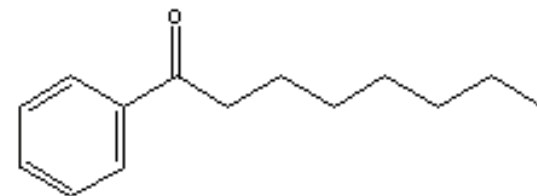
Amitriptyline (B)
m.w. 277.40



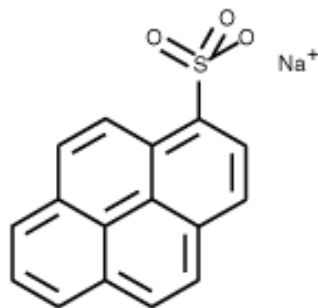
Imipramine (B)
m.w. 280.40



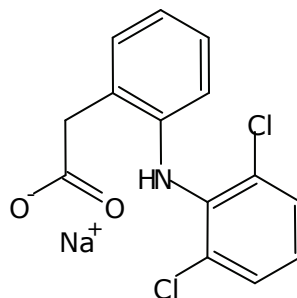
Flavone (N)
m.w. 222.24



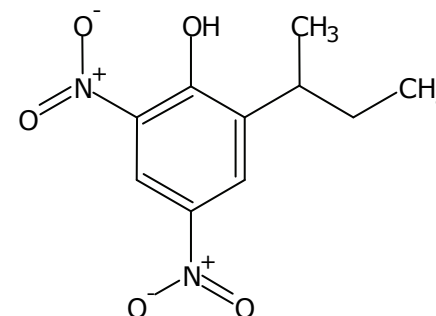
Octanophenone (N)
m.w. 204.31



1-Pyrenesulfonic acid (A)
m.w. 304.3

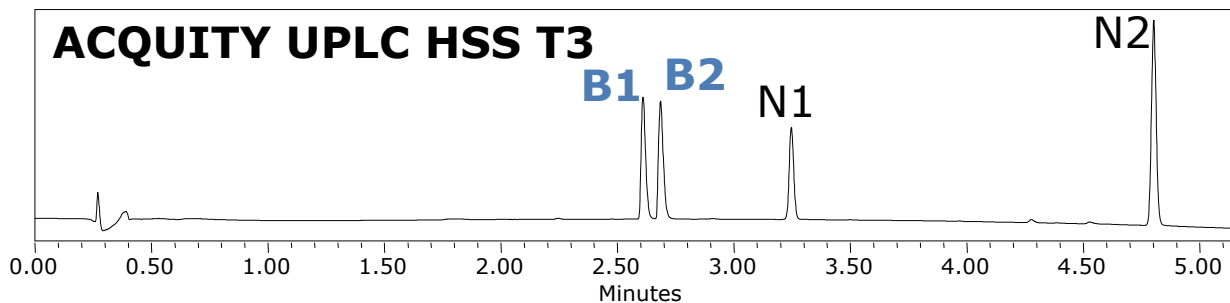
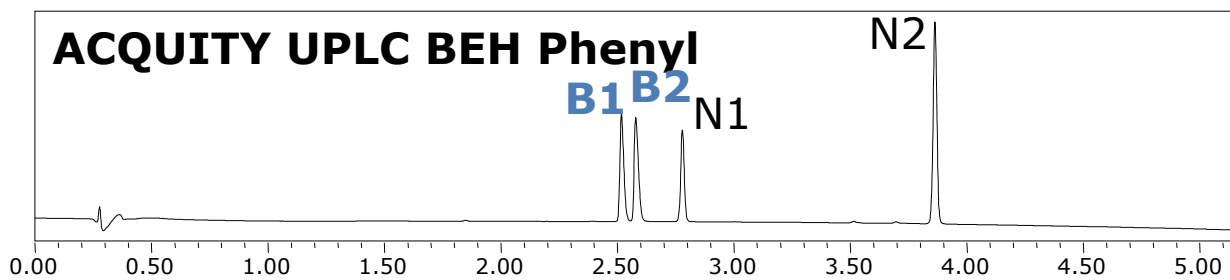
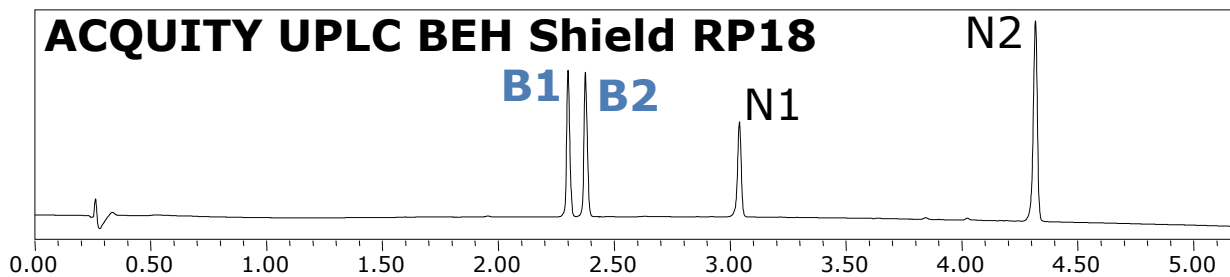
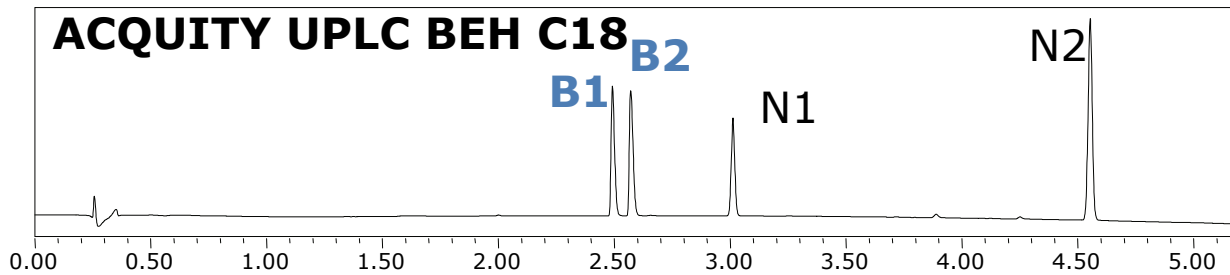


diclofenac (WA)
m.w. 318.13



Dinoseb (A)
m.w. 240.21

Stationary Phase Selectivity: Basic and Neutral Compounds



Acetonitrile pH 3.0

Test Probes:

- B1** imipramine
- B2** amitriptyline
- N1** flavone
- N2** octanophenone

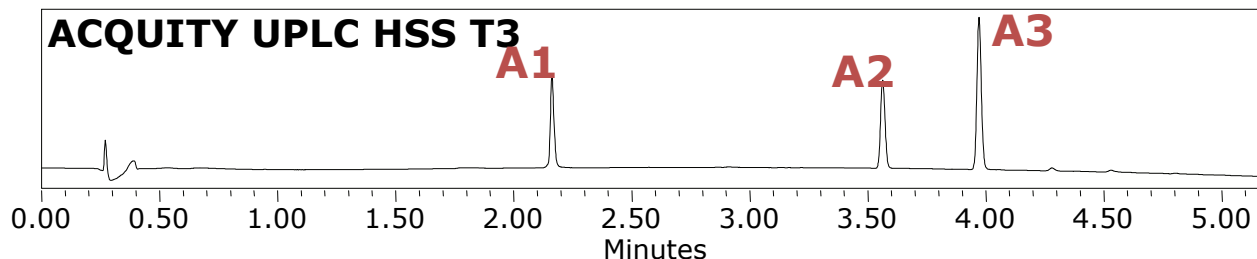
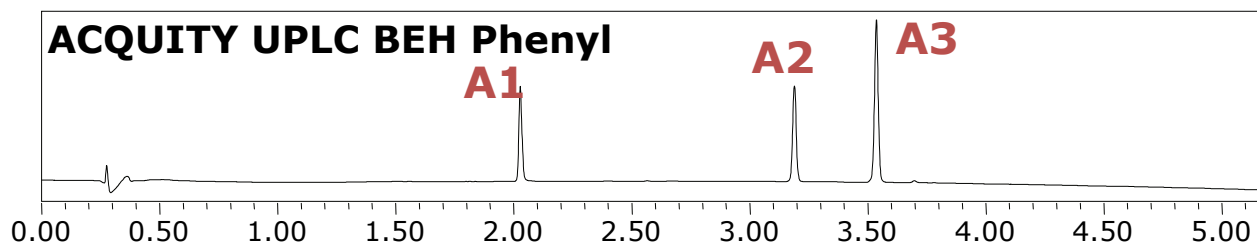
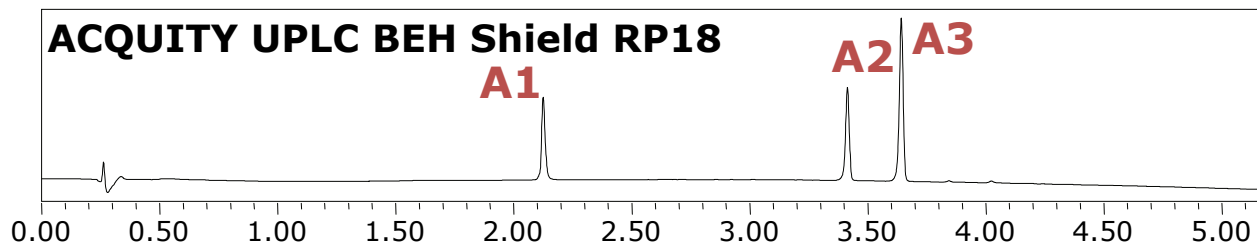
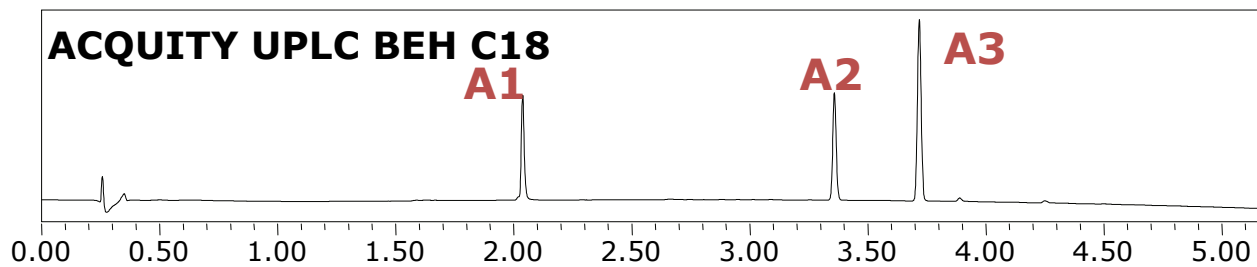
Small differences
in stationary
phase selectivity
with acetonitrile,
low pH

Stationary Phase Selectivity: Acidic Compounds

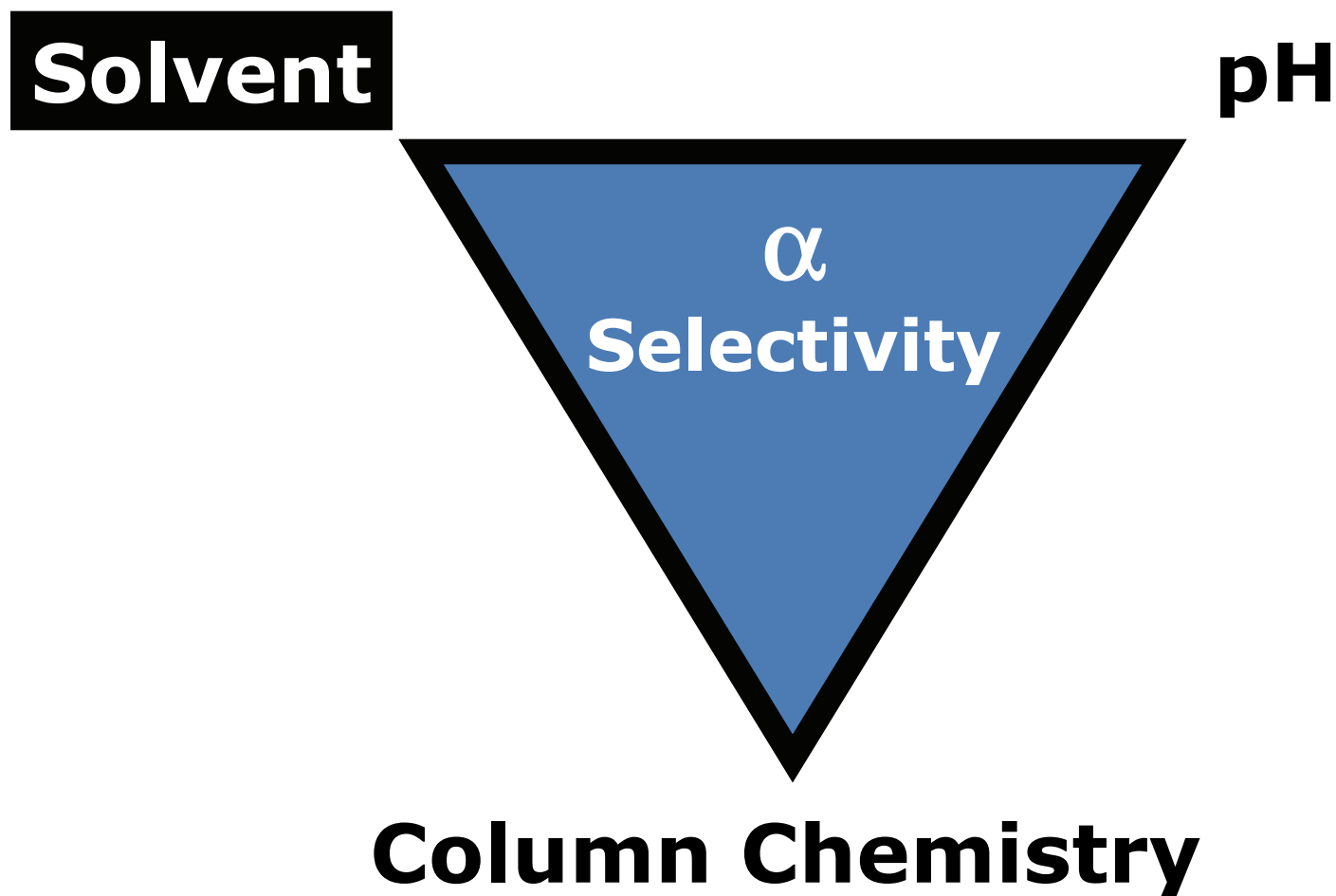
Acetonitrile pH 3.0

Test Probes:

- A1** 1-pyrenesulfonic acid
- A2** diclofenac
- A3** dinoseb



Small differences
in stationary
phase selectivity
with acetonitrile,
low pH

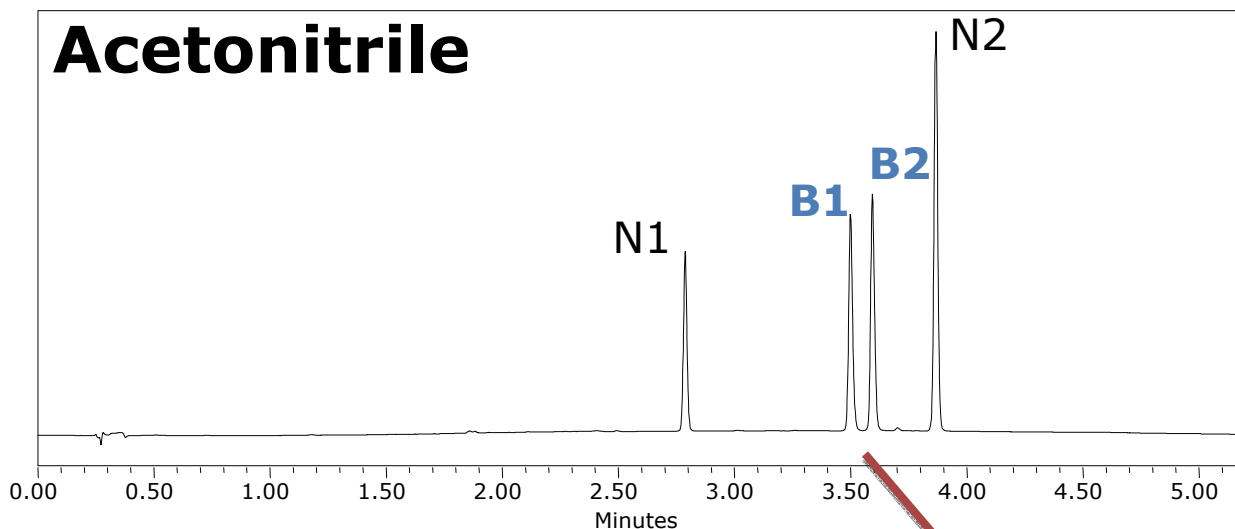


- Methanol
 - Weaker eluent
 - H-bonding solvent
- Acetonitrile
 - Aprotic solvent
 - Stronger eluent
 - Lower viscosity

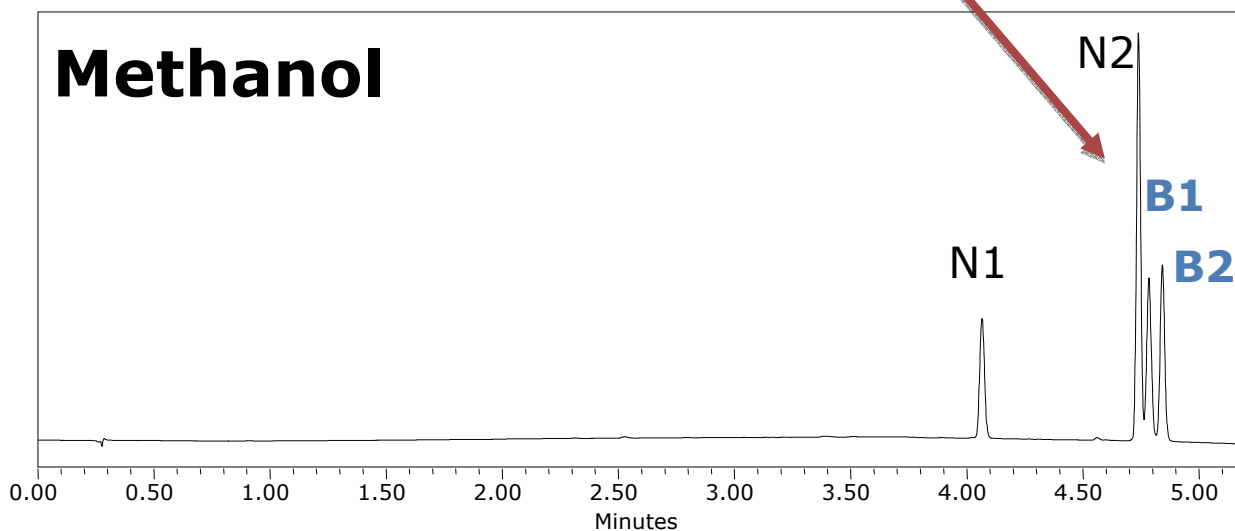
Solvent Selectivity: Basic and Neutral Compounds

ACQUITY UPLC BEH Phenyl

Acetonitrile



Methanol



Test Probes:

B1 imipramine

B2 amitriptyline

N1 flavone

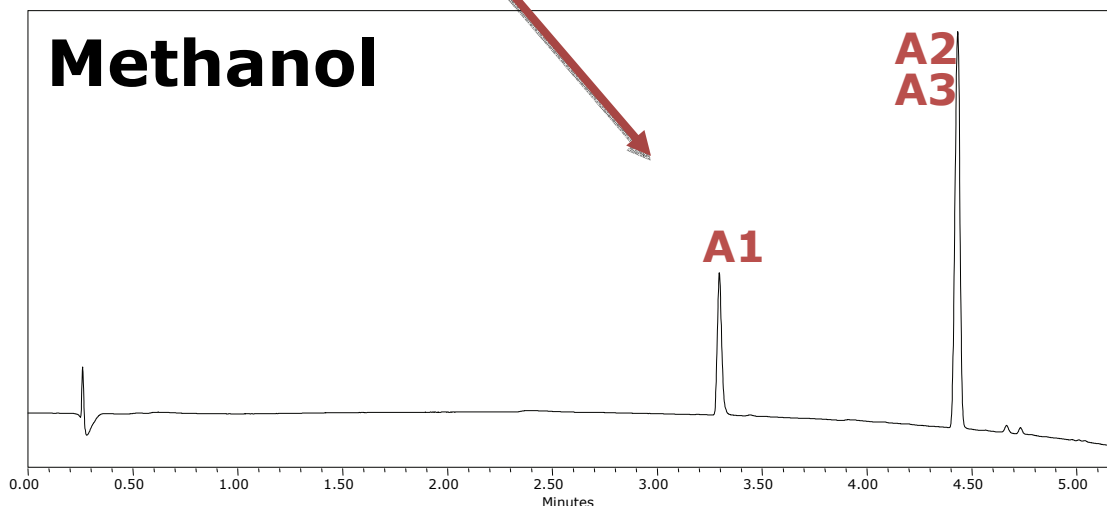
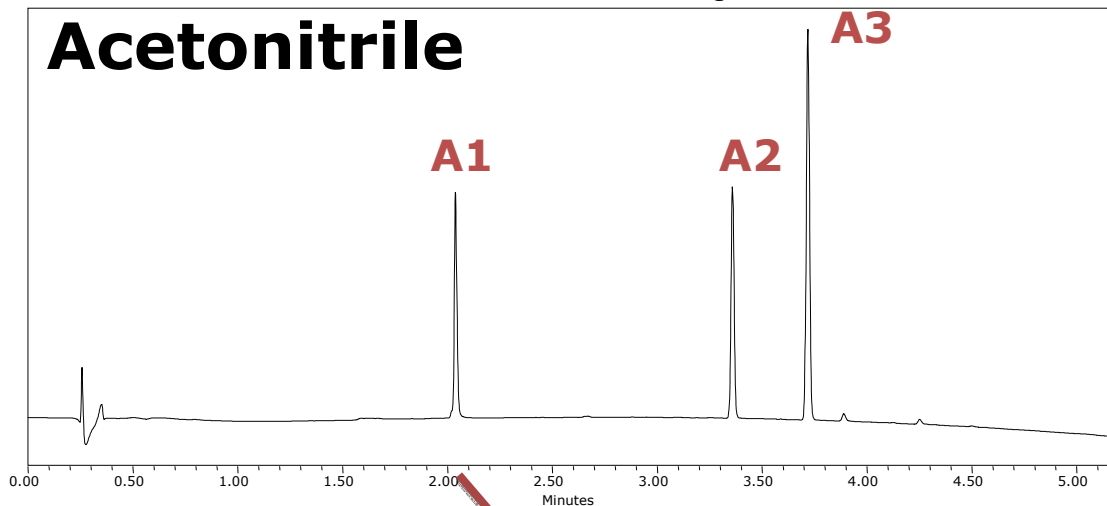
N2 octanophenone

Methanol is weaker elution solvent, resulting in greater retention for all analytes

Greater retention of bases respective to neutral probes

Solvent Selectivity: Acidic Compounds

ACQUITY UPLC BEH C18



Test Probes:

A1 1-pyrenesulfonic acid

A2 diclofenac

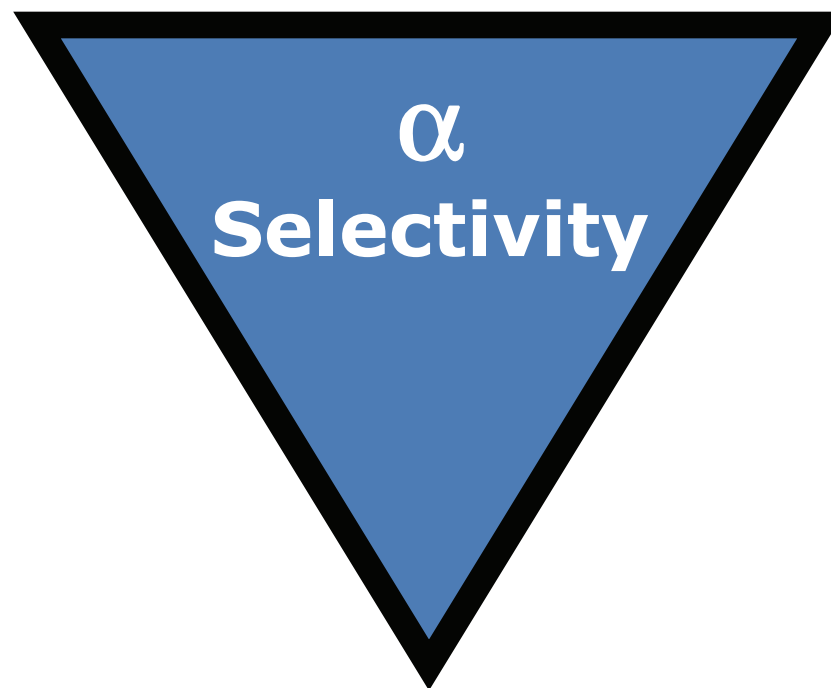
A3 dinoseb

Methanol is weaker elution solvent resulting in greater retention for all analytes

Elution order change

Solvent

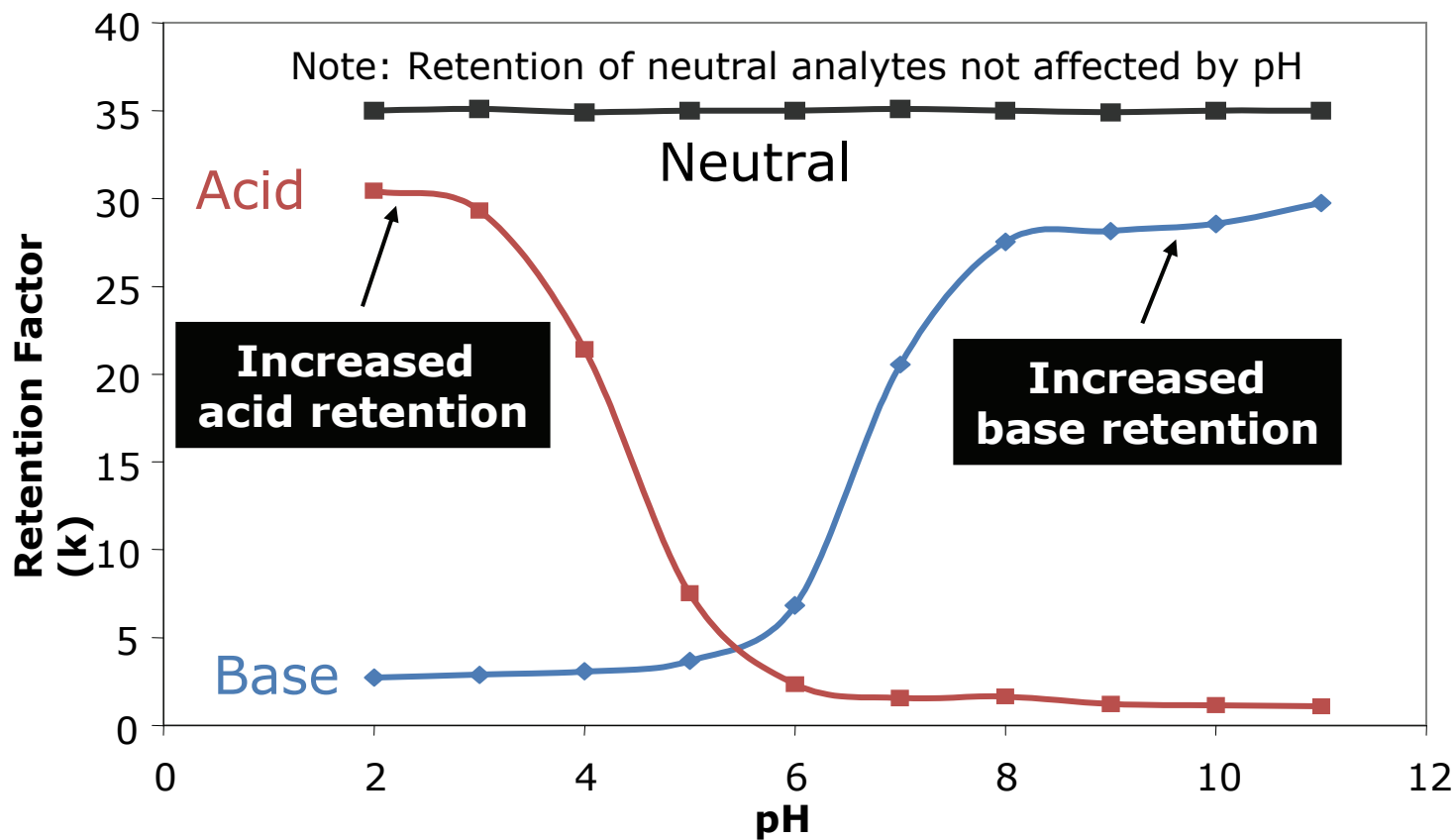
pH



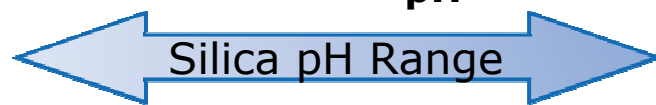
Column Chemistry

- Affects only analytes with ionizable functional groups
 - Amines
 - Carboxylic acids
 - Phenols
- Some compounds contain one or more ionizable function
- Strongest selectivity effects can be caused by pH changes

Reversed-Phase Retention Map: The Importance of Mobile Phase pH



HSS



BEH



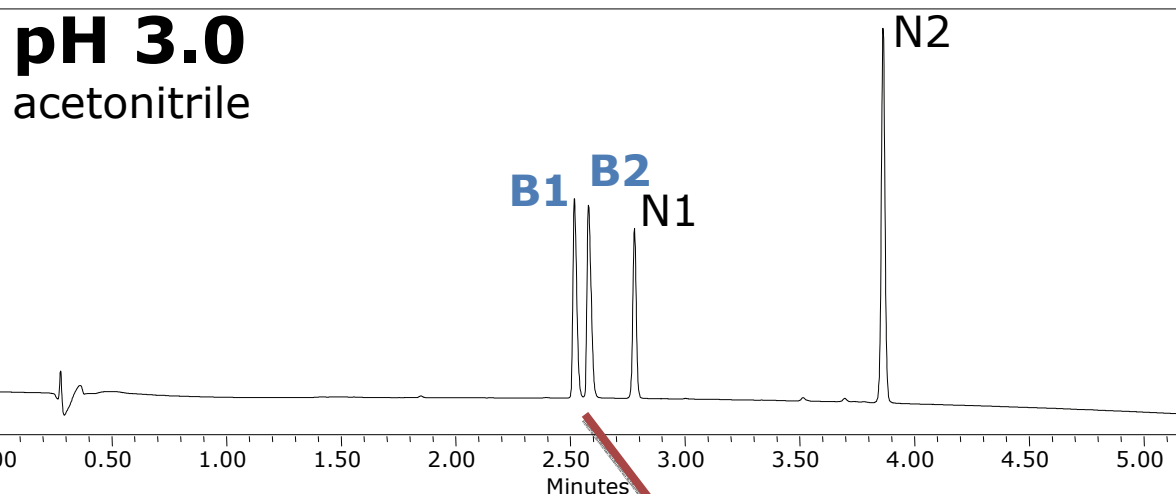
pH Selectivity: Basic and Neutral Compounds

ACQUITY UPLC BEH Phenyl

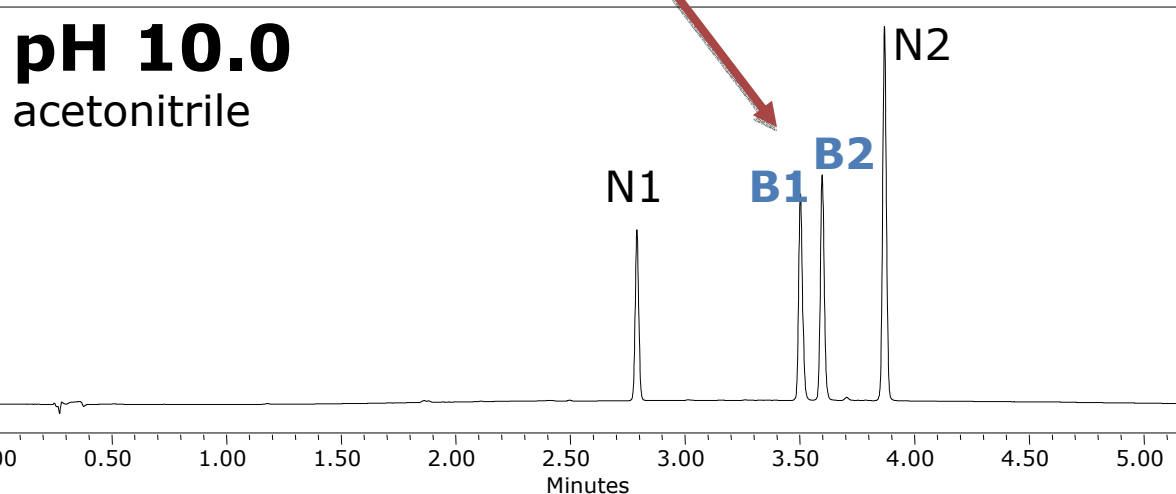
Test Probes:

- B1** imipramine
- B2** amitriptyline
- N1** flavone
- N2** octanophenone

pH 3.0
acetonitrile



pH 10.0
acetonitrile

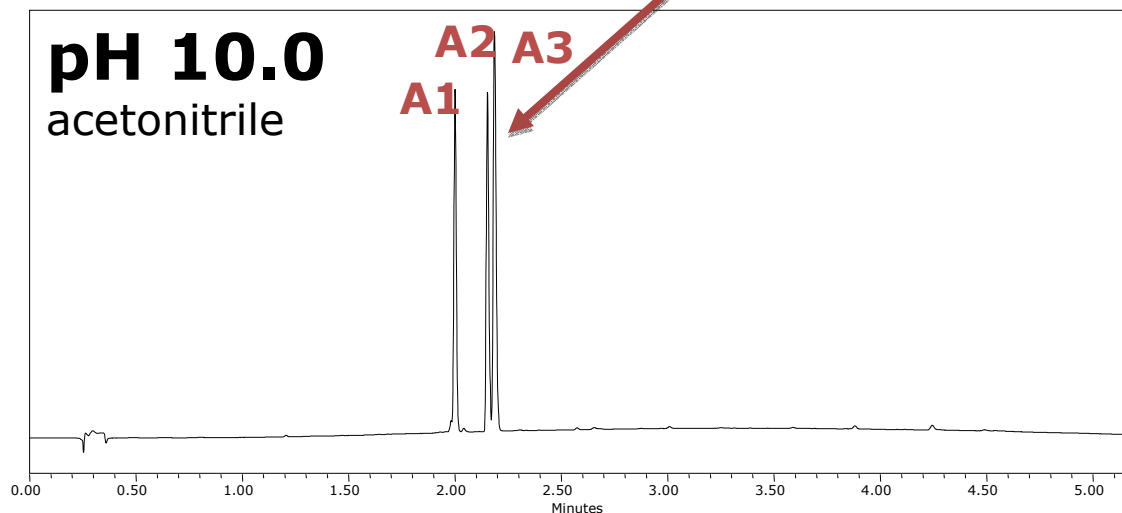
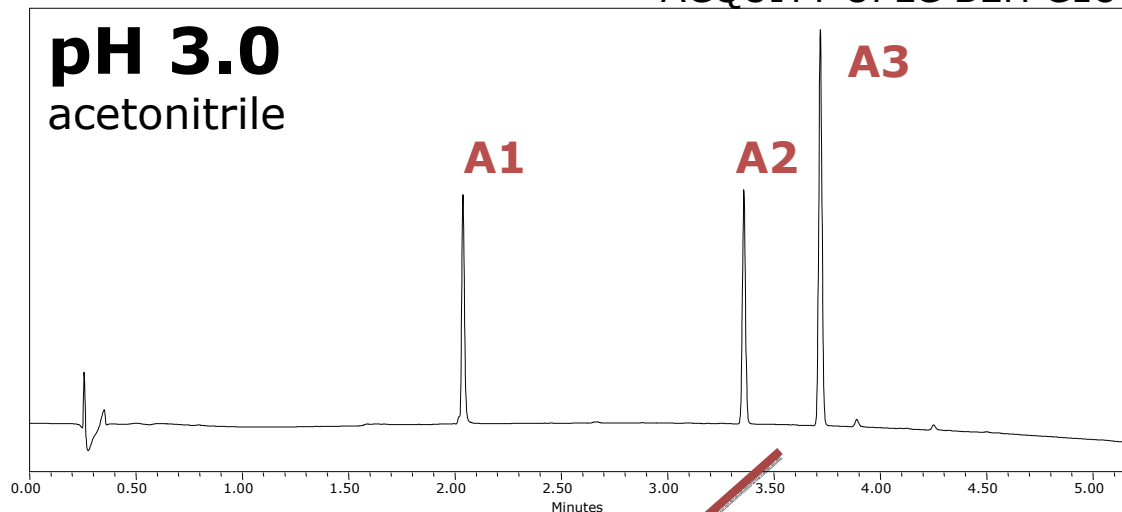


Neutrals
unaffected by pH

At alkaline pH,
bases are in their
un-ionized form,
resulting in
greater retention

pH Selectivity: Acidic Compounds

ACQUITY UPLC BEH C18



Test Probes:

- A1** 1-pyrenesulfonic acid
- A2** diclofenac
- A3** dinoseb

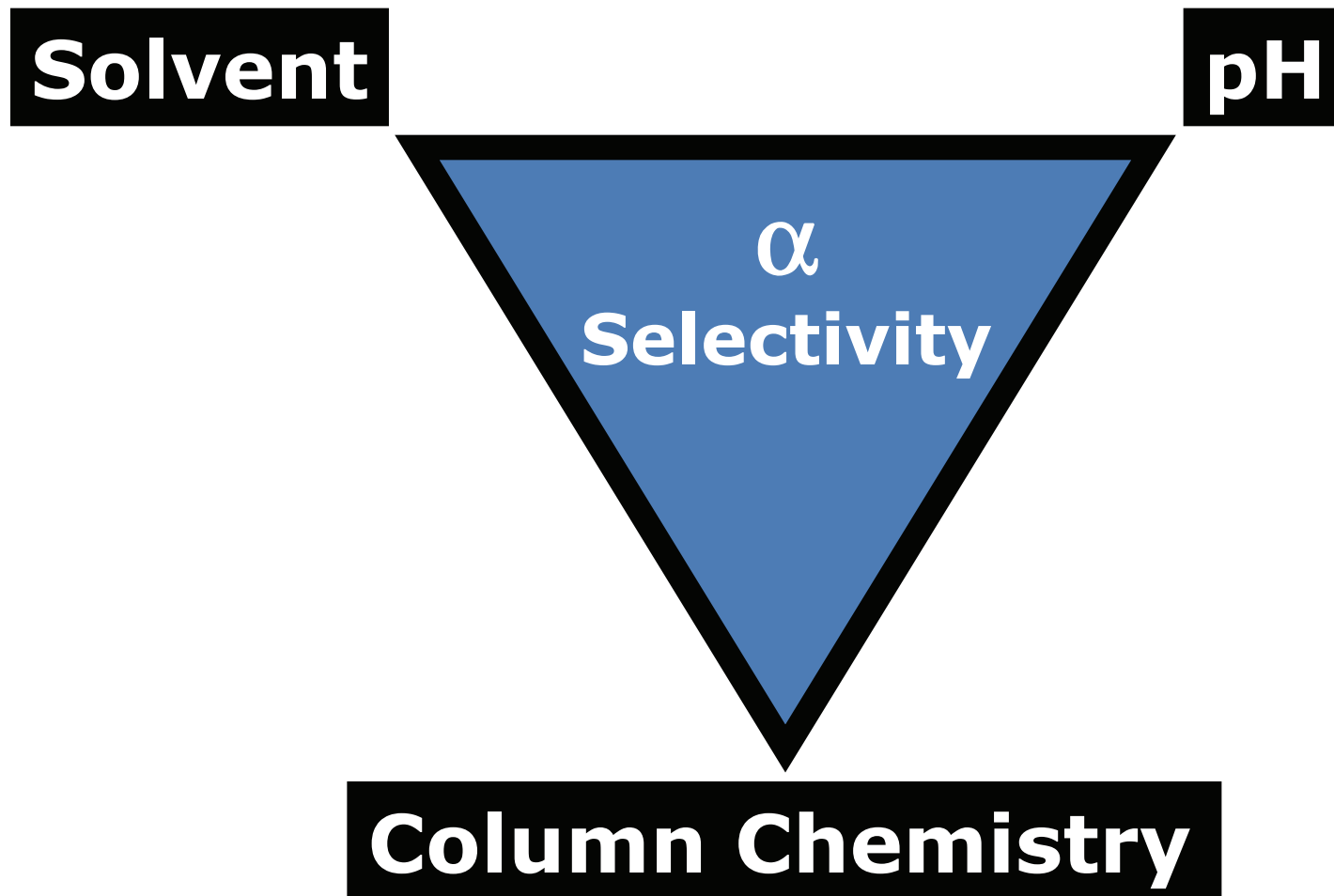
At acidic pH, acids are in their un-ionized form resulting in greater retention

Same elution order

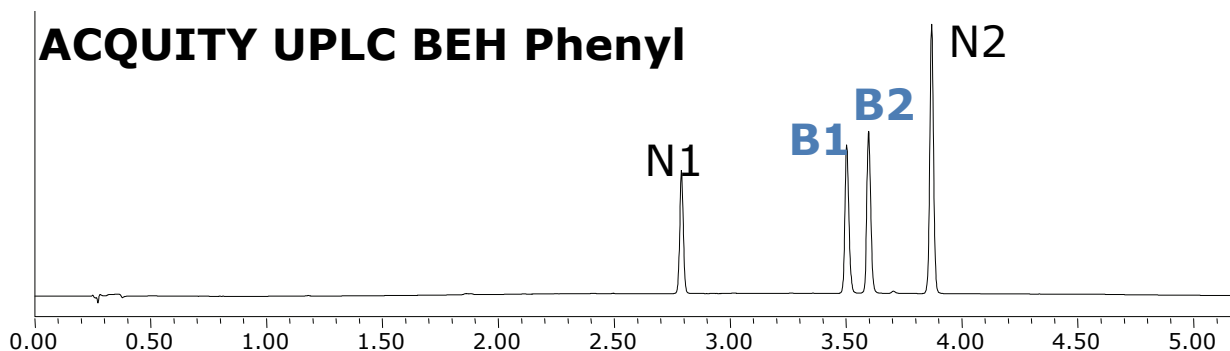
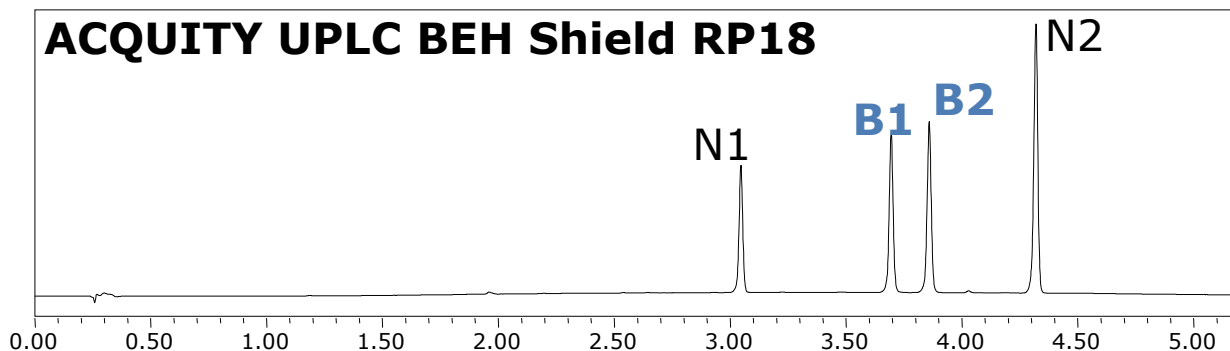
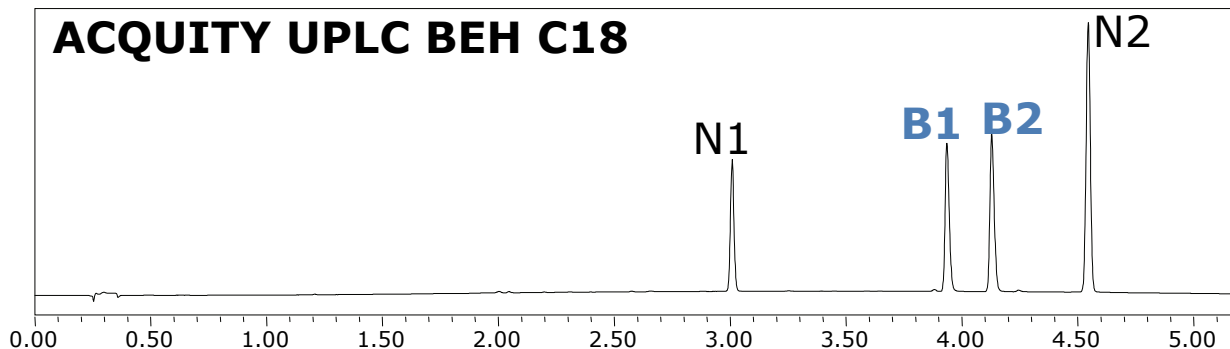
Dramatic change in selectivity

Selectivity Tools: Combining Chemical Factors

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Maximizing Selectivity Differences: Basic and Neutral Compounds



Acetonitrile, pH 10.0

Test Probes:

B1 imipramine

B2 amitriptyline

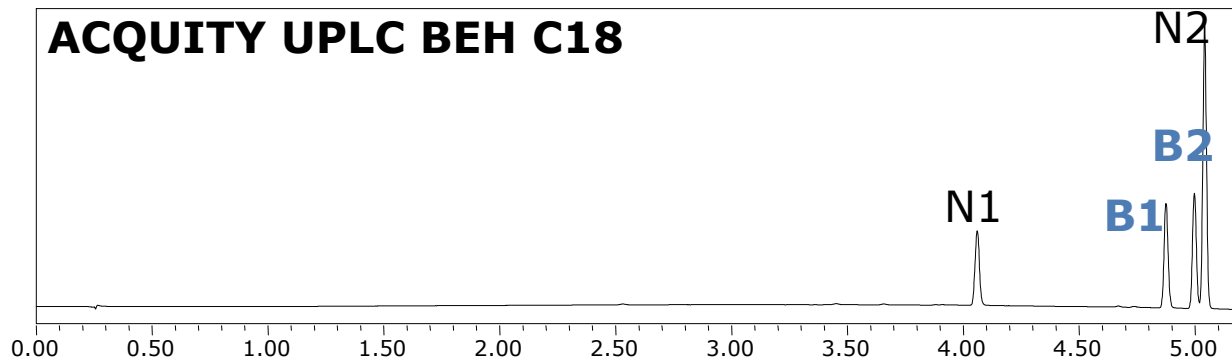
N1 flavone

N2 octanophenone

Small differences
in stationary
phase selectivity
occur with
un-ionized
analytes and
acetonitrile as
organic modifier

Maximizing Selectivity Differences: Basic and Neutral Compounds

ACQUITY UPLC BEH C18



Methanol, pH 10.0

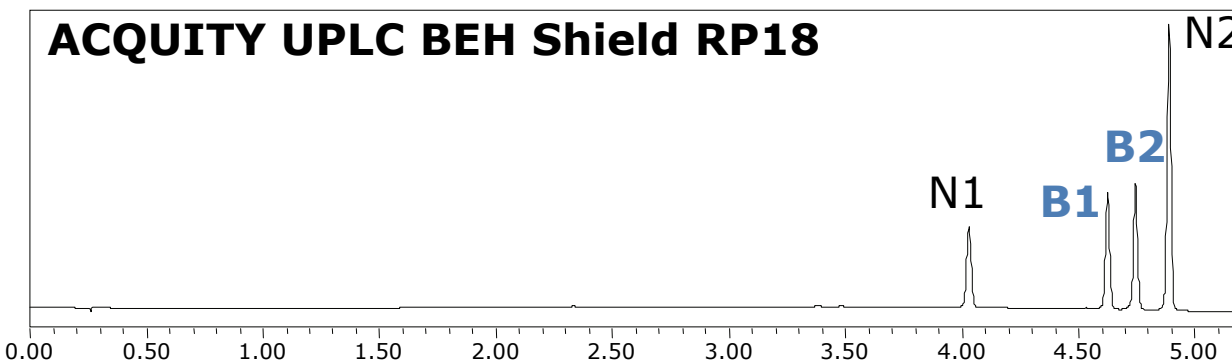
Test Probes:

B1 imipramine
B2 amitriptyline

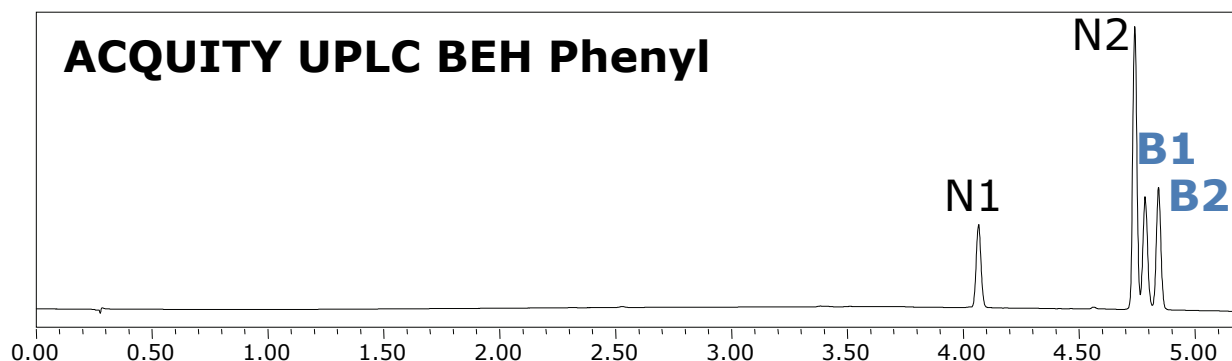
N1 flavone

N2 octanophenone

ACQUITY UPLC BEH Shield RP18

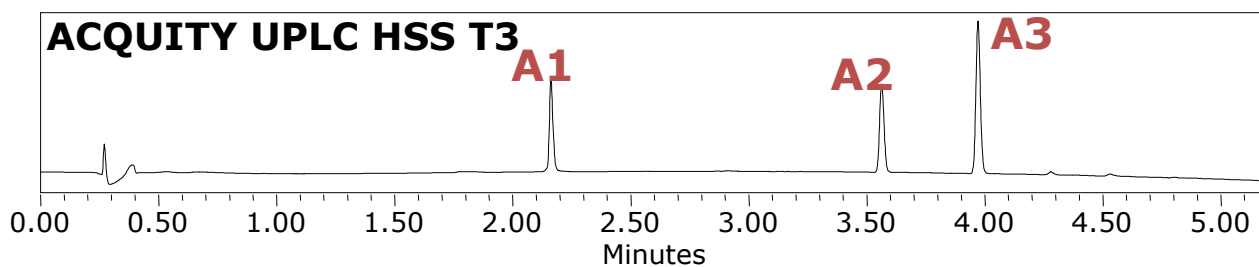
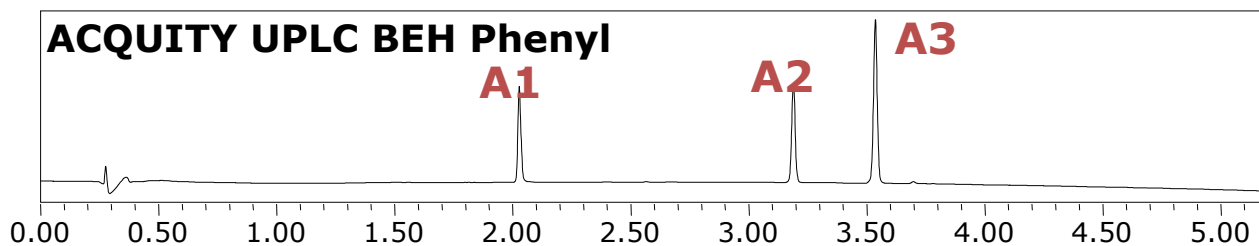
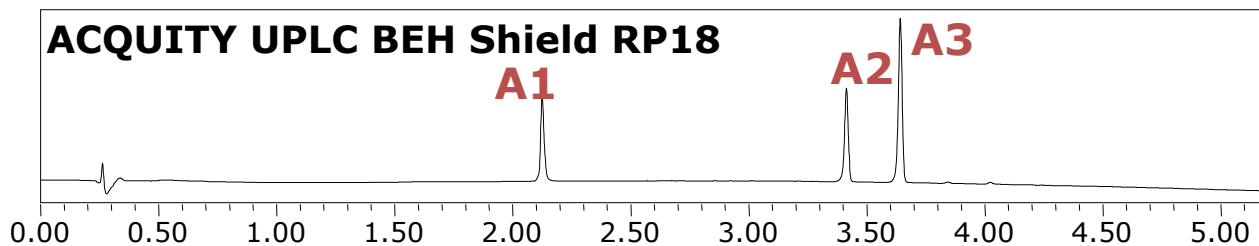
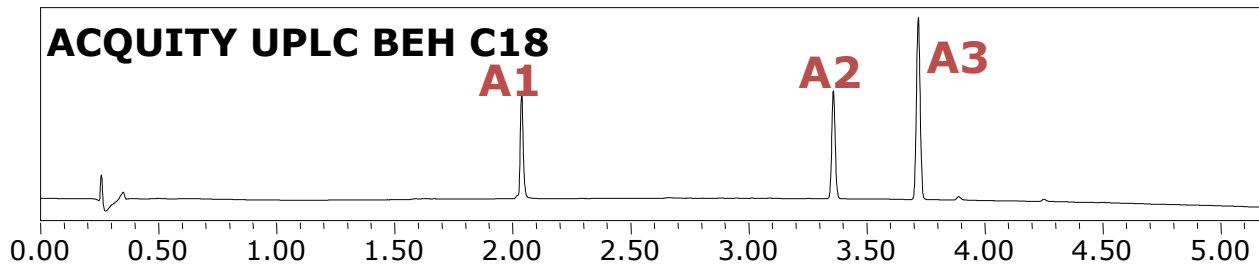


ACQUITY UPLC BEH Phenyl



Large differences
in stationary
phase selectivity
with un-ionized
analytes and
methanol as
organic modifier

Maximizing Selectivity Differences: Acidic Compounds



Acetonitrile, pH 3.0

Test Probes:

A1 1-pyrenesulfonic acid

A2 diclofenac

A3 dinoseb

Small differences
in stationary
phase selectivity
with un-ionized
analytes
and acetonitrile
as organic
modifier

Maximizing Selectivity Differences: Acidic Compounds

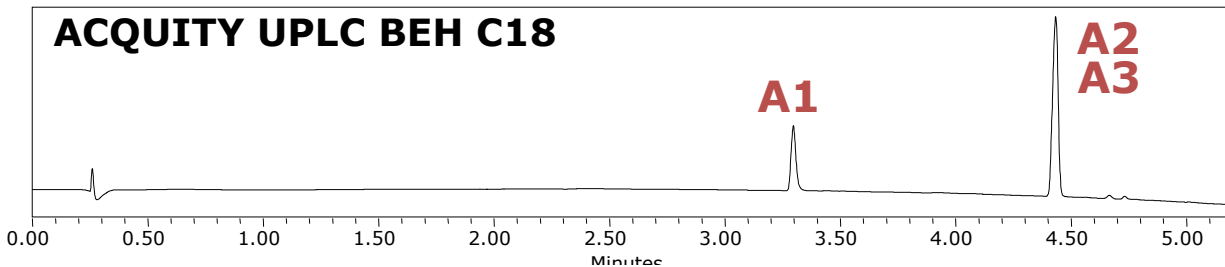
Methanol, pH 3.0

Test Probes:

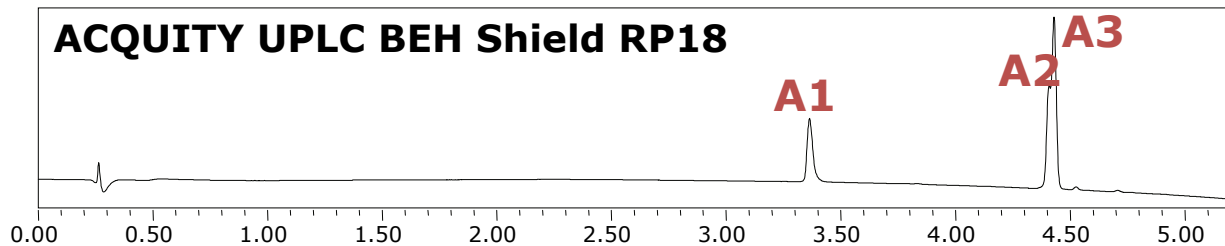
- A1** 1-pyrenesulfonic acid
- A2** diclofenac
- A3** dinoseb

Large differences
in stationary
phase selectivity
with un-ionized
analytes and
methanol as
organic modifier

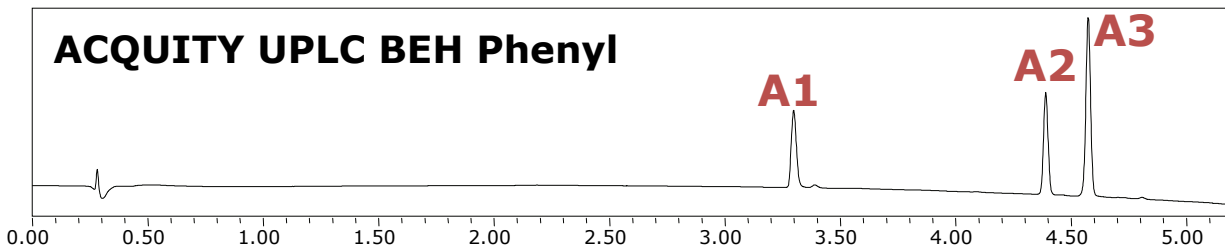
ACQUITY UPLC BEH C18



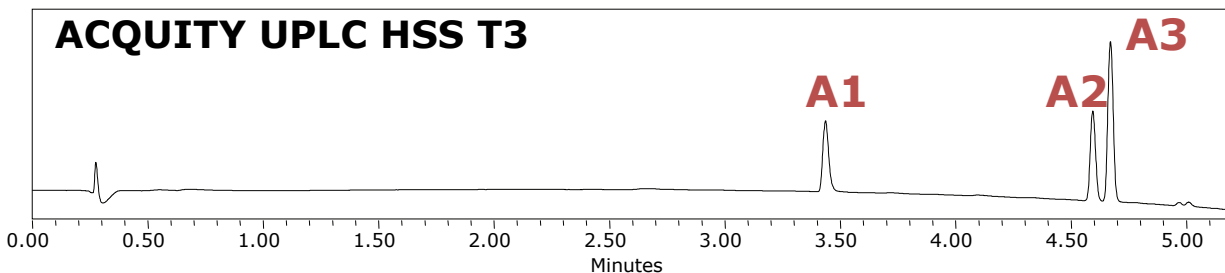
ACQUITY UPLC BEH Shield RP18



ACQUITY UPLC BEH Phenyl



ACQUITY UPLC HSS T3



- Analytes in their un-ionized forms yield greater retention
- Methanol increases retention of all components compared to acetonitrile
- Large differences in selectivity are observed when change in pH alters charge state
- These four bonded phases yield similar results employing acetonitrile
- **Largest selectivity** differences between bonded phases occurred with **methanol** as organic modifier and **analytes** in their **un-ionized** state

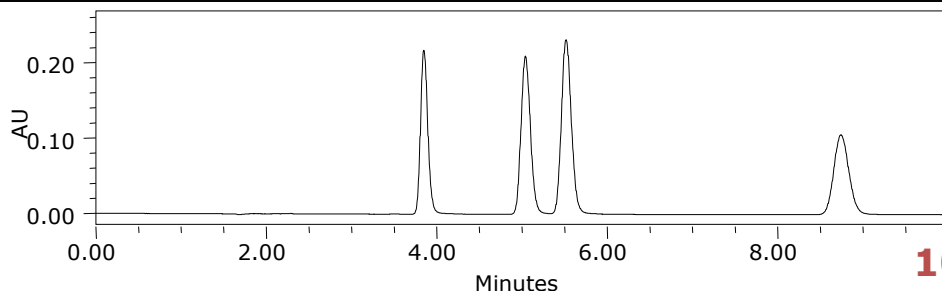
- Manipulation of parameters for method development remain the same, HPLC or UPLC separations
 - Acetonitrile and methanol
 - pH 3 and pH 10
- BEH particle technology enables the exploration of pH extremes in method development
 - Stability from pH 1 - 12
- Under the conditions and analytes of this study, large selectivity differences occurred between bonded phases with
 - Methanol as organic modifier
 - Analytes in their un-ionized state
- Evaluation of data from the complete screening matrix was essential to fully understand the analytes' chromatographic behavior

Why UPLC technology for method development?

Same Resolution and Selectivity with Increased Speed - Constant L/dp

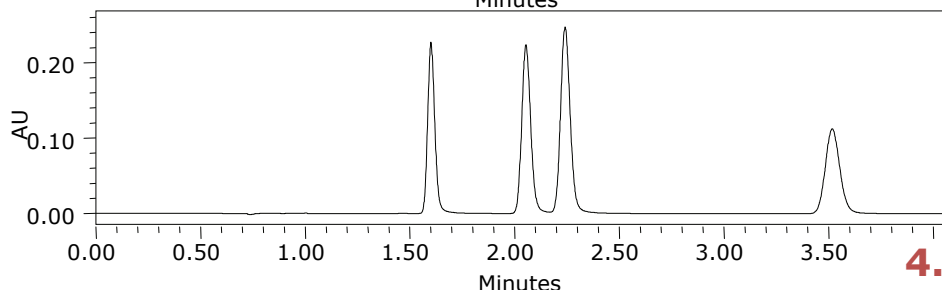
Waters

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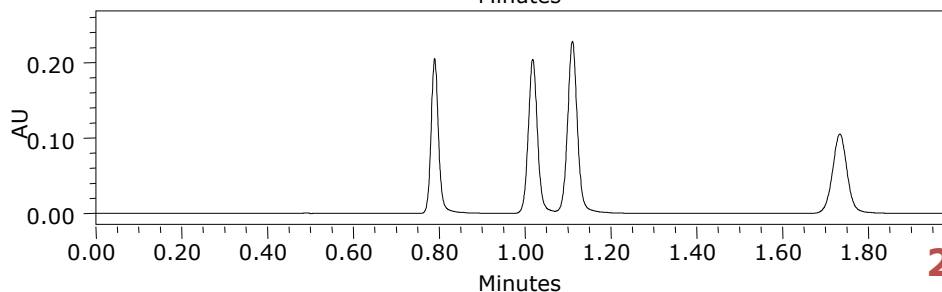
5 μm - 150 mm
F = 200 μL/min
Injection = 5.0 μL
 $Rs_{(2,3)} = 2.28$

10.00



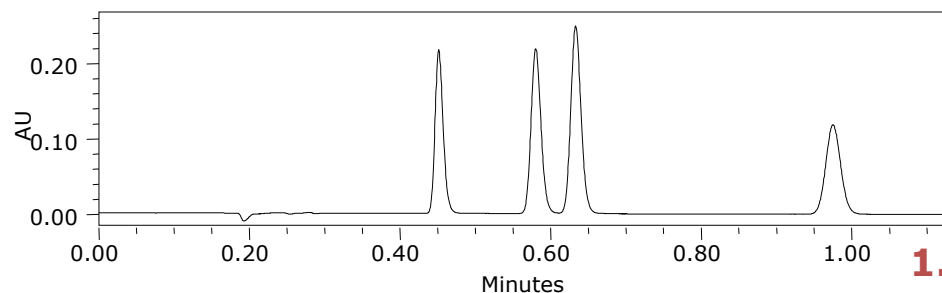
3.5 μm - 100 mm
F = 300 μL/min
Injection = 3.3 μL
 $Rs_{(2,3)} = 2.32$

4.00



2.5 μm - 75 mm
F = 500 μL/min
Injection = 2.5 μL
 $Rs_{(2,3)} = 2.34$

2.00



1.7 μm - 50 mm
F = 600 μL/min
Injection = 1.7 μL
 $Rs_{(2,3)} = 2.29$

1.10

Develop Methods Faster with UPLC Technology: Time Savings

UPLC Gradient Conditions:

Column Dimensions: 2.1 x 50 mm, 1.7 μm

Flow Rate: 0.5 mL/min

| Gradient: | <u>Time</u> | <u>Profile</u> | | <u>Curve</u> |
|-----------|-------------|----------------|----|--------------|
| | (min) | %A | %B | |
| | 0.0 | 95 | 5 | 6 |
| | 5.0 | 10 | 90 | 6 |

Equivalent HPLC Gradient Conditions:

Column Dimensions: 4.6 x 100 mm, 3.5 μm

Flow Rate: 1.4 mL/min

| Gradient: | <u>Time</u> | <u>Profile</u> | | <u>Curve</u> |
|-----------|-------------|----------------|----|--------------|
| | (min) | %A | %B | |
| | 0.0 | 95 | 5 | 6 |
| | 15.0 | 10 | 90 | 6 |

Peak Capacity (P_c) = 150

Peak Capacity (P_c) = 150

$$P_c = 1 + \frac{t_g}{W}$$

Develop Methods Faster with UPLC: Time Savings versus 3.5 μ m HPLC Column

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UPLC Method Development Protocol

2.1 x 50 mm, 1.7 μ m, 0.5 mL/min

pH 3/ acetonitrile Time

| | |
|-----------------------------------------|--------|
| Flow ramp | 5 min |
| Column conditioning (2 blank gradients) | 11 min |
| Sample injection (2 replicates) | 11 min |

pH 3/ methanol

| | |
|-----------------------------------------|--------|
| Flow ramp | 5 min |
| Column conditioning (2 blank gradients) | 11 min |
| Sample injection (2 replicates) | 11 min |
| Column purge | 6 min |

pH 10/ acetonitrile

| | |
|-----------------------------------------|--------|
| Flow ramp | 5 min |
| Column conditioning (2 blank gradients) | 11 min |
| Sample injection (2 replicates) | 12 min |

pH 10/ methanol

| | |
|-----------------------------------------|---------|
| Flow ramp | 5 min |
| Column conditioning (2 blank gradients) | 11 min |
| Sample injection (2 replicates) | 11 min |
| Column purge | 6 min |
| | 120 min |

SCOUTING TIME **2 Hours/ Hybrid column
x 3 columns**

**1 Hour/ Silica column
x 1 column**

TOTAL SCOUTING TIME **7 HOURS**

EQUIVALENT HPLC Method Development Protocol, 3.5 μ m

4.6 x 100 mm, 3.5 μ m, 1.4 mL/min

pH 3/ acetonitrile Time

| | |
|-----------------------------------------|----------|
| Flow ramp | 5 min |
| Column conditioning (2 blank gradients) | 37.7 min |
| Sample injection (2 replicates) | 37.7 min |

pH 3/ methanol

| | |
|-----------------------------------------|-----------|
| Flow ramp | 5 min |
| Column conditioning (2 blank gradients) | 37.7 min |
| Sample injection (2 replicates) | 37.7 min |
| Column purge | 20.56 min |

pH 10/ acetonitrile

| | |
|-----------------------------------------|----------|
| Flow ramp | 5 min |
| Column conditioning (2 blank gradients) | 37.7 min |
| Sample injection (2 replicates) | 37.7 min |

pH 10/ methanol

| | |
|-----------------------------------------|------------|
| Flow ramp | 5 min |
| Column conditioning (2 blank gradients) | 37.7 min |
| Sample injection (2 replicates) | 37.7 min |
| Column purge | 20.56 min |
| | 362.72 min |

SCOUTING TIME **6 Hours/ Hybrid column
x 3 columns**

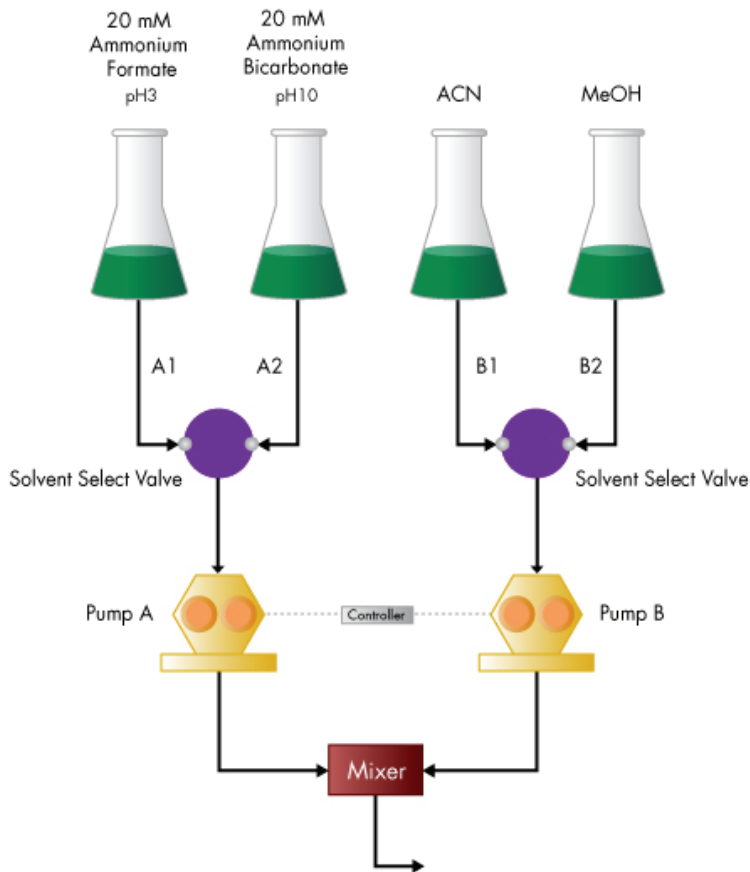
**3 Hours/ Silica column
x 1 column**

TOTAL SCOUTING TIME **21 HOURS**

UPLC scouting **3X faster** than 3.5 μ m HPLC

- Approaches Towards Method Development
- Selectivity and Retention Tools
 - Stationary Phase
 - Solvent
 - pH
- ■ Method Development Strategy
- Applications
- Conclusions

Automated Method Development and Validation



- Automated Method Development
 - ACQUITY UPLC Column Manager, 4-column selection device
 - Utilize 2.1 x 50 mm columns
 - ACQUITY UPLC Binary Solvent Manager, solvent select valves
 - Utilize pH 3 and pH 10 buffers
 - Utilize methanol and acetonitrile
 - Fast 5-minute gradient 5 – 90% B, Flow = 0.5 mL/min
- Automated Method Validation
 - Empower™ 2 Method Validation Manager, streamlines method validation process



Selectivity Scouting Protocol

2.1 x 50 mm, <2 μm

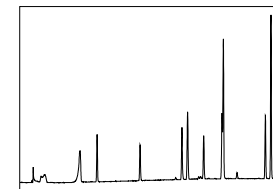
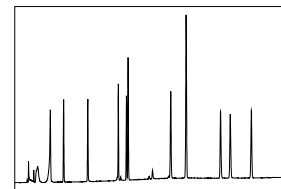
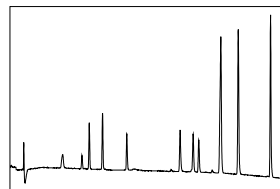
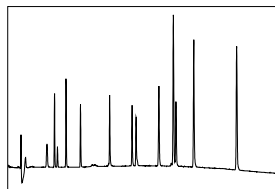
pH 3, ACN

pH 3, MeOH

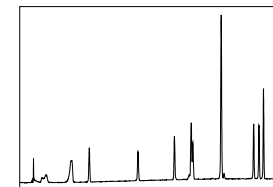
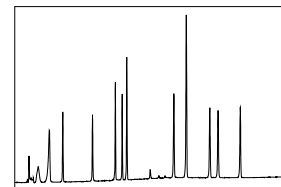
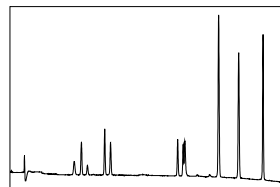
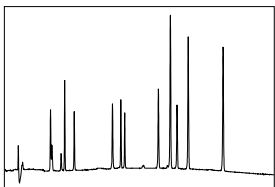
pH 10, ACN

pH 10, MeOH

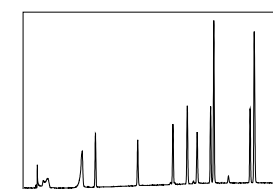
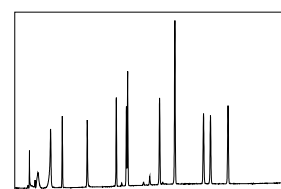
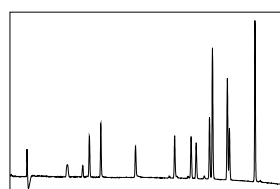
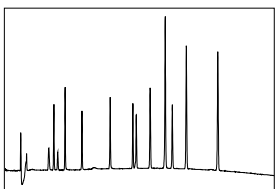
ACQUITY UPLC
BEH C₁₈



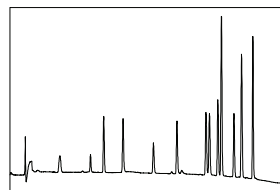
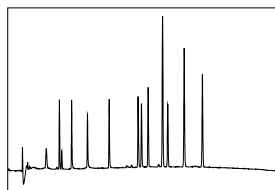
ACQUITY UPLC
BEH Shield RP₁₈



ACQUITY UPLC
BEH Phenyl



ACQUITY UPLC
HSS T3



Optimization

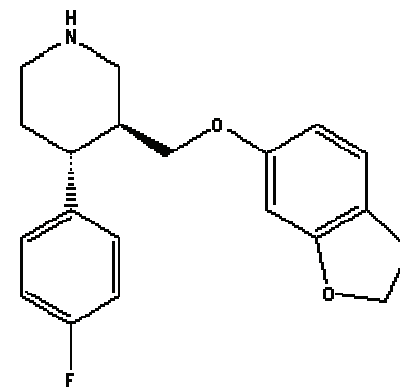
- Approaches Towards Method Development
- Selectivity and Retention Tools
 - Stationary Phase
 - Solvent
 - pH
- Method Development Strategy
- ■ Applications
 - Method Optimization
- Conclusions

- As we go through the data for each separation, we should have some questions in mind to help select the best pH, organic solvent and column
 - Do I need resolution of every peak or simply my main component?
 - How much resolution do I need?
 - How much retention do I need and what is the retention profile?
 - What are the tailing factors?
 - Do I want a gradient or isocratic method?
- Step 1: Review data and select pH
- Step 2: Review data and select organic solvent and column
- Step 3: Optimize/fine-tune separation

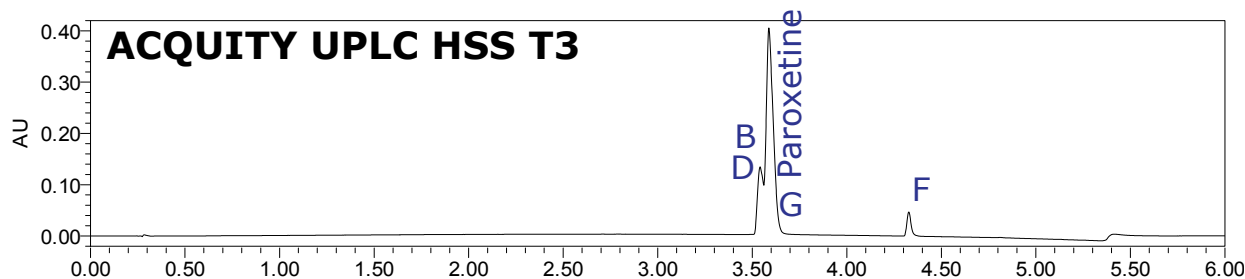
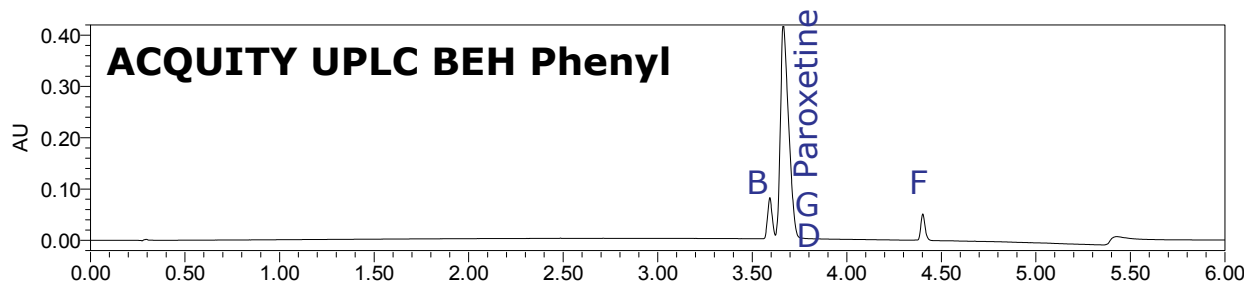
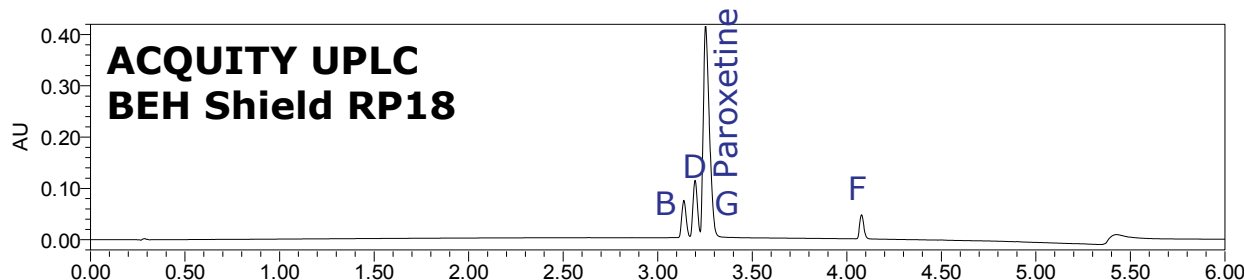
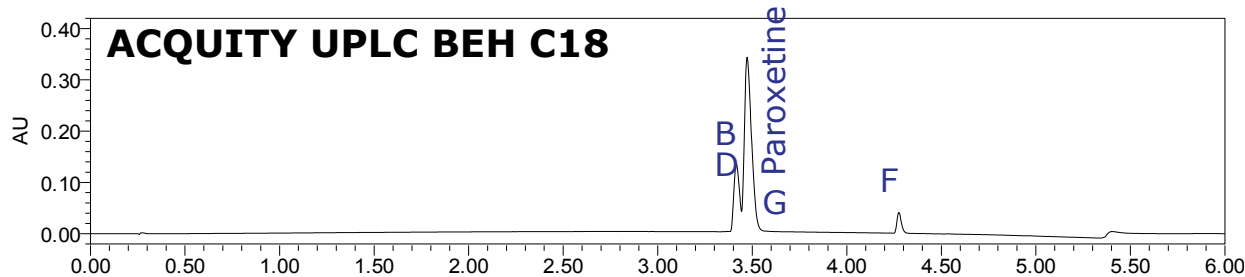
Automated Method Development and Validation of Paroxetine and Related Compounds

- Method Development
 - Use systematic screening protocol
 - Paroxetine (API) concentration: 0.2 mg/mL in 50:50 MeOH:H₂O
 - Related compounds at 10% concentration of API for easy identification during scouting
- Method Optimization
 - Related compounds at 0.1% concentration of API
 - Related compounds B, D, F and G
- Method Validation
 - Empower™ 2 Method Validation Manager

Paroxetine
m.w. 374.8



Stationary Phase Selectivity: Paroxetine



Methanol pH 3.0

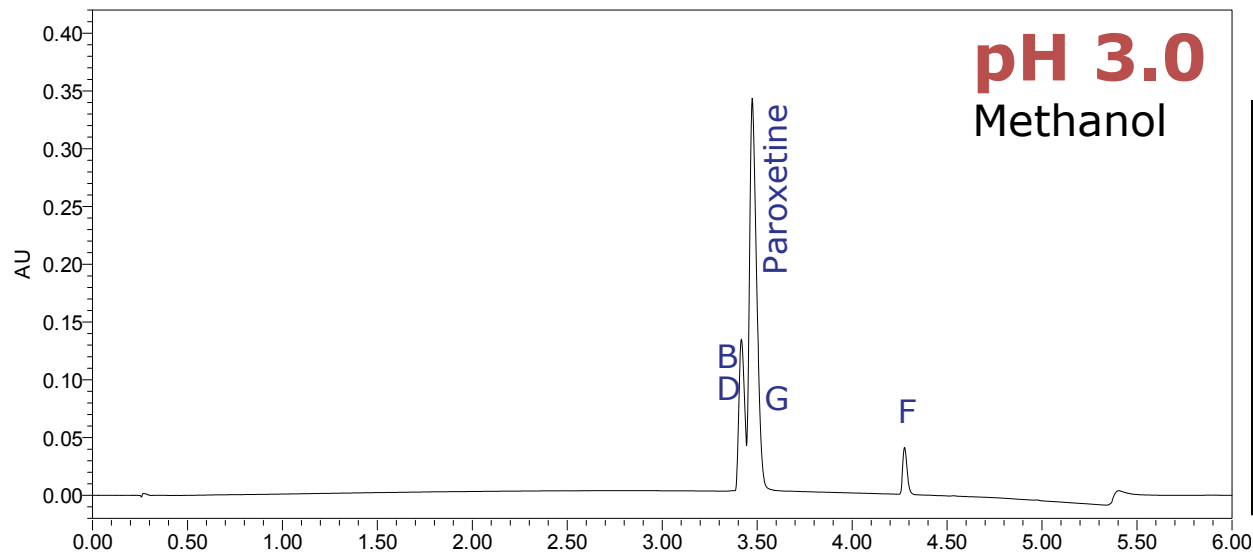
Observation:

Poor resolution of
paroxetine and
related compounds
(RC)

Action:

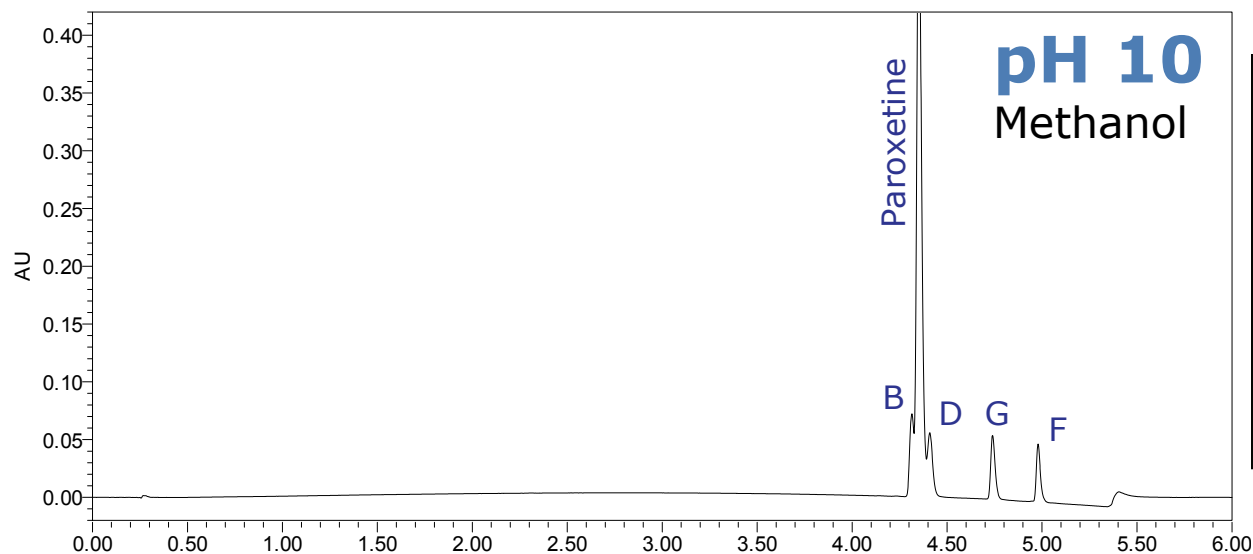
Investigate high pH

pH Selectivity: Paroxetine



Observation:

Better retention and resolution of API from RC due to neutral charge state of analytes at alkaline pH



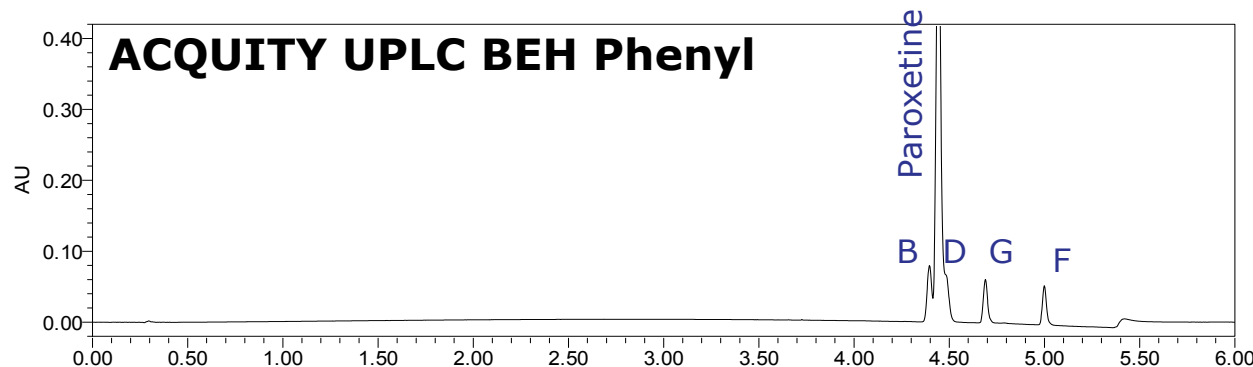
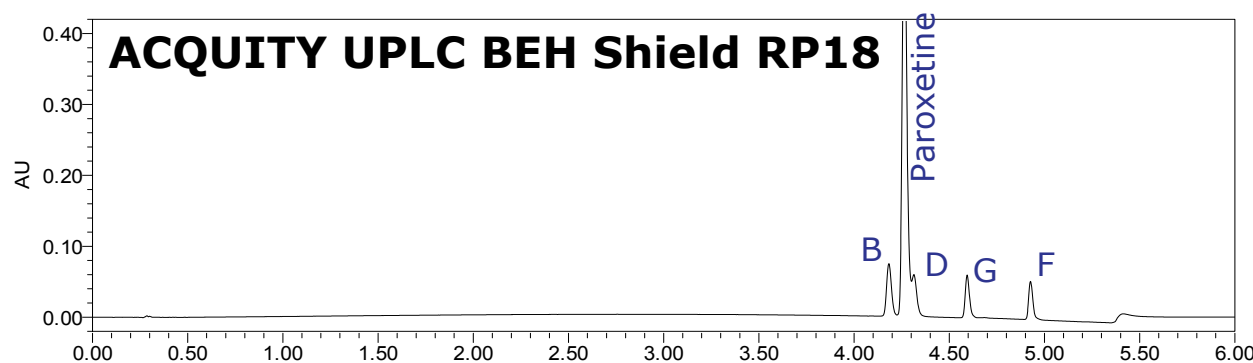
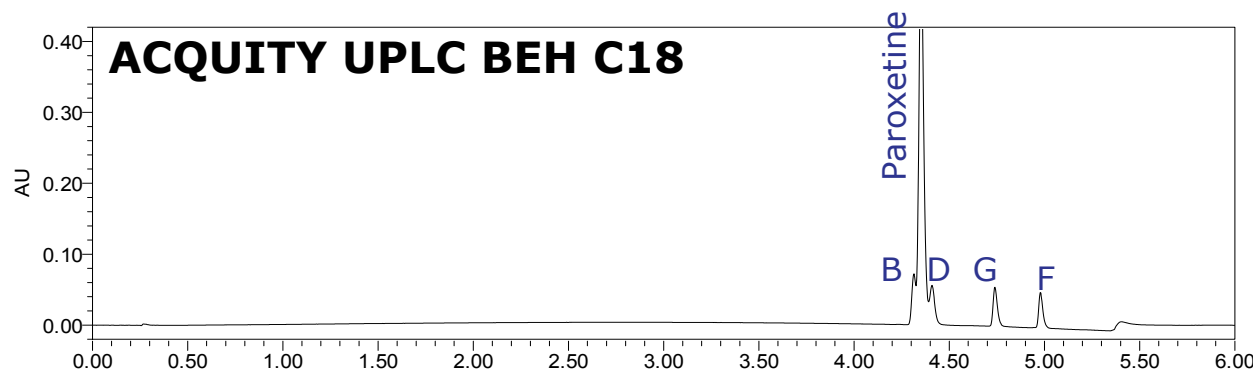
Actions:

Select pH 10 due to better separation

Compare stationary phase selectivity with pH 10 buffer

Stationary Phase Selectivity: Paroxetine

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Methanol pH 10.0

Observation:

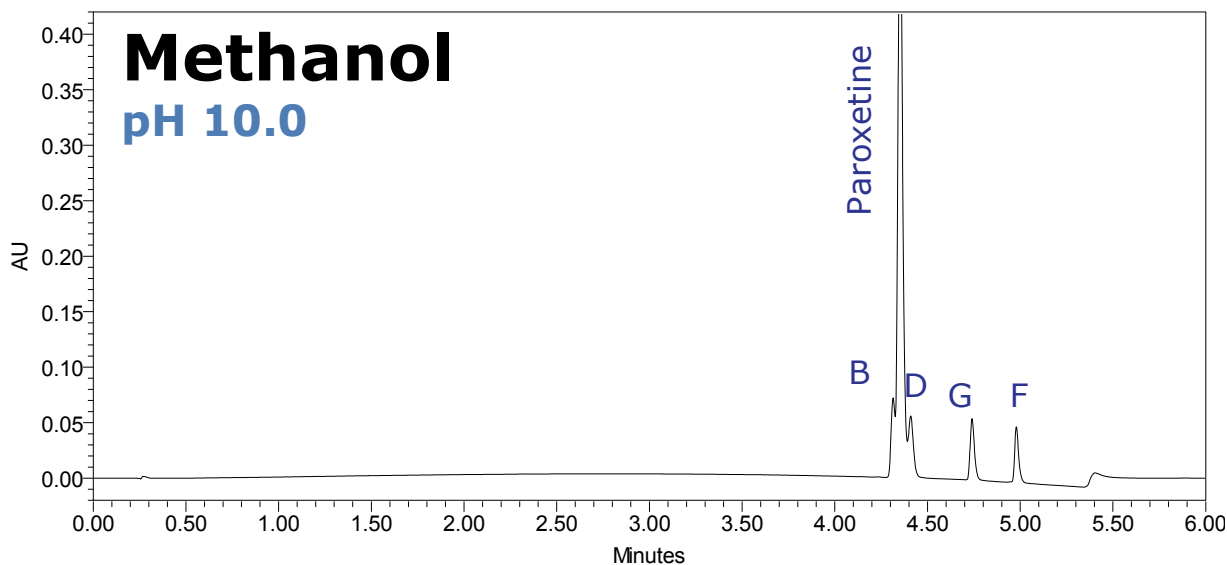
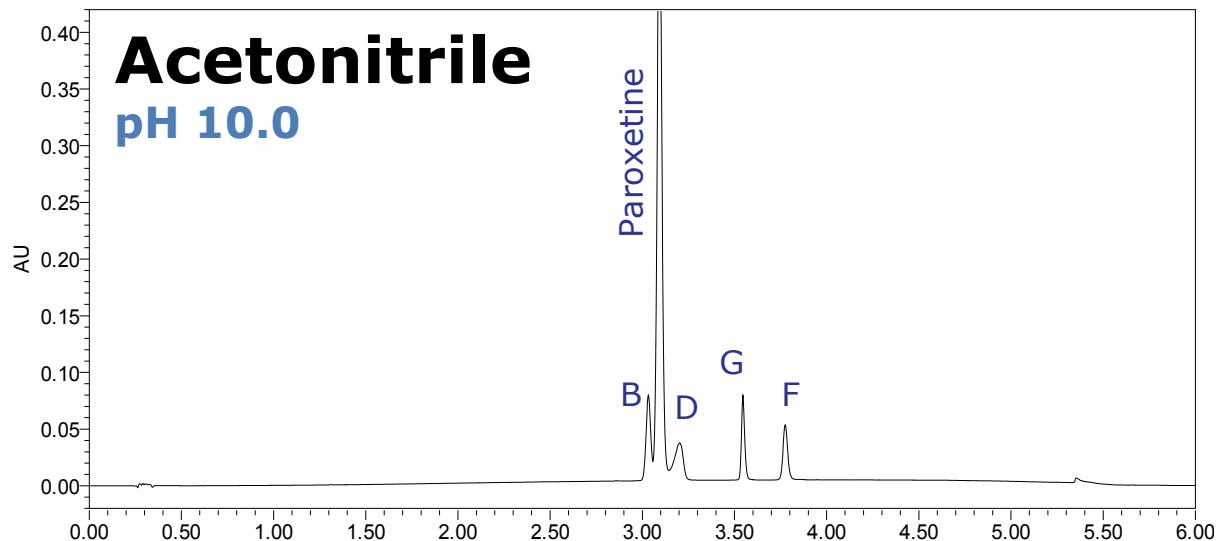
Any column may provide successful separation

Actions:

Select ACQUITY UPLC BEH C18

Compare selectivity between organic modifiers

Solvent Selectivity: Paroxetine



Observations:

Methanol is weaker elution solvent resulting in greater retention

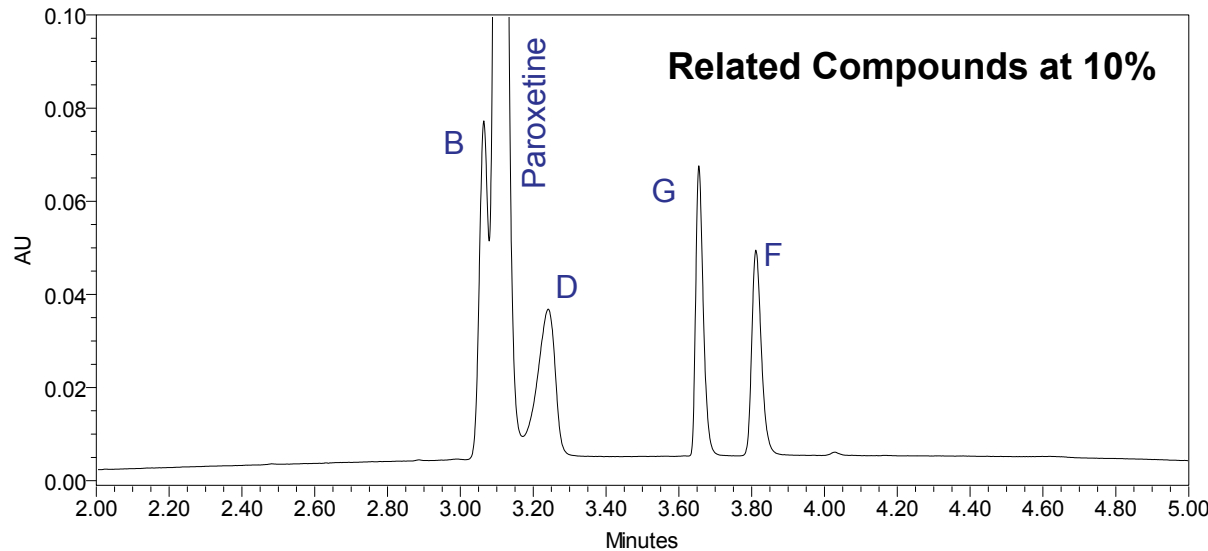
Better resolution exhibited with acetonitrile as organic modifier

Actions:

Select acetonitrile as organic modifier

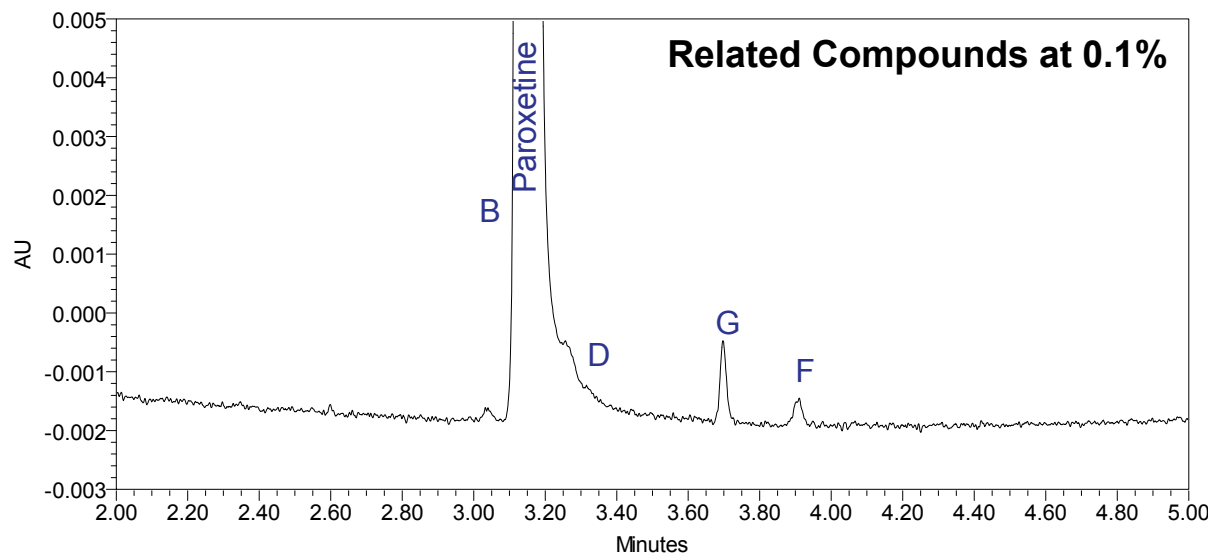
Optimize separation using appropriate concentration of RC

Related Compounds at 0.1% Concentration of Paroxetine



Observation:

Inadequate resolution among paroxetine and related compounds B and D due to disparate levels of concentration



Action:

Optimize Separation

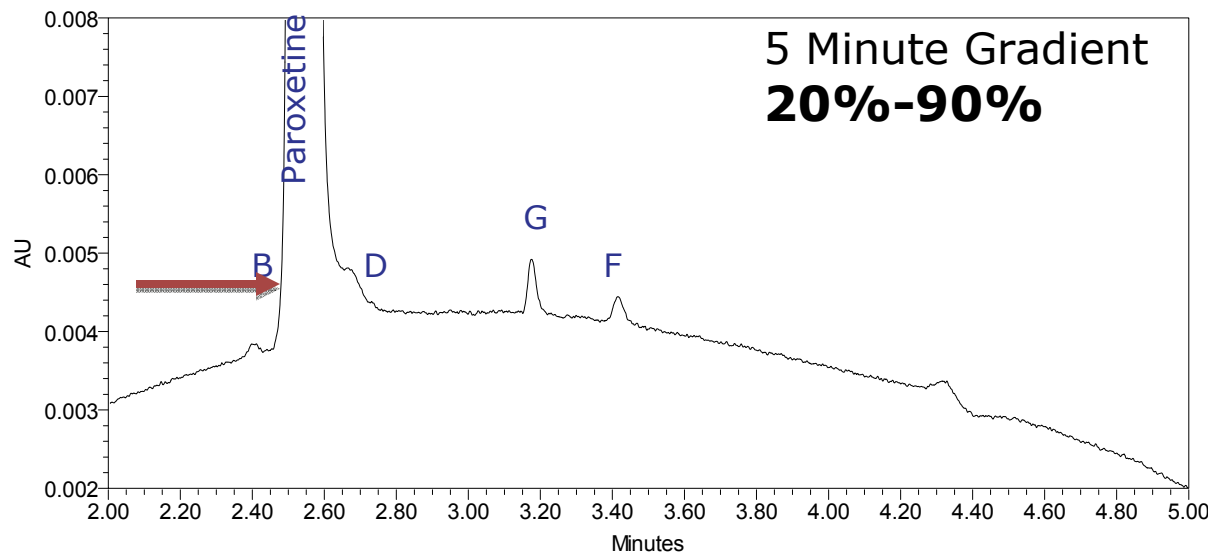
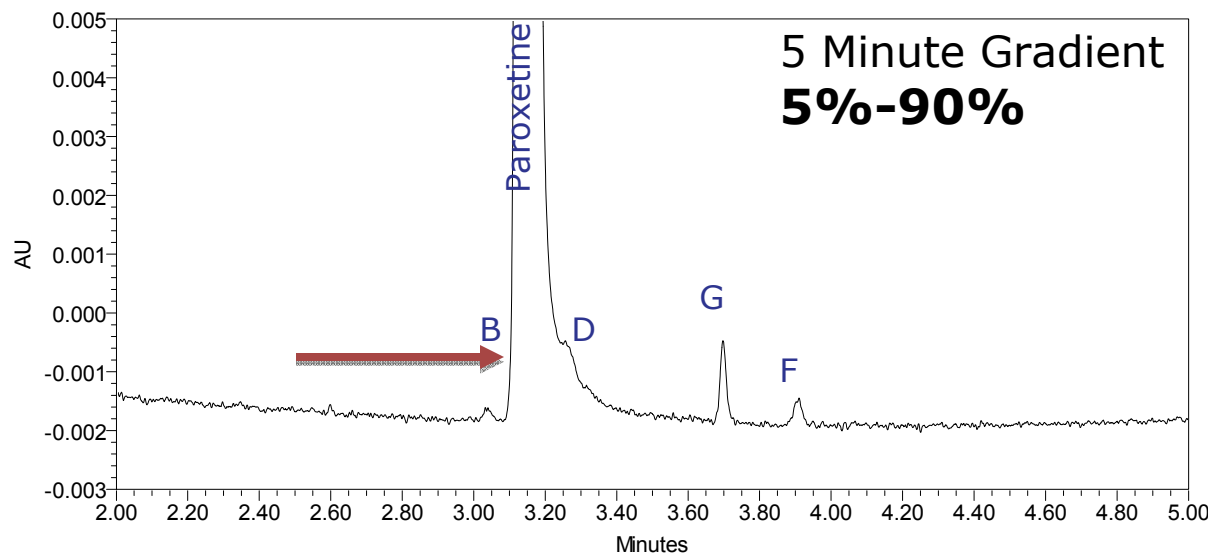
Factors that Control Retentivity and Selectivity: Optimization Tools

- Once the pH, mobile phase, and column are selected, the method can be fine-tuned and optimized
- Parameters for optimization
 - Gradient slope
 - Temperature

Method Optimization: Gradient Slope

- Shallower gradient slope may improve resolution
 - Decreasing gradient slope will decrease sensitivity
- Steeper gradient slope may compress the peaks and often reduce the resolution
 - Increasing gradient slope will increase sensitivity
- Changing gradient slope is a balance between peak heights relative to resolution
- Changes in retentivity and selectivity

Method Optimization: Gradient Slope



Acetonitrile pH 10.0
30 °C

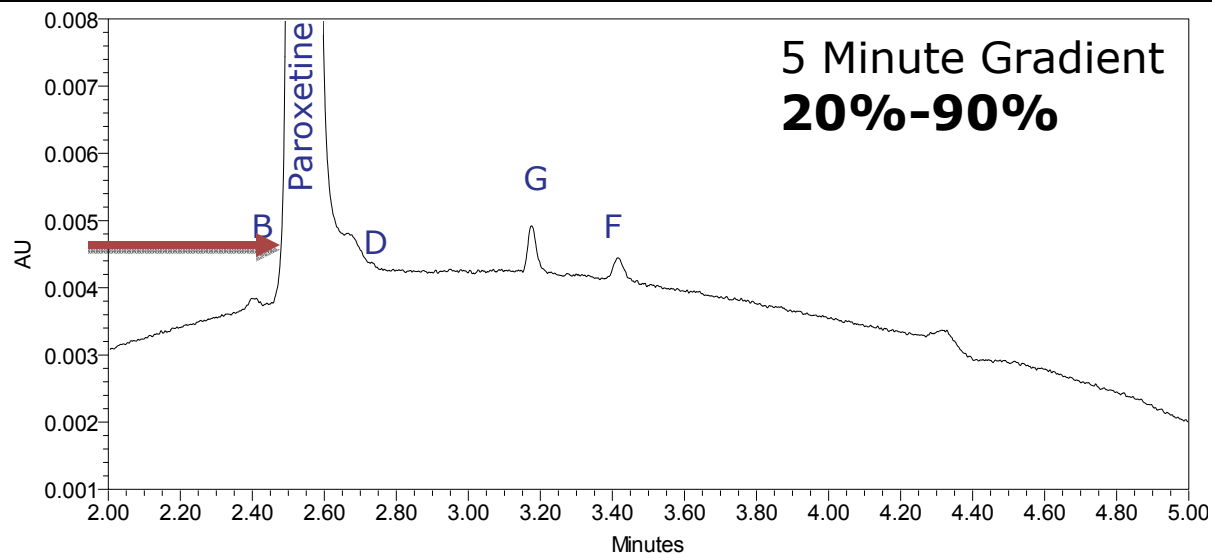
Observations:

Marginal improvement in separation of RC from parent compound with shallow gradient slope

Action:

Alter gradient endpoint to produce shallower slope

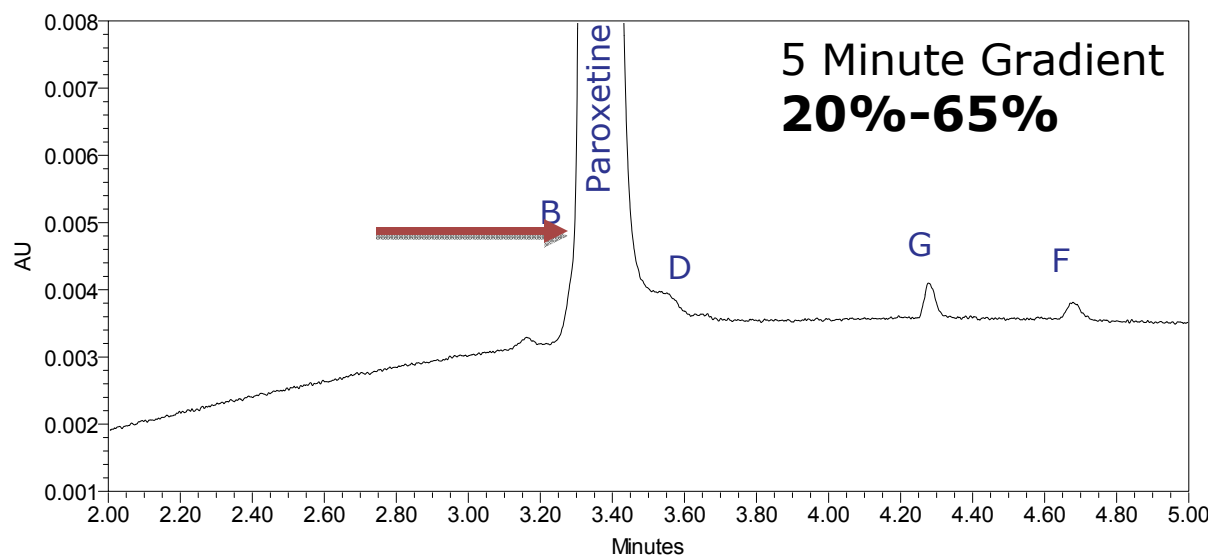
Method Optimization: Gradient Slope



Acetonitrile pH 10.0
30 °C

Observations:

Resolution remains inadequate with shallow gradient slope



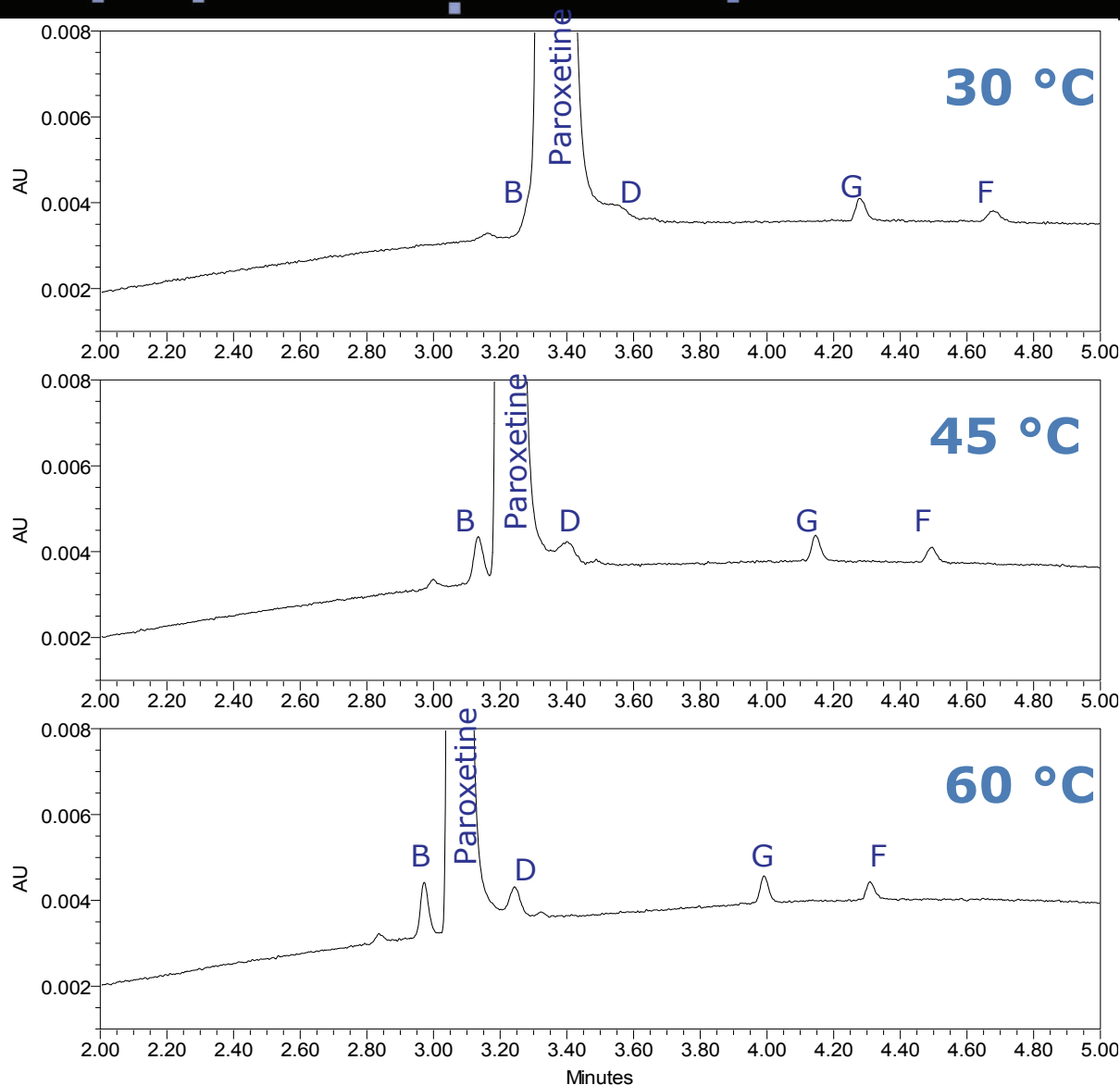
Action:

Investigate column temperature

Method Optimization: Influence of Temperature

- Reduced mobile phase viscosity
- Lower backpressure
 - If flow rate is held constant
- Improve analyte diffusivity
 - Higher optimal linear velocity
- Changes in retention and selectivity

Method Optimization: Column Temperature



Acetonitrile pH 10.0
Observations:

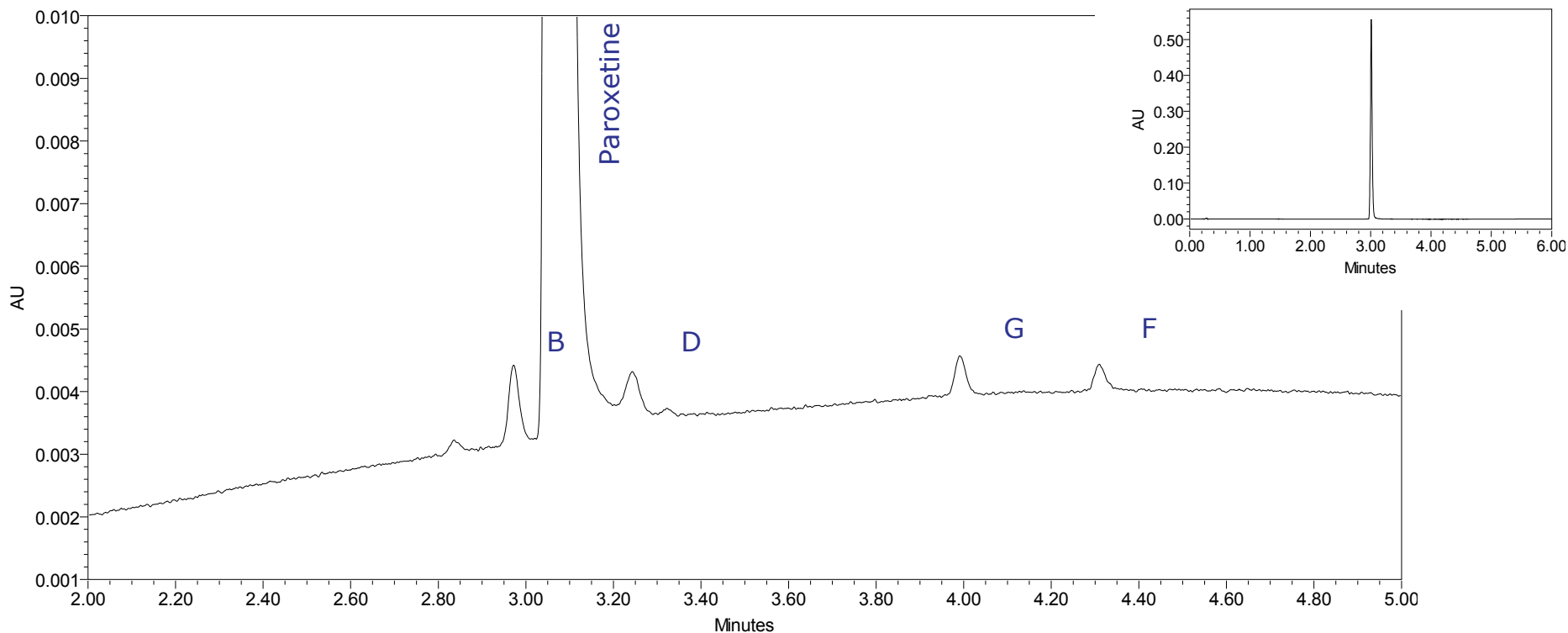
Higher temperature improves separation of RC from paroxetine

Peak shape improves as temperature increases

Action:

Select 60 °C for best resolution and peak shape

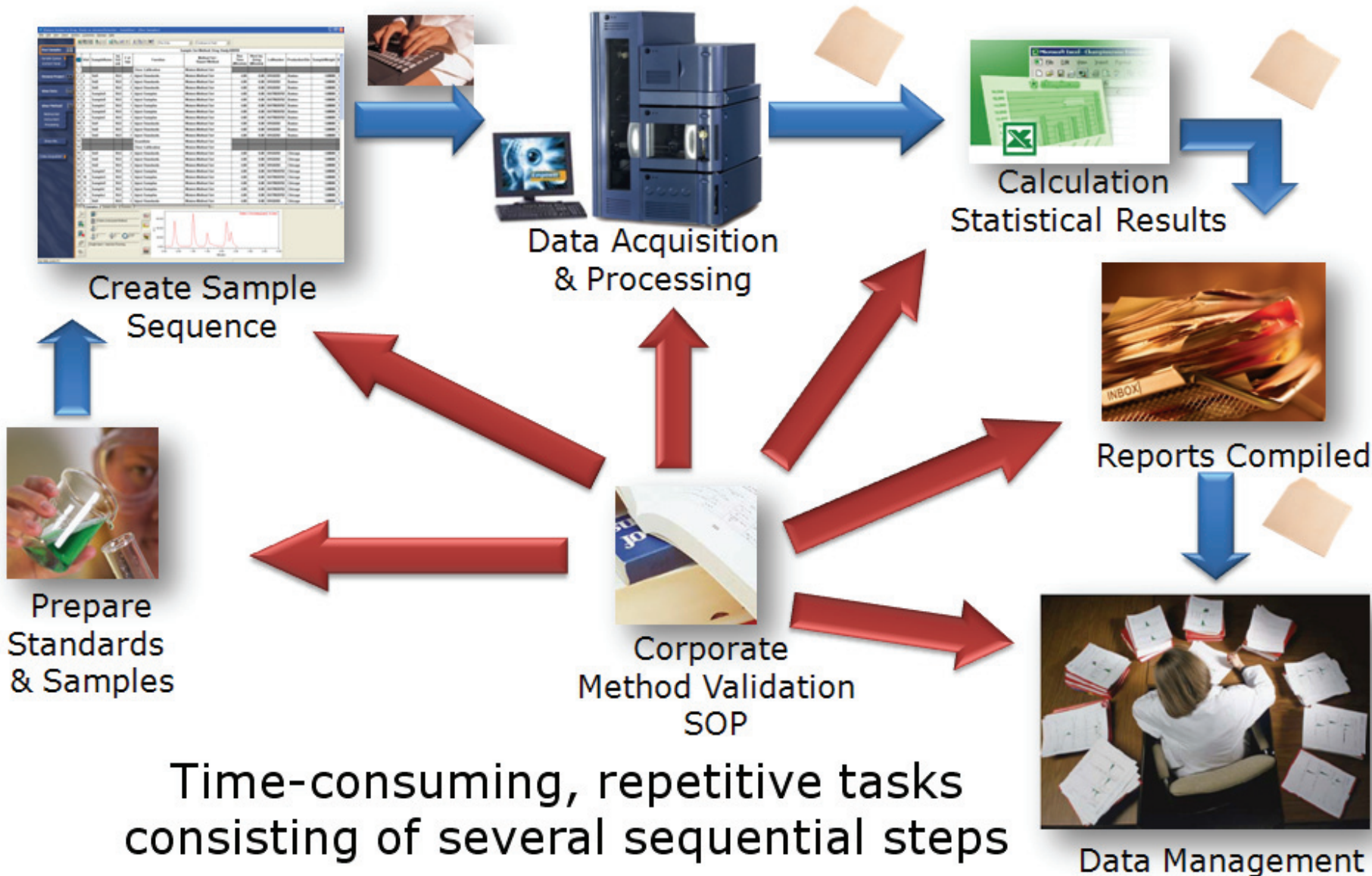
Final Method: Paroxetine and Related Compounds at 0.1%



| Compound | USP Rs |
|--------------------|--------|
| Related compound B | |
| Paroxetine | 1.95 |
| Related compound D | 3.07 |
| Related compound G | 13.00 |
| Related compound F | 6.74 |

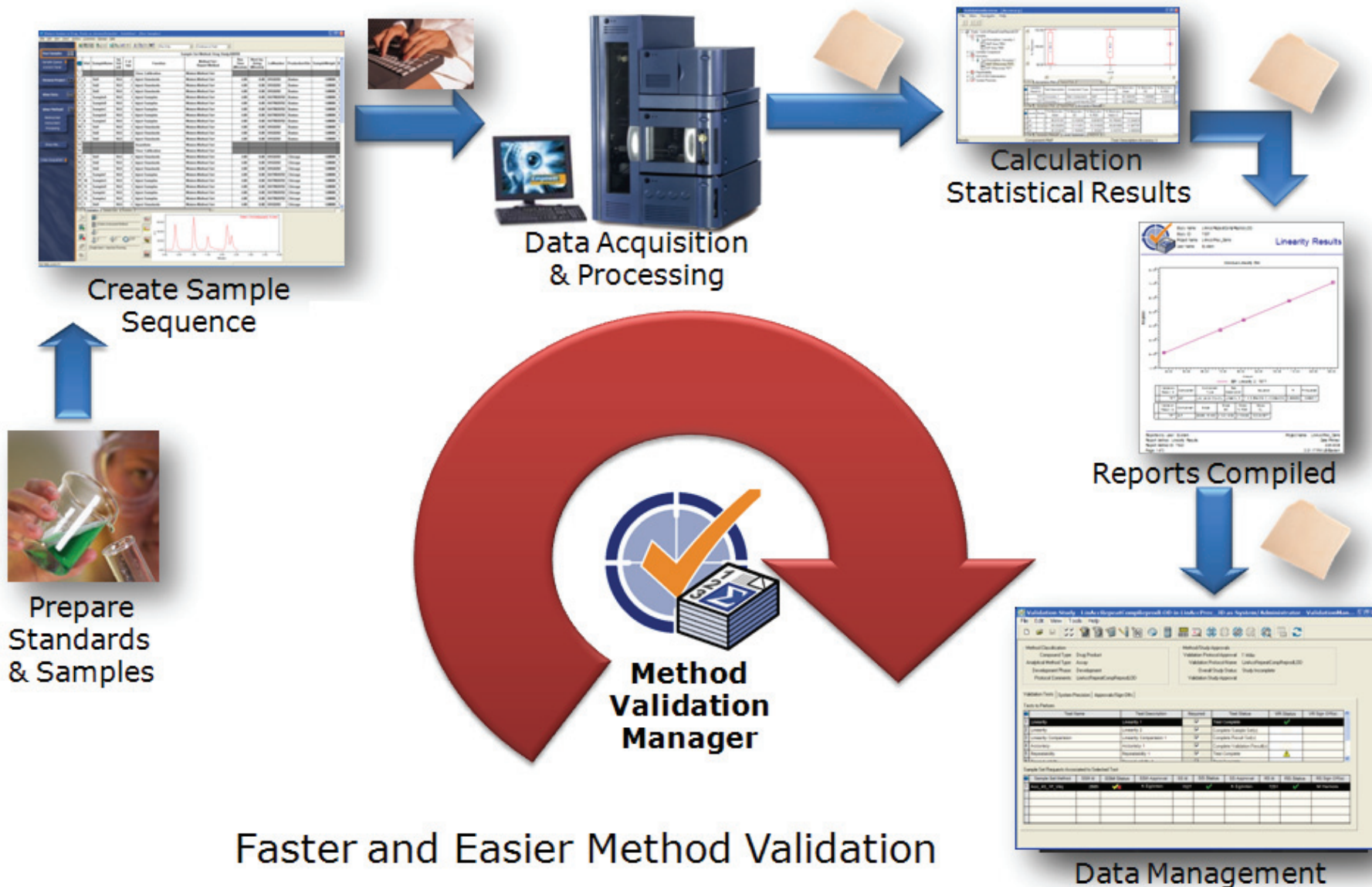
ACQUITY UPLC BEH C18
2.1 x 50 mm, 1.7 μ m
20 mM ammonium bicarbonate, pH 10
60 °C
5 Min Gradient, 20%-65% acetonitrile
F = 0.5 mL/min
Inj. Vol. 4 μ L
UV @ 295 nm

Analytical Method Validation



Analytical Method Validation with Empower 2 Method Validation Manager

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Automated Method Validation Manager: Paroxetine Validation Results

| Parameter | Acceptance Criteria | Reported Value | Pass/Fail |
|-----------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------|-----------------------------------------------|-----------|
| Linearity | $R^2 \geq 0.995$ | 0.9999 | pass |
| | Residuals $\leq 2.0\%$ RSD | 1.74% RSD | pass |
| Accuracy | 80 – 120 % | 97 – 102 % | pass |
| Intermediate Precision | Variance Component $\leq 10\%$ RSD Peak Area (Analyst, Instrument, Column) | Analyst 1.33% RSD | pass |
| | | Instrument 7.72% RSD | pass |
| | | Column 0.00% RSD | pass |
| | | | |
| LOD of impurities | Impurities 0.1% of active at 0.2 mg/mL | 0.05% of active at 0.2 mg/mL (s/n 2.2 – 6.23) | pass |
| Method Robustness Peak Area | Variance Component $\leq 2\%$ RSD Peak Area (Buffer strength, Additive Conc., Column Temperature) | 0.06 – 1.64% RSD | pass |
| Method Robustness Retention Time | Variance Component $\leq 5\%$ RSD Retention Time (Buffer strength, Additive Conc., Column Temperature, Flow Rate, Injection Volume) | 0.00 – 2.57% RSD | pass |

Method Development and Validation Timeline: Instrument Time

UPLC Method Development and Validation Timeline

2.1 x 50 mm, 1.7 μ m, 0.5 mL/min

Screening

4 Columns, 2 Organics, 2 pHs

Time

7.0 hours

Optimization

Gradient Slope and Temperature

1.7 hours

Validation

Accuracy, linearity, repeatability,
Reproducibility, LOD/LOQ,
Intermediate precision, robustness

21.1 hours

TOTAL TIME

29.8 HOURS

- Achieve **more resolution, faster** by utilizing sub-2 μm UPLC columns
- Principles of method development remain the same
- **Broad range of column selectivity** to successfully develop methods efficiently
 - BEH C₁₈, BEH Shield RP18, BEH Phenyl and HSS T3
- UPLC Technology allows for **efficient method development**
 - Systematic scouting protocol
 - Automated column selection
 - High resolution sub 2 μm column technology
- UPLC Technology and Empower 2 software can significantly **improve laboratory productivity and compliance**

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