SHIMADZU

LC-MS/MS method development of aflibercept using Fab-selective proteolysis nSMOL technology

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1. Introduction

Aflibercept is a biopharmaceutical drug inhibiting of vascular endothelial growth factor (VEGF) signaling and composed of the extracellular domains of human VEGF receptors 1 and 2 that are fused to the Fc portion of the human IgG1 immunoglobulin. It is important to determine an appropriate dose for medical optimization, but little amount of intraocular fluid is able to collect as a specimen in the treatment of retina. To offer the quantitative assessment method of aflibercept in biological matrix, the primary structure was confirmed using a quadrupole time-offlight (Q-TOF) mass spectrometer LCMS-9030, and quantitative analysis was performed with a triple quadrupole mass spectrometer LCMS-8060 by using nSMOL (nano-surface and molecular orientation limited proteolysis) technology.

2. Methods and Materials

The tryptic digest of aflibercept was analyzed by MS using a Q-TOF, and MS/MS analysis of the VEGF receptor region was performed to select transitions for multiple reaction monitoring (MRM) assay. The observed fragments were selected for the quantitative analysis using a triple quadrupole LC-MS/MS. A serial dilutions of aflibercept were prepared with human plasma and 5 uL was used for each assay. Using nSMOL Antibody BA Kit, aflibercept and endogenous IgGs were captured in resin pore via Protein A and selectively proteolyzed with trypsin immobilized on the surface of nanoparticles to collect the signature peptides for aflibercept.

2-1. Sample treatment for sequence confirmation of aflibercept peptides

Aflibercept (10 µg) was dissolved in 10 µL of 2 M urea containing 1 mM TCEP, reacted at room temperature for 30 minutes, then 100 µL of 20 mM tris-HCl buffer (pH 8.0) and trypsin 1 µg were added, and the mixture was treated at 37°C for 16 hours.

2-2. MS and MS/MS analysis conditions

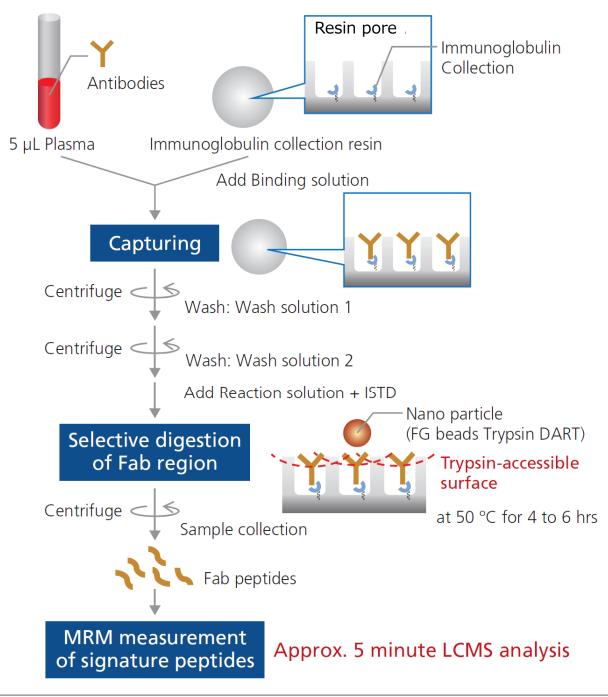
<nexera systematics<="" th="" x3=""><th>em></th><th></th><th></th></nexera>	em>									
Column	: Shim-pack GISS-HP C18 (100 mm x 2.1 mm Ι.D., 3 μm)									
Mobile phase A	: 0.1% Formic acid / Water									
Mobile phase B	: 0.1% Formic acid / Acetonitrile	Flow rate	: 0.2 mL/min							
Time program	: B conc. 1% (0-5 min) → 40% (40 min) → 95% (40.01-50 min)									
	→ 1% (50.01-60 min)									
Column temp.	: 40 °C	Injection vol.	: 5 μL							
<lcms-9030></lcms-9030>										
Ionization	: ESI, Positive mode	MS range	: <i>m/z</i> 100-1300							
CID	: 10-30 V	DL temp.	: 250 °C							
Interface temp.	: 300 °C	Heat block temp.	: 400 °C							
Nebulizer gas	: 3.0 L/min	Heating gas	: 10 L/min							
Drying gas	: 10 L/min									

2-3. LC-MS condition for MRM assay

<nexera n<="" or="" th="" x2=""><th>exera X3 system></th><th></th><th></th><th></th></nexera>	exera X3 system>			
Column	: Shim-pack GISS C18 (50 mm x 2	2.1 mm I.D., 1.9 ur	m)	
Mobile phase A	•	, - I	/	G
Mobile phase B				
Flow rate	: 0.4 mL/min (0-5.51, 6.72-7.5 min), 1 mL/min (5.53-	·6.7 min)	T
Time program	: B conc. 1% (0-2 min) → 40% (5.		-	
	→ 1% (6.35-7.5 min)	, , , , , , , , , , , , , , , , , , ,	,	G
Column temp.	: 50 °C	Injection vol.	: 10 μL	D
<lcms-8060></lcms-8060>				Y
Ionization	: ESI, Positive mode	DL temp.	: 200 °C	Ť
Interface temp.	: 400 °C	Heat block temp). : 350 °C	
Nebulizer gas	: 3.0 L/min	Heating gas	: 10 L/min	VE
Drying gas	: 10 L/min			Di

2-4. nSMOL sample preparation

The nSMOL kit was used for pretreatment for quantification of aflibercept in human plasma. After collection of aflibercept from 5 uL of sample with immunoglobulin collection resin peptide fragments were obtained by trypsin immobilized on nanoparticles. The protocol scheme is described Figure 1



3. Result

3-1. Prediction of aflibercept signature peptides

Figure 2 shows the peptide sequence of aflibercept. Aflibercept is composed of human VEGFR1, human VEGFR2 and IgG Fc region. The signature peptide candidates are underlined.

TGRPFVEM	YSEIPEIIHM	TEGRELVIPC	RVTSP <mark>N</mark> ITVT	LKK <u>FPLDTLI</u>	PDGKRIIWDS	60 <mark>-</mark>
_	-					
GFIIS <mark>N</mark> AT	YKEIGLLT <mark>C</mark> E	ATVNGHLYK <u>T</u>	NYLTHRQTNT	IIDVVLSPSH	GIELSVGEKL	120
	NVGIDFNWEY	DCCVIIOUVVI				100
	NVGIDENWEI	PSSKUQUKKL	VIIRDLK IQ5G	2EWKKL T21T	TIDGVIRDQ	180
YT <mark>C</mark> AASSG	LMTKK <mark>N</mark> STFV	RVHEKDKTHT	CPPCPAPELL	GGPSVFLFPP	KPKDTLMISR	240
EVT <mark>C</mark> VVVD	VSHEDPEVKF	NWYVDGVEVH	NAKTKPREEQ	YNSTYRVVSV	LTVLHQDWLN	300
		TOWAROODD				200
EIKCKVSN	KALPAPIEKT	ISKAKGQPRE	PQVYTLPPSR	DELIKNQVSL		360
AVEWESNG	QPENNYKTTP	PVLDSDGSFF	LYSKLTVDKS	RWQQGNVFSC	SVMHEALHNH	420
	~			~~		_
OKSLSLSP	G					

YTQKSLSLSP G

Figure 2. Peptide sequence of aflibercept.

/EGFR1 region: 1-104, VEGFR2 region: 105-205, IgG Fc region: 206-431, Disulfide bridge: Intrachain) 30-79, 124-185, 246-306, 352-410, Interchain) 211-211', 214-214' /cochain addition (potential): 36, 68, 196. The arrows show disulfie bonding.

Table 1. MS analysis of aflibercept.										Table	3. MRM tran	sition of af	flibercept.		
Region	Peptide	RT [min]	Observed <i>m/z</i>	Charge	Calculated m/z	Delta [ppm]	Area [cps]	Pe	eptide	Region	Μ	RM transit	tion (Collision [V]	Role
166-177	FLSTLTIDGVTR	27.6	661.8693	+2	661.8694	-0.0801	2,170,787	TNYLT	HR	90-96	452.7	75 >689.35	5 (y5+)	-19	Quantitation
56-61	IIWDSR	21.4	395.2163	+2	395.2163	-0.0076	1,444,997				452.7	75 >413.25	5 (y3+)	-22	Structure
97-119	QTNTIIDVVLSPSHGIELSV GEK	33.5	812.7707	+3	812.7709	-0.2596	979,639				452.7	75 >526.30) (y4+)	-20	Structure
1-24	SDTGRPFVEMYSEIPEIIHM TEGR	34.8	699.3345	+4	699.3349	-0.5834	897,543	IIWDS	۲	56-61	395.2	20 >563.25	5 (y4+)	-14	Quantitation
90-96	TNYLTHR	15.6	302.1591	+3	302.1594	-0.8638	480,001				395.2	20 >262.15	5 (y2+)	-21	Structure
157-164	TQSGSEMK	6.8	434.1976	+2	434.1975	0.1382	461,589				395.2	20 >227.20) (b2+)	-13	Structure
150-153	LVNR	7.7	251.1608	+2	251.1608	0.1155	399,945	FLSTL	TIDGVTR	166-177	661.8	35 >761.40) (y7+)	-23	Quantitation
44-54	FPLDTLIPDGK	29.6	608.3343	+2	608.3346	-0.4208	382,417				661.8	5 >1062.60) (y10+)	-24	Structure
128-144	TELNVGIDFNWEYPSSK	31.3	666.9858	+3	666.9863	-0.7826	350,038				661.8	35 >432.25	5 (y4+)	-35	Structure
63-72	GFIISNATYK	24.0	557.3005	+2	557.3006	-0.1992	288,647	P14R	lı lı	nternal stand	dard 512.	10 >292.30) (b3+)	-20	Quantitation
6-24	PFVEMYSEIPEIIHM TEGR	35.2	760.0335	+3	760.0343	-1.1000	228,865				512. ⁻	10 >660.40) (y6+)	-17	Structure
145-148	HQHK	1.2	183.7680	+3	183.7679	0.3646	94,214				512.7	10 >563.30) (y5+)	-17	Structure
157-165	TQSGSEMKK	5.0	332.4987	+3	332.4991	-1.2000	50,805								
44-55	FPLDTLIPDGKR	27.9	457.9255	+3	457.9259	-0.8953	44,064		Table 4. S	ummary of (CV% and acc	uracy (ave	erage, n = 7	7) of aflibercep	t.
145-149	HQHKK	1.2	226.4666	+3	226.4662	1.7221	17,632								
						Run	Concentratio	n TN	<u>YLTHR</u>		IWDSR	FLS	ILTIDGVTR		
	3-2. MS ana								[µg/mL]	CV%	Accuracy%	CV%	Accuracy	/% CV%	Accuracy%
	Aflibercept p	peptide wa	s identified usin	g Q-TOF	mass spectromet	er LCMS-903	0 and the	1	0.049	77.35	44.53	11.38	97.00	14.97	67.76
	identified peptides (with no cysteine residue containing) are shown in Table 1 in descending				lescending		0.15	31.64	69.58	2.76	87.13	12.07	76.53		
	order of sense	sitivity.							2.34	3.61	97.45	2.96	97.53	2.56	102.97
									160	3.52	96.37	3.49	101.17	7 3.22	96.79
		•	nd partial valida		'h			2	0.049	42.72	64.43	6.11	91.84	34.08	78.23
MRM was optimized using the full digest of aflibercept and transitions were selected by					•		0.15	16.43	53.00	4.51	91.57	6.15	101.64		
	MS/MS analysis using Q-TOF. Furthermore, a serial dilutions of aflibercept with human						2.34	9.31	96.10	6.50	98.43	6.19	102.44		
plasma were prepared and treated by nSMOL kit for evaluating the prepared MRM. There are					160	3.08	95.09	2.96	104.70) 3.13	97.41				
	two optimized buffers for enzyme reaction of nSMOL, thus we examined using each of them. The concentration of lower limit of detection of each peptide within 0.24 to 250 µg/mL were shown Table 2, and we have selected 3 candidate signature peptide (IIWDSR, TNYLTHR,						3	0.049	89.42	24.81	8.13	89.61	9.24	99.66	
)						•		0.15	16.15	69.06	9.74	86.24	10.41	92.89	
					cause retention ti	•			2.34	5.01	94.41	3.75	97.76	4.43	95.59
	unstable, it v	,					Populae 13		160	2.88	95.37	2.62	102.32	2 3.16	95.84



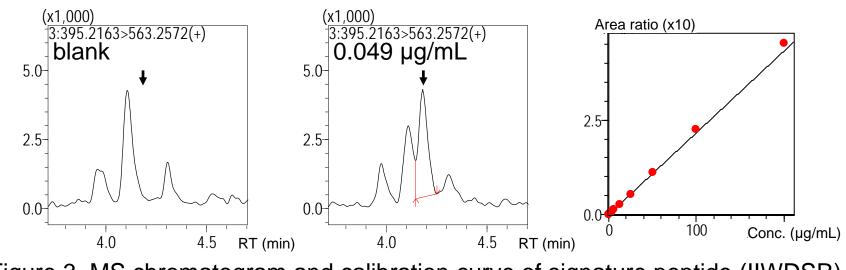
Region	Dentide	Limit of detection [µg/mL]				
	Peptide	reaction sol.				
56-61	IIWDSR	<0.24	<0.24			
90-96	TNYLTHR	<0.24	0.49			
166-177	FLSTLTIDGVTR	<0.24	0.49			
150-153	LVNR	<0.24	0.98			
44-55	FPLDTLIPDGKR	0.12	0.49			
63-72	GFIISNATYK	0.12	15.6			
128-144	TELNVGIDFNWEYPSSK	0.24	0.49			
157-164	TQSGSEMK	0.98	15.6			
157-165	TQSGSEMKK	0.98	62.5			
145-148	HQHK	31.25	31.25			
1-24	SDTGRPFVEMYSEIPEIIHM TEGR	31.25	125			
145-149	HQHKK	125	15.63			
97-119	QTNTIIDVVLSPSHGIELSV GEK	125	62.5			
44-54	FPLDTLIPDGK	125	-			
6-24	PFVEMYSEIPEIIHM TEGR	-	-			

3-4. Lower limit of quantification, CV% and accuracy The partial validation was in accordance with the Guideline on Bioanalytical Method Validation in Pharmaceutical Development of the Ministry of Health, Labor and Welfare. Because the lower limit of detection with reaction solution of these three selected peptides were 0.049 μ g/mL, partial validation was verified at 0.049 to 200 μ g/mL.



Table 2. Limit of detection of each peptide of aflibercept

As the QC sample, LLOQ, LOQ, MOQ, and HOQ were set as 0.049, 0.15, 2.34, and 160 ug / mL, respectively. The MRM transition, CV value and average of accuracy (n = 7) of each peptide are shown in Tables 3 and 4. From the table 5, since CV% and Accuracy% were achieved with IIWDSR, this was designated as the signature peptide. Figure 3 shows the IIWDSR MS chromatogram and calibration curve.



4. Conclusions

- and does not compete with the biological matrix.

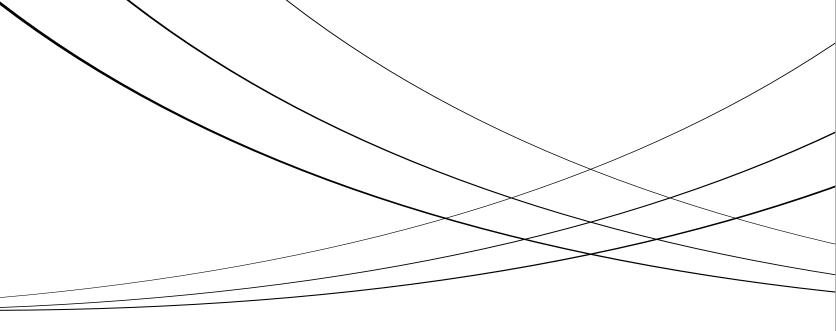


Figure 3. MS chromatogram and calibration curve of signature peptide (IIWDSR).

> We established a method to analysis of aflibercept with high sensitivity and high stability. > Aflibercept can be quantified in the range of 0.049 to 200 µg/mL with a small amount of specimen using IIWDSR as the signature peptide.

> It was confirmed that the signature peptide does not overlap with the endogenous protein