

Automated compound identification using product ion scanning with accurate mass measurement and compound database searching for non-targeted metabolomics

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1. Introduction

Non-targeted metabolomics entails data exploration to find the important metabolites from the detected features. Liquid chromatography mass spectrometry (LC-MS) is frequently used for non-targeted metabolomics because of the analytical sensitivity for a wide variety of compounds. In non-targeted metabolomics, compound identification is an important step to translate the information obtained from the instrument such as retention time and *m/z* into biologically relevant information such as chemical name and structure. We therefore developed a compound identification technique with scoring using both prediction formulae and assignment of product ions for narrowing the candidates. Using this approach, each candidate is evaluated not only using partial structural information from the spectral assignment, but also using all the molecular information provided by formula prediction.



Fig. 1 Schematic view of compound identification system

2. Methods and Materials

To evaluate the developed technique, we used dried green tea leaves which had been ranked by a sensory evaluation test. An aliquot of extracted samples was injected into an LCMS-IT-TOF system (Shimadzu Co.) with an ESI source. We applied this technique to find compounds which are important to the quality of the tea by constructing a quality prediction model using multivariate analysis. Formula Predictor (Shimadzu Co.) was used for formula prediction. These formulae were then used for database searching by an in-house developed searching interface and Application Programming Interface derived from ChemSpider. After predicting the list of candidate compounds, the score for each candidate was calculated based on mass accuracy and comparison of observed and predicted tandem mass spectra.

column	: Shim-pack XR-ODS (2.0 mm I.D. × 50 mm L., 2.2 µm)						
mobile phase A	: Water containing 0.1% formic acid						
mobile phase B	: Methanol						
gradient program : 2%B (0 min) – 60%B (10 min) – 98%B (10.01-14 min) – 2%B (14.01 – 19 min)							
flow rate	: 0.4 mL/min	column temp.	: 40°C				
ionization	: ESI (+/- switching)	scan range	: <i>m/z</i> 100 – 1000				
CDL temp.	: 200°C	BH temp.	: 200°C				

3. Result

3-1. Chromatograms of green tea extract

3742 peaks were detected from tea extract. These peaks were narrowed to 462 by filtering of isotopic peaks and p-value. This peak set was used to construction of tea quality evaluation model.



Fig. 2 Typical TIC chromatogram of green tea leaf extract.

3-2. Construction of tea quality evaluation model

To select compounds which shows importance to tea quality, quality evaluation model using PLS regression model were constructed. Compound identification for top 20 compounds which shows highest impact in variable importance in the projection plot were performed.



Fig. 3 Quality evaluation model of green tea

3-3. Compound identification

The result of compound identification for var_337 was shown here as an example. Formula prediction for var_337 was performed using MS¹⁻³ spectra (Figs. 4a-b). The score of the chemical formula (Formula Score) was calculated based on comparison of theoretical and observed *m/z* value and isotopic patterns using Formula Predictor. By using this formula list as a query for database searching, 218 candidate compounds were retrieved (Fig. 4c). The score of the assignment (Assignment Score) was calculated based on rate of assigned ion among product ion spectrum. As a result of automatic assignment of product ion spectrum, eight candidates received a highest score (Fig. 4d). Finally, 218 candidates were narrowed to 6 candidates by the scoring based on formula prediction and automatic assignment (Fig. 4e).



Fig. 4 Compound identification result for Var_337

ID	m/z	R.T. (min)	Candidate	Formula	lon	Final Score
UK -001	195.087	4.807	caffeine	C8H10N4O2	[M+H]+	94.24
UK -002	307.080	4.063	gallocatechin	C ₁₅ H ₁₄ O ₇	[M+H]+	87.86
UK -003	459.092	5.156	gallocatechin gallate	C22H18O11	[M+H]+	93.07
UK -004	443.097	6.392	catechin gallate	C22H18O10	[M+H]+	89.95
UK -005	261.169	5.429	1-(4-amino-6,7,8,9-tetrahydro-1h- imidazo[4,5-c]quinolin-1-yl)-2- methylpropan-2-ol	C ₁₄ H ₂₀ N ₄ O	[M+H]+	73.06
UK -006	345.080	1.003	theogalline	C ₁₄ H ₁₆ O ₁₀	[M+H]+	93.84
UK -007	339.106	5.719	coumaroyl quinic acid	C16H18O8	[M+H]+	82.14
UK -008	217.068	4.805	(Sodium ion adduct of UK-001)			
UK -009	261.169	4.034	1-(4-amino-6,7,8,9 -tetrahydro -1h- imidazo[4,5-c]quinolin -1-yl) -2- methylpropan -2-ol	C ₁₄ H ₂₀ N ₄ O	[M+H]+	75.44
UK -010	361.088	5.717	(Sodium ion adduct of UK-007)			
UK -011	273.074	6.393	(Fragment of UK -004)			
UK -012	365.159	4.4	ethyl-5-(acetylamino)-2,3,4,5-tetradeoxy- 2-methylidene-4-nitro-d-glycero-d- galacto-nononate	C14H24N2O9	[M+H]+	58.90
UK -013	291.086	3.958	catechin	C ₁₅ H ₁₄ O ₆	[M+H]+	93.65
UK -014	417.172	6.971	3-[[4-(2,4-dimethylphenyl)-5-(1- naphthylmethyl) -1,2,4-triazol -3- yl]sulfanyl]propanamide	C24H24N4OS	[M+H]+	59.30
UK -015	275.185	5.841	[4-amino-1-(2-methylpropyl)-6,7,8,9- tetrahydro-1h-imidazo[4,5-c]quinolin-2- yl]methanol	C15H22N4O	[M+H]+	76.53
UK -016	181.072	2.541	4-hydroxy -6-methyl -3,4-dihydropteridin - 2(1H)-one	C7H8N4O2	[M+H]+	86.80
UK -017	471.090	8.433	Luteolin 7-b-D-Glucopyranoside	C ₂₁ H ₂₀ O ₁₁	[M+Na]+	84.23
UK -018	565.157	7.027	3,4-dihydroxy-9,10-dioxo-9,10- dihydroanthracen-2-yl-6-O-(6-deoxy- alpha-L -mannopyranosyl) -beta-D- glucopyranoside	C ₂₆ H ₂₈ O ₁₄	[M+H]+	53.25
UK -019	579.150	4.123	(2r,3s)-2-(3,4-dihydroxyphenyl)-8-{2,3- dihydroxy-5-[(2r,3s)-3,5,7-trihydroxy-3,4- dihydro-2h-chromen-2-yl]phenyl}-3,4- dihydro-2h-chromene-3,5,7-triol	C ₃₀ H ₂₆ O ₁₂	[M+H]+	64.55
UK -020	307.083	1.834	gallocatechin	C ₁₅ H ₁₄ O ₇	[M+H]+	87.86

Table 2 Candidates representing the 20 components that most impact tea quality as determined by VIP value

4. Conclusions

The LCMS-IT-TOF provides positive and negative MSⁿ data with low variability. In this study, MS¹ data was used to generate a spectrally aligned data array of mass intensity and retention time pairs for multivariate analysis. A PLS regression was performed using this peak set and a tea quality contest ranking as the free and bound variables, respectively. The quality evaluation model was constructed successfully using a PLS regression model. Whole samples except test samples which ranked 10th, 20th and 30th places were used as the training set. We selected 20 features for compound identification which the quality prediction model indicated high importance. Thousands of candidates were initially returned by the database search. By using an automatic workflow involving formula prediction and product ion assignment, the number of candidates were narrowed successfully without non-trivial tasks such as the manual assignment of the product ion spectrum and literature searching. In addition to well-known compounds such as caffeine and catechins, these candidates include various esters of organic acids. This technique is not limited to the analysis of secondary metabolites as reported here, but is also applicable for the prediction of a wide range of compounds, including additives and impurities in polymers and pesticides.

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