Ion Chromatography Mass Spectrometry methods for cationic polar pesticides analysis

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Abstract

The hyphenation of Ion Chromatography to Triple Quadrupole Mass Spectrometry (IC-MS/MS) is increasing in popularity for the analysis of Polar Pesticides. Advances in the Quick Polar Pesticides (QuPPe) method of sample preparation has made this analysis fast, accurate and routine for anionic polar pesticides such as glyphosate, but the development of methods for cationic polar pesticides has fallen behind. Nevertheless, cationic polar pesticides may occur as residues in food, but are often excluded from pesticide monitoring programs due to the difficulty in the determination of these target analytes.

Introduction

Evaluation of target analytes

The EURL lists 39 target polar pesticides of interest, 28 of which can potentially have positive charge in acidic pH and are therefore good candidates for separation using a cation exchanger. All selected pesticides were run one-by-one on an IonPac CS17 column (2 x 250 mm) analytical column; the results are summarized in Table 2.

IonPac CS17 method optimization

A mixture of 11 pesticides were separated using gradient elution on the lonPac CS17column. Eluent flow rate, column temperature and eluent gradient were optimized as shown in Figure 2 and Table 3. Using the optimized method 16 polar pesticide along with 6 common cations were analyzed as shown in Figure 1. To test the response using IC-MS/MS the gradient was modified slightly to make the run-time faster but otherwise left unchanged. 13 cationic polar pesticides were evaluated using IC-MS/MS, see Figure

Figure 3. Gradient separation of polar pesticides on IonPac CS17 column with suppressed MS/MS detection



The EURL-SRM (European Union Reference Laboratory for pesticides requiring Single Residue Methods) lists 39 polar pesticides of interest, from which 23 fall into the category of cationic species based on their ionic properties and molecular structures. This work is aimed to demonstrate capabilities of several cation-exchange columns in the analysis of the listed cationic polar pesticides via ion chromatography.

Materials and methods

Sample preparation

Stock solutions (1000 mg/L in water) for all standards for target analytes were prepared by diluting the calculated amount of pure compound in reagent water. The standard solutions at lower concentrations were prepared by serial dilutions with reagent water.

Instrumentation

All chromatography work was performed using a Thermo Scientific[™] Dionex[™] ICS-6000⁺ ion chromatography system. Eluent was electrolytically generated using a Thermo Scientific[™] Dionex[™] x EGC MSA eluent generator cartridge.

Mass spectrometry was performed using a Thermo Scientific[™] TSQ Altis[™] Triple Quadrupole Mass Spectrometer equipped with a heated electrospray ionization (H-ESI) source operating in the positive ion mode. Data acquisition was performed in the selected reaction monitoring (SRM) mode. Spray voltage was set to 2.8 kV, sheath gas to 45 arbitrary units (Arb), auxiliary gas to 2.5 Arb, sweep gas to 2.0 Arb, ion transfer tube temperature set to 350°C, and vaporizer temperature set to 300°C. The collision gas pressure was set to 1.5 mTorr and the source fragmentation voltage to 10 V. All SRM transitions (parent, quantifier, and qualifier ions) were individually tuned for each target analyte by injecting the corresponding standard solution (100 μ g/L), as shown in Table 4.

Figure 1. Gradient separation of polar pesticides and 6 common cations on IonPac CS17 with suppressed conductivity detection



Table 2. Summary of suitability of target analytes for cation-exchange IC analysis

Analyte	Positive at low pH	Detection Method	Analyte	Positive at Iow pH	Detection Method
Ethephon	No	n.a.1	Daminozide	Yes	N.S.Cond. ⁵
HEPA	No	n.a.	Chlormequat	Yes	S.Cond.
Glufosinate	No	n.a.	Mepiquat	Yes	S.Cond.
N-acetyl-glufosinate	No	n.a.	Difenzoquat	Yes	n.d.
MPPA	No	n.a.	Propamocarb	Yes	S.Cond.
Glyphosate	No	n.a.	Melamine	Yes	S.Cond.
AMPA	Yes	n.d.	Diquat	Yes	S.Cond.
Phosphonic acid	No	n.a.	Paraquat	Yes	S.Cond.
N-acetyl AMPA	No	n.a.	N,N-Dimethylhydrazine	Yes	S.Cond.
Fosetyl-Al	Yes	n.d.	Nereistoxin	Yes	S.Cond.
Maleic hydrazide	Yes	UV	Streptomycin	Yes	S.Cond.
Perchlorate	No	n.a.	Kasugamycin	Yes	S.Cond.
Chlorate	No	n.a.	Morpholine	Yes	S.Cond.
Bialaphos	No	n.a.	Diethanolamine	Yes	S.Cond.
Cyanuric acid	Yes	UV	Triethanolamine	Yes	S.Cond.
Amitrol	Yes	UV	1,2,4-Triazole	Yes	S.Cond.
ETU	Yes	UV	Triazole-alanine	Yes	N.S.Cond.
PTU	Yes	n.d.	Triazole-acetic acid	Yes	UV
Cyromazine	Yes	UV	Triazole-lactic acid	Yes	UV
Trimesium	Yes	S.Cond. ⁴			
¹ n.a. – Not applicable	9				

²n.d. – Not detected/eluted under any tested condition ³UV – Non suppressed UV detection at 204 nm

⁴S.Cond. – Suppressed Conductivity detection

⁵N.S.Cond. – Non-Suppressed Conductivity detection

Figure 2. Temperature effect on the separation of some polar pesticides on IonPac CS17 column

Figure 3: Column : Dionex IonPac CS17 2 x 250 mm. Injection volume: 5 µL. Flow rate: 0.4 mL/min. Eluent: 0 min – 1.0 mM MSA, 4 min – 3.2 mM MSA, 10 min – 15 mM MSA, 14 min – 40 mM MSA. SRM transitions as shown in Table 4.

IonPac CS21-Fast-4µm method development

The Thermo Scientific[™] IonPac[™] CS21-Fast-4µm stationary phase column consists of a novel highly hydrophilic carboxylate functionality attached to the surface of a 4 µm macroporous polymeric substrate. This column has unique selectivity allowing the isolation of high levels of matrix ions such sodium, potassium, magnesium and calcium in two narrow windows, see Figure 4, while separating the four quaternary ammonium compounds outside these windows, see Figure 5.

Figure 4. Separation of the matrix components in a simulated matrix using the Dionex IonPac CS21-Fast-4µm column with suppressed conductivity detection.



Figure 1: Column : Dionex IonPac CS17 2 x 250 mm. Injection volume: 2 µL. Flow rate: 0.4 mL/min. Eluent: 0 min – 2 mM MSA, 8.5 min – 2 mM MSA, 14.5 min – 10 mM MSA, 20.5 min – 60 mM MSA

Results

Evaluation of off-the-shelf columns

In order to reduce the number of columns necessary for evaluation of cationic polar pesticides, existing cation exchange columns were reviewed with the following requirements.

- Weak cation exchanger. Many of the target analytes are divalent or polyvalent. The high affinity of strong cation exchangers for divalent analytes makes them unsuitable.
- Low to very low hydrophobicity. Many of the target analytes are polar or have high hydrophobicity. Thus, the column needs to have low or very low hydrophobicity.
- · Good for simple and complex amines. Most of the target analytes are amines or diamines. Columns that are targeted towards complex amines are preferred.
- **Compatible with IC-MS**. The target application is IC-MS/MS. Therefore, the column needs to have low bleed in order to ensure longevity of the ESI probe.

Table 1 shows available off-the-shelf weak cation exchange columns. The IonPac CS17 column was found to meet all requirements and was the most suitable column for further evaluation.

Table 1. Summary of weak cation-exchange columns characteristics and separation capabilities

Column	Capacity	Hydro-	Common	Simple	Complex	IC-MS



Figure 2: Column: Dionex IonPac CS17 2 x 250 mm. Injection volume: 2 µL. Flow rate: 0.4 mL/min. Eluent: 0 min – 2 mM MSA, 9.3 min – 2mM MSA, 16 min – 10 mM MSA, 22 min – 60 mM MSA.

Table 4. Selected reaction monitoring (SRM) transitions

Compound	Collision Energy	Transition Type	Parent Ion (<i>m/z</i>)	Product Ion (<i>m/z</i>)
Chlormequat	30	Quantifier Qualifier	122.1 122.1	57.9 62.9
Chlormequat-d4	30	Quantifier	126.0	57.9
Mepiquat	30	Quantifier Qualifier	114.1 114.1	98.1 58.0
Mepiquat-d16	30	Quantifier	130.0	110.0
Paraquat	19	Quantifier Qualifier	93.0 93.0	171.0 85.0
Paraquat-d8	19	Quantifier	97.0	179.0
Diquat	19	Quantifier Qualifier	92.0 92.0	84.5 157.1
Diquat-d8	19	Quantifier	96.0	88.5
Morpholine	23	Quantifier	88.0	44.0
Diethanolamine	16	Quantifier	106.0	70.0
Triethanolamine	15	Quantifier	150.0	132.0
Trimethylsulfonium	16	Quantifier	77.0	62.0
Nereistoxin	20	Quantifier	150.0	105.0
Melamine	19	Quantifier	127.0	85.0
Cyromazine	18	Quantifier	167.0	125.0
Propamocarb	20	Quantifier	189.0	102.0
Streptomycin	33	Quantifier	600.0	263.0

Figure 4: Column : Dionex IonPac CS21-Fast-4µm 2 x 250 mm. Injection volume: 10 µL. Flow rate: 0.3 mL/min. Eluent: 0 min - 3.0 mM MSA, 3.6 min - 3.0 mM MSA, 6.0 min - 22.0 mM MSA, 15 min -25 mM MSA.

Figure 5. Separation of the four quaternary ammonium polar pesticides chlormequat and mepiquat (Analyte Zone 1), and paraquat and diquat (Analyte Zone 2) using the Dionex IonPac CS21-Fast-4µm column with suppressed Mass Selective detection.



Figure 4: Column : Dionex IonPac CS21-Fast-4µm 2 x 250 mm. Injection volume: 10 µL. Flow rate: 0.3 mL/min. Eluent: 0 min - 3.0 mM MSA, 3.6 min - 3.0 mM MSA, 6.0 min - 22.0 mM MSA, 15 min -25 mM MSA. SRM transitions as shown in Table 4.

Conclusions

	(µ⊨q.)	phobicity	Cations	Amines'	Amines ²	compatible
IonPac CS12A	700	Med/Low	Excellent	Good	Poor	Yes
IonPac CS14	325	Med/High	Good	Good	Good	Yes
IonPac CS15	700	Med/Low	Excellent	Good	Poor	Yes
IonPac CS16	3,000	Medium	Excellent	Excellent	Poor	Yes
IonPac CS17	363	Very Low	Poor	Good	Excellent	Yes
IonPac CS18	290	Medium	Good	Excellent	Good	Yes
IonPac CS19	600	Medium	Poor	Good	Excellent	No
IonPac CS20	750	Medium	Poor	Excellent	Good	Yes

¹Hydrophobic amines such as methylamines and polar amines such as alkanolamines ²Polyvalent amines such as alkyldiamines and moderately hydrophobic amines such as biogenic amines

The IonPac CS17 column was found to be an excellent column for the screening of cationic polar pesticides. However, its inability to separate paraguat and diquat makes it unsuitable for the quantification of the four quaternary ammonium polar pesticides.

The lonPac CS21-Fast-4µm column was developed for the specific purpose of separating and thus quantification of the four quaternary ammonium polar pesticides by IC-MS/MS.

Methods were developed for both the IonPac CS17 column and IonPac CS21-Fast-4µm column.

Acknowledgements

We would like to thank Christopher Pohl for his invaluable input during the development of the IonPac CS21-Fast-4µm column.

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