A Multiresidue Method for Pesticide Analysis Using an Orbitrap Tribrid Mass Spectrometer and Automatic **Background Exclusion**

Figure 2. Obtained LOD (A) and LOQ (B) levels per number of analytes

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ABSTRACT

Purpose: Develop a multiresidue method for pesticides quantitation and improved untargeted screening on the Thermo Scientific™ Orbitrap ID-X™ Tribrid™ mass spectrometer

Methods: Chromatographic separation was performed on a Thermo Scientific™ Vanquish™ UHPLC system using a Thermo Scientific™ Accucore™ aQ C18 column

Mass spectrometric analysis was performed on an Orbitrap ID-X Tribrid mass spectrometer using AcquireX workflow, for automated generation of background exclusion list

mzVault library for pesticide screening

Results: Excellent limits of quantitation, reproducibility, linearity and accuracies were obtained

Data processing was performed with the Thermo Scientific™ TraceFinder™ software utilizing the

Automated generation of background exclusion list, using the AcquireX workflow, resulted in a significant increase in the number of identified pesticides, through libraries matches, compared to DDA, at low ppb concentration levels

INTRODUCTION

Pesticides are routinely applied to crops for preventing, destroying or controlling pest activity. In order to protect the consumers and ensure they are not being exposed to pesticide levels harmful for their health, pesticides are regulated and several countries have established maximum residue levels (MRLs). Given the large number of pesticides used and the globalization of the food industry, multiresidue methods offer a great advantage allowing analysis of hundreds of pesticides in a single run. We have implemented a multiresidue method for the analysis of 250 pesticides on an Orbitrap ID-X Tribrid mass spectrometer utilizing AcquireX for automated generation of background exclusion

MATERIALS AND METHODS

Sample Preparation

Strawberry samples were obtained from a local retail store. Following homogenization, strawberry samples were extracted using a QuEChERS extraction kit. Briefly 10 g of sample was weighed into the QuEChERS extraction tube and 10ml of ACN was added. Samples were shaken, centrifuged and the supernatant was collected. Matrix-matched standards were prepared by spiking the pesticide standards (250 pesticides) into the extracted matrices at concentration levels ranging from 0.05 to

Liquid Chromatography

Chromatographic separation was performed on a Vanguish UHPLC system using an Accucore aQ C18 column (100 × 2.1 mm, 2.6 µm). Mobile phase A consisted of 98% water, 2% methanol containing 5mM ammonium formate and 0.1% formic acid. Mobile phase B consisted of 98% methanol, 2% water containing 5mM ammonium formate and 0.1% formic acid. The flow rate was 300 µl/min and the column temperature was set at 25 °C. Analysis time was 15 min including 3 min equilibration time. Injection sample volume was 1 ul.

Mass Spectrometry

Mass spectrometric analysis was performed on an Orbitrap ID-X Tribrid mass spectrometer using AcquireX workflow, for automated generation of background exclusion list, or data dependent acquisition (DDA). The exclusion list was automatically generated from the matrix blank and automatically implemented in the acquisition method for all subsequent analyses to reduce background ion fragmentation. The AGC target value was set at 2e5 for the full MS and at 1e4 for the MS/MS scans. The resolution was set to 60,000 for the full MS scan and at 15,000 for the MS/MS scan events. HCD was performed with stepped collision energy (%) of 20, 40, and 70.

Data Processing

Data processing was performed with the TraceFinder software. Quantitation was performed on the precursor ion and mass tolerance was set to 5 ppm.

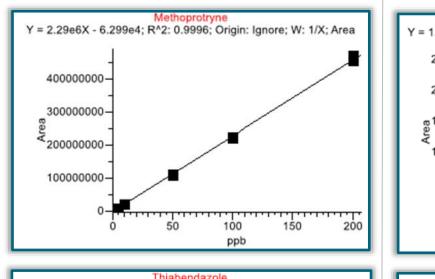
RESULTS

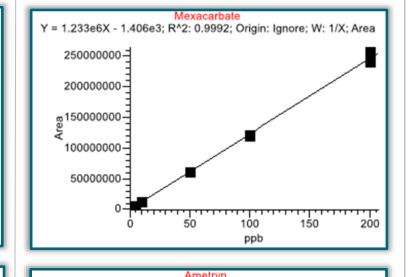
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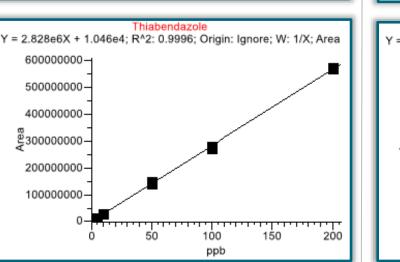
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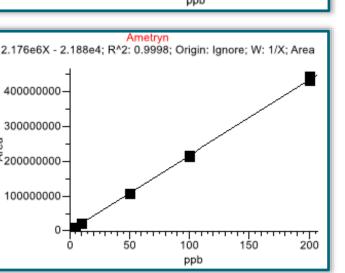
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Figure 1. Representative calibration curves







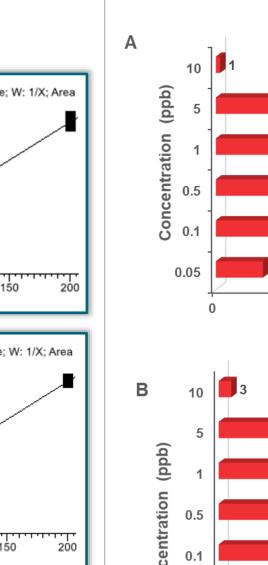


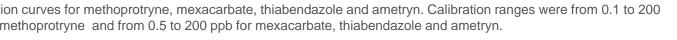
Calibration curves for methoprotryne, mexacarbate, thiabendazole and ametryn. Calibration ranges were from 0.1 to 200 ppb for methoprotryne and from 0.5 to 200 ppb for mexacarbate, thiabendazole and ametryn.

Table 1. Obtained % Diff, % RSD and % at LOQ levels for representative pesticide residues

Pesticide Residue	LOQ (ppb)	% Diff Injection 1	% Diff Injection 2	% Diff Injection 3	% RSD	% CV
Ametryn	0.5	-0.07	-6.27	-8.01	4.38	4.48
Carbaryl	1.0	4.14	-6.79	3.43	6.10	5.10
Chloridazon	1.0	-0.13	-3.33	8.17	5.84	5.86
Clomazone	0.5	0.54	4.46	-9.50	7.31	7.24
Cyanazine	0.5	-4.15	-1.42	-1.71	1.54	1.53
Cyazofamid	0.5	-4.48	-4.40	-7.92	2.13	2.01
Dicrotophos	0.5	3.66	2.87	9.13	3.24	3.62
Dinotefuran	1.0	2.14	8.90	1.67	3.88	4.88
Fensulfothion	0.5	1.08	-4.20	5.61	4.87	4.74
Fuberidazole	0.5	-1.24	7.81	1.97	4.46	4.47
Hexazinone	0.5	-5.17	-0.35	-2.01	2.51	2.55
Heptonophos	1.0	-0.37	3.72	-0.37	2.34	2.40
Methabenzthiazuron	0.5	3.38	6.83	2.54	2.18	2.13
Metosulam	1.0	-1.21	-1.81	-2.98	0.92	0.72
Ofurace	0.5	6.41	-1.51	-0.11	4.16	4.14
Tebufebpyrad	1.0	1.39	-5.94	6.06	6.02	5.60
Thiabendazole	0.5	1.49	-2.71	-5.99	3.84	3.81
Tricyclazole	0.5	-1.04	-4.31	-2.49	1.68	1.73

The % Diff represents the difference between the calculated amount and the expected amount, divided by the expected amount. % RSD is based on the calculated amount and % CV is based on the peak areas.

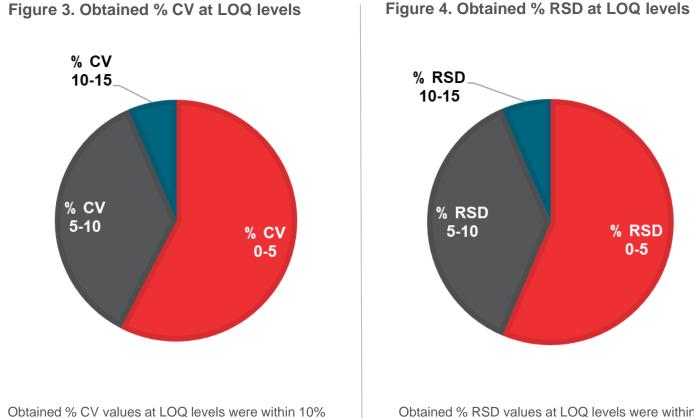




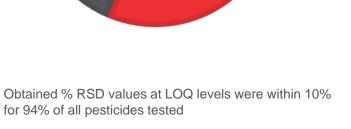
pesticide residues tested.

Number of Pesticide Residues

Number of Pesticide Residues

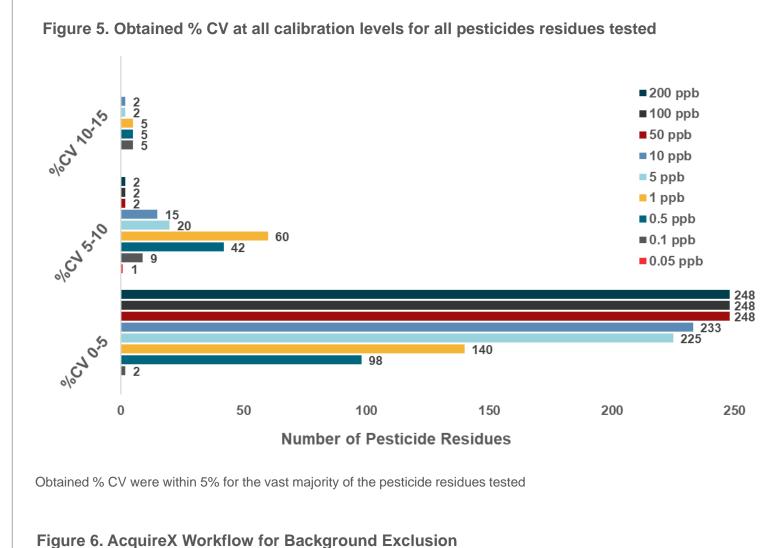


for 94% of all pesticides tested



% RSD

% RSD



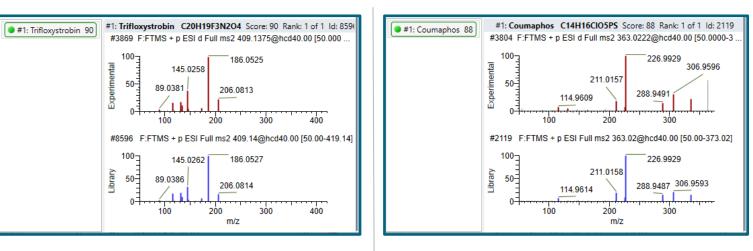


Exclusion List

Generation

Matrix Blank Injection

Figure 7. Representative Examples of mzCloud Spectral Library Matches



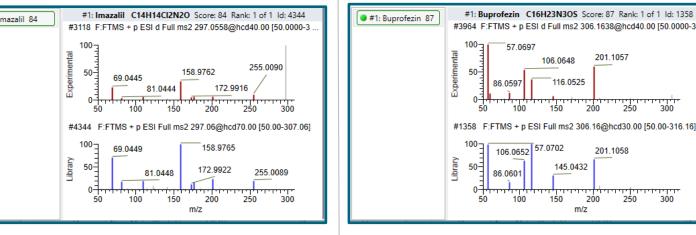
Instrument Method

Method to Exclude

106.0648 201.1057

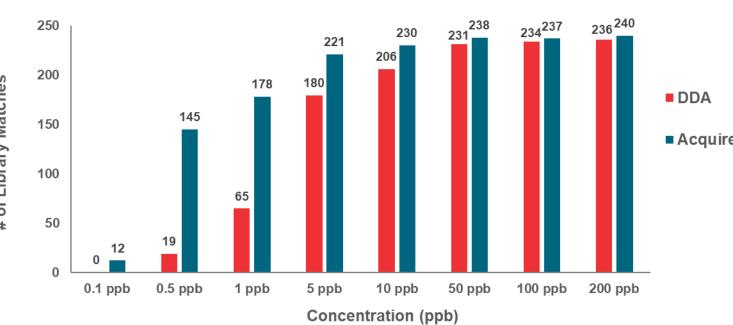
116.0525

Update with Exclusion list



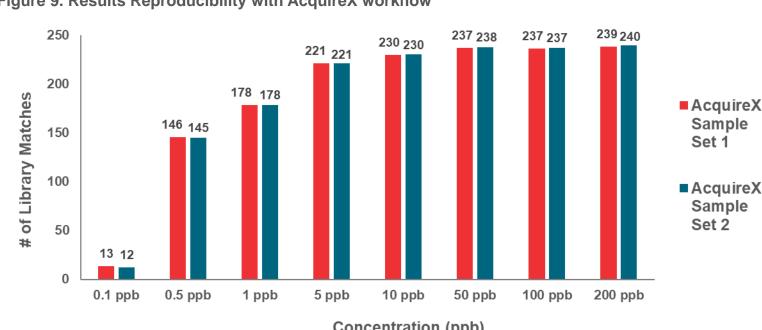
Spectra library matches for trifloxystrobin at a spiked concentration of 0.5 ppb, coumaphos at spiked concentration of 0.5ppb, imazalil at spiked concentration of 1 ppb and buprofeszin at spiked concentration of 1 ppb.

Figure 8. Number of Identified Pesticides with Background Exclusion using Acquire X and Data Dependent Acquisition (DDA)



Automated generation of background exclusion list with AcquireX workflow significantly increases the number of identified pesticides, compared to DDA, at low ppb concentration levels

Figure 9. Results Reproducibility with AcquireX workflow



Number of identified pesticides obtained with automated generation of background exclusion list with AxquireX workflow are highly reproducible. In the data presented above matrix-matched standards were prepared and analyzed on two different days

CONCLUSIONS

The Orbitrap ID-X Tribrid mass spectrometer delivers confident, accurate and reproducible

LOQ levels for 86% of the pesticide residues tested were at or below 1 ppb

AcquireX workflow enables the generation of background exclusion list in an automated fashion to increase productivity, efficiency and quality of results

Automated generation of background exclusion list with AcquireX workflow significantly increases the number of identified pesticide residues at low ppb levels compared to DDA

TRADEMARKS/LICENSING

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