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Evaluation of New Peak Detection Performance in MAM Using a Controlled Peptide Spike-in Strategy

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Introduction

The [Multi Attribute Method \(MAM\)](#) is widely adopted for monitoring of multiple product quality attributes (PQAs) in biotherapeutic products using LC/MS.¹ In addition to targeted monitoring of known critical quality attributes (CQAs), MAM workflows include New Peak Detection (NPD) to enable non targeted purity assessment.² NPD compares MS1 precursor ion features from a test sample against a reference standard to identify unexpected or emerging species. While highly powerful, implementation of NPD in regulated environments remains challenging due to the trade off between sensitivity and specificity. Lower detection thresholds increase the ability to detect low level impurities but also increase the risk of false positives arising from noise, adducts, or sample preparation variability. Regulatory guidance (USP <1060>, ICH Q2) emphasizes molecule specific optimization and robust performance characterization.³ However, standardized approaches for evaluating NPD performance are limited. In this study, we introduce a controlled peptide spike in strategy to systematically evaluate the sensitivity, specificity, and robustness of an NPD workflow implemented within a MAM software environment.

Experimental

A tryptic digest of the NIST mAb was used as the reference (0.5 µg on column). A PRTC peptide mixture was spiked into the NISTmAb digest at 0.5–4 pmol, corresponding to approximately 0.2–1.5% of the total digest. LC/MS analyses were performed under standardized peptide mapping conditions, with data acquired in positive ion mode on an Agilent 6230C TOF mass spectrometer. Full separation and acquisition parameters are provided in Table 1.

Table 1. LC/MS acquisition parameters.

LC Conditions	
Column	AdvanceBio Peptide Mapping C18, 2.1 x 150 mm, (p/n 653750-902)
Solvent A	Water with 0.1% FA
Solvent B	ACN with 0.1% FA
Gradient	0 min, 2% B 75 min, 36% B 75.5 to 77 min, 80%B 79 min, 35% B 79.5 to 80 min, 80% B 80.5 min, 2%B 84 min, 2%B
Flow rate	0.4 mL/min
Injection volume	4 ul
Column temperature	40 °C

Experimental

MS Conditions (G6230C)	
Ion Source	Agilent Dual AJS ESI source
Polarity	Positive
MS Scan Range	m/z 200 to 3000
Acquisition Rate	1 spectra/sec
Fragmentor	45 V
Skimmer	35 V
Data Storage	Profile
Drying Gas Flow	13 L/min
Nebulizer Pressure	35 psi
Sheath Gas Flow	12 L/min
Sheath Gas Temperature	300 °C
Capillary Voltage	3000 V
Nozzle Voltage	0 V
Gas Temperature	300 °C

Data processing:

Non-targeted MS data were processed using MAM software with an internally developed NPD algorithm. Differential analysis of MS1 features was performed between spiked samples and the NIST mAb reference using mass accuracy and retention time tolerances. Peak intensity thresholds were evaluated to balance true-positive detection and artifact control, while fold-change thresholds were applied as predefined criteria for feature classification. An overview of the workflow is shown in Figure 1.

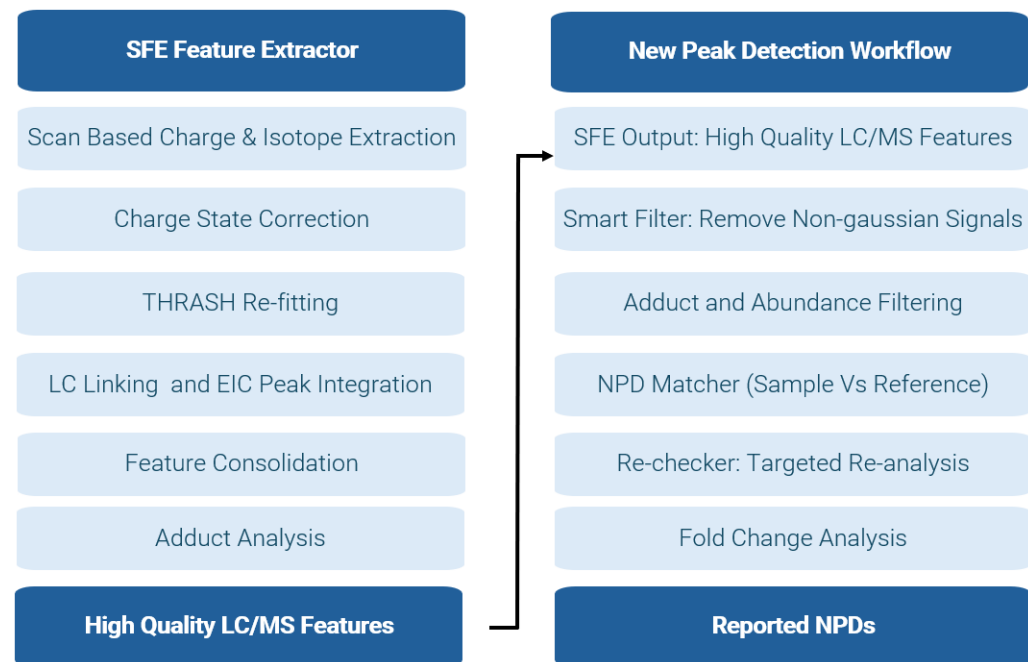


Figure 1. Untargeted SFE extracts and refines high quality LC/MS features from MS1 data using charge, isotope, LC linking, and adduct analysis. These features are processed by the NPD workflow using smart filtering, sample to reference matching, targeted re-analysis, and fold change criteria to report new peaks.

Results and Discussion

NPD Parameter Optimization

New Peak Detection Settings

Data Processing

Integration

Algorithm *
Agile2

Smoothing *
5

Manual Integration Boundary...
2

Feature Extraction

Peak Height *
1800

m/z range *
200 - 3000

Allowed Ion Species

☒ +H
☐ +Na
☐ +K
☐ +Fe
☐ +NH4

New Peak Detection

Limit of Detection *
Absolute 4500

Peak Detection RT Window *
10 - 50

Charge State *
2 - 3

Peak RT Tolerance *
1

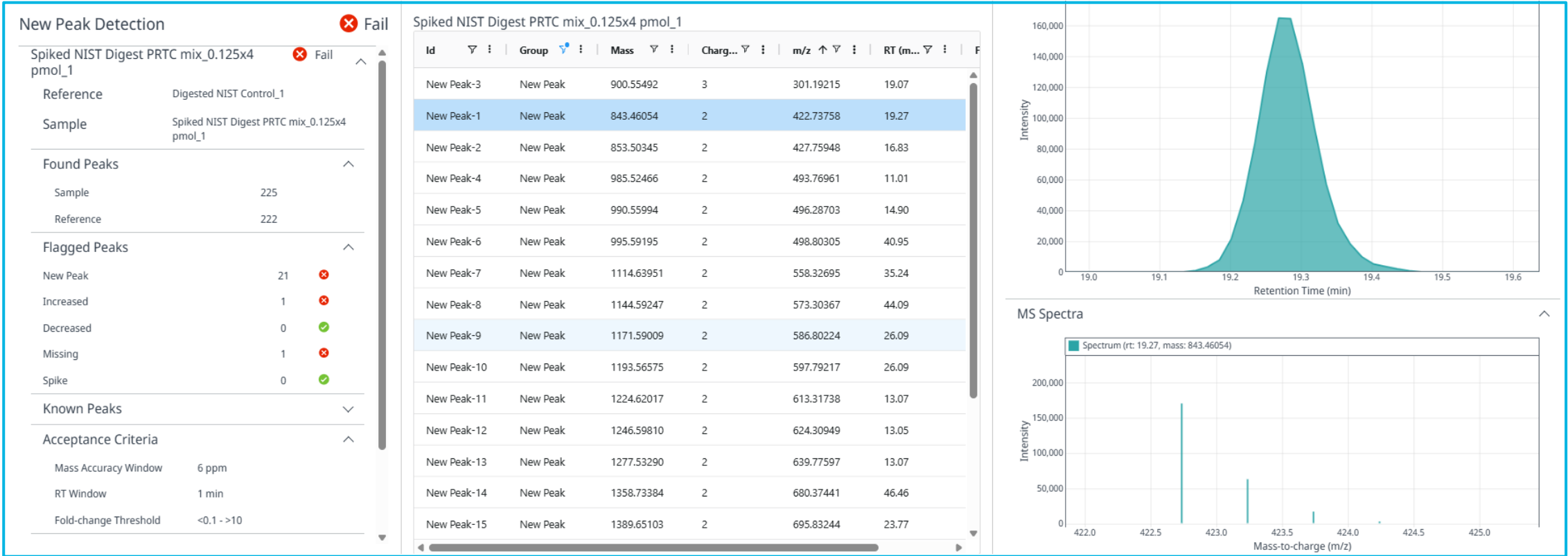
Peak m/z Tolerance *
6

New Peak Threshold *
Above and below threshold

0.1 - 10

Figure 2. Key NPD parameters were optimized to balance sensitivity for low-level impurity detection with control of false positive reporting.

NPD Results Visualization and Review Interface



Specificity Assessment of NPD Using Replicate Reference Samples

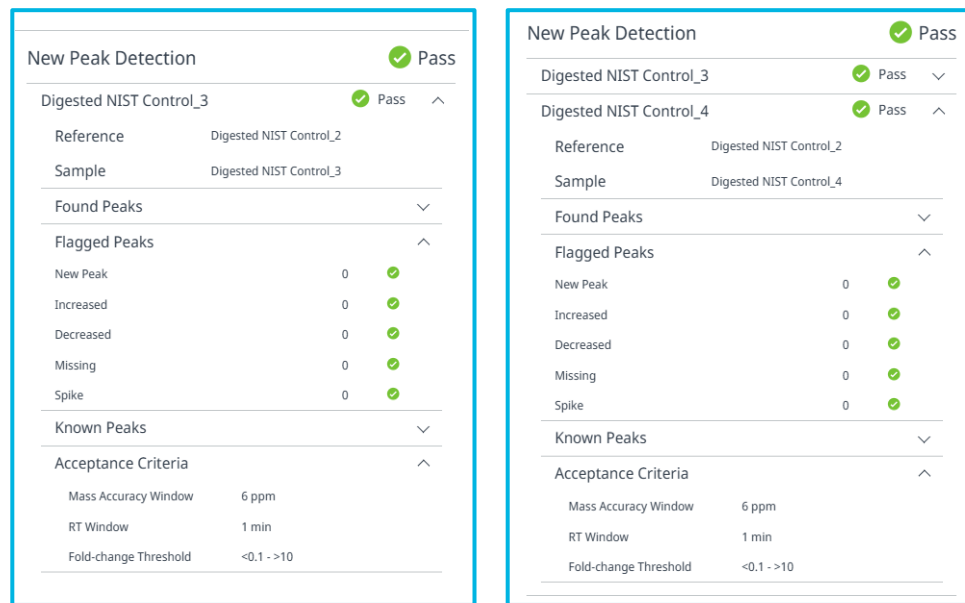


Figure 5. Comparison of replicate injections against a reference sample resulted in zero flagged features (New, Increased, Decreased, or Missing), demonstrating high specificity under optimized NPD parameter setting.

Fold-change validation: 2 pmol vs 1 pmol spiked samples.

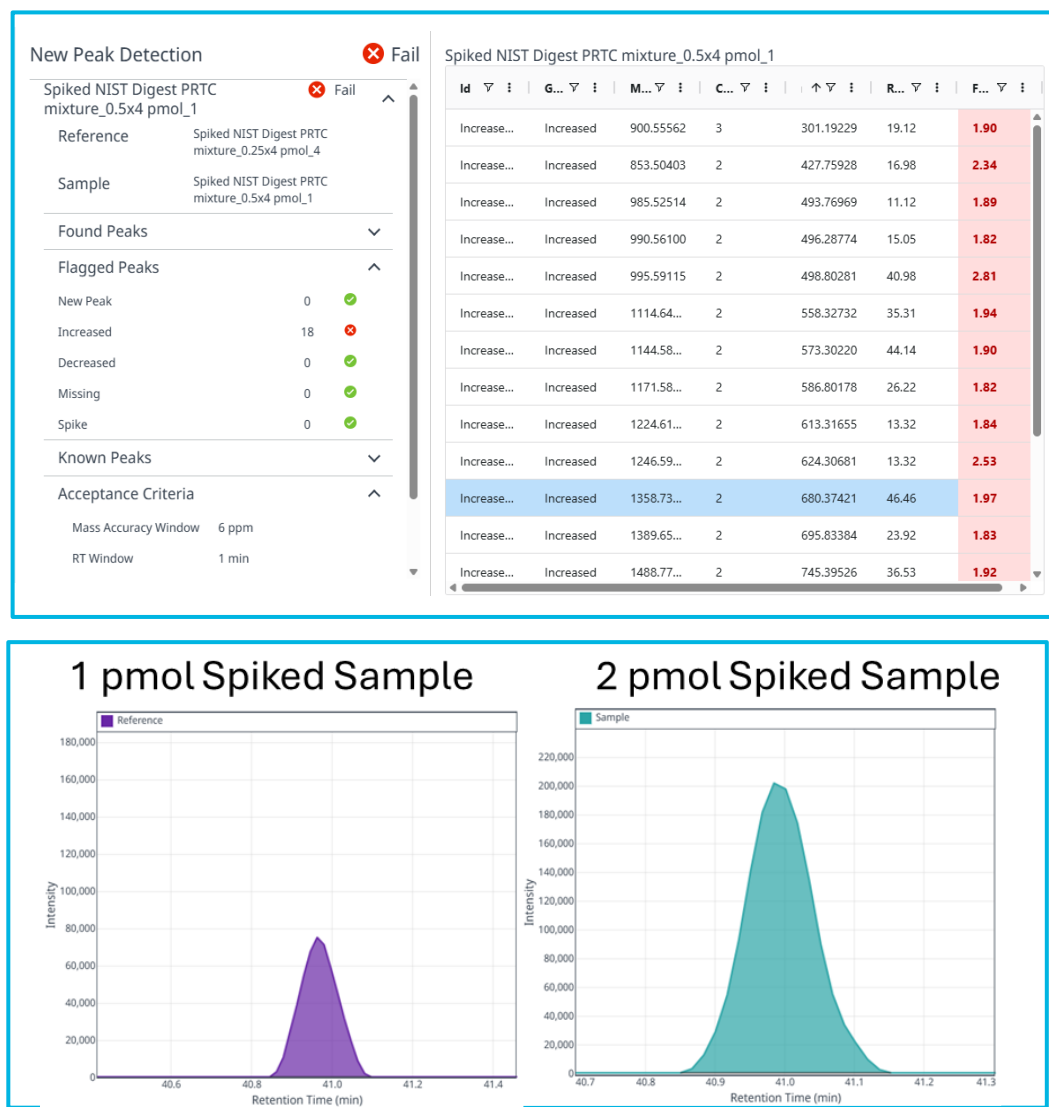


Figure 6. (Top panel) Table of features identified as increased in the 2 pmol sample relative to the 1 pmol sample. (Bottom panel) EICs for a representative peptide (m/z 680.3742) at 1 pmol and 2 pmol demonstrate a proportional increase in signal intensity. The observed fold change values are consistent with the expected ~2 increase, confirming accurate relative quantitation by the NPD algorithm.

NPD reporting and result summary in MAM

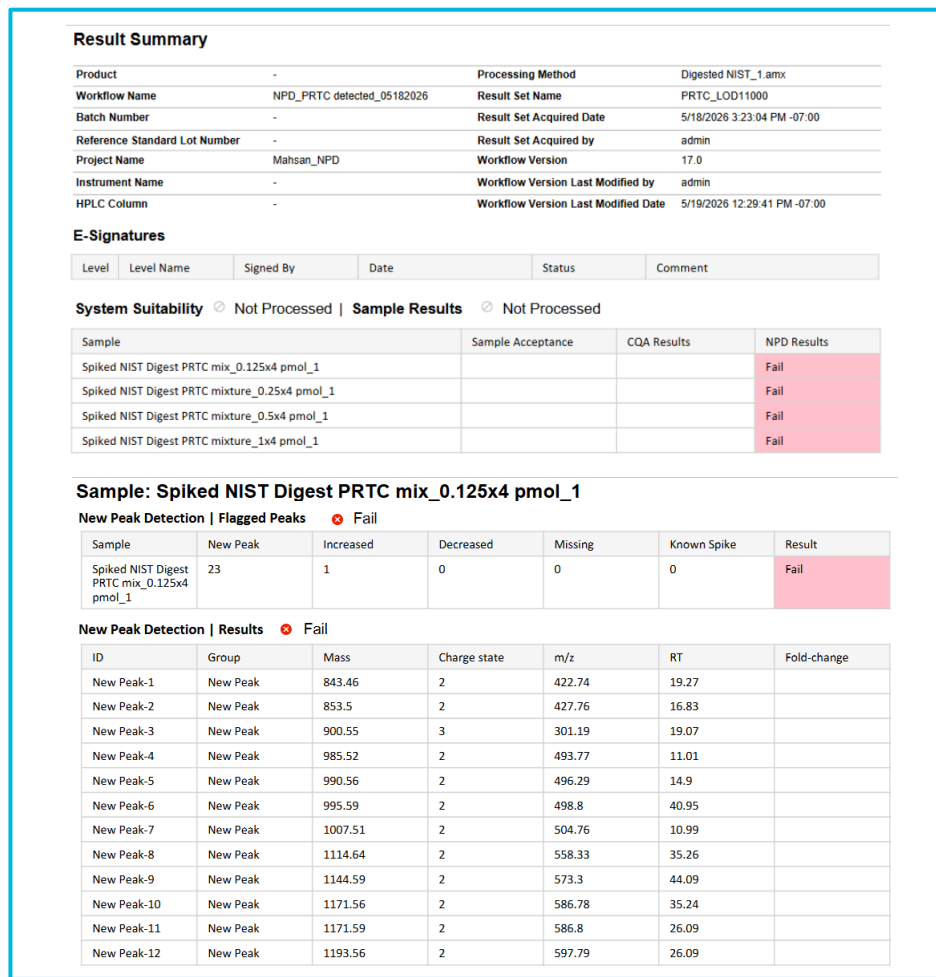


Figure 7. Structured NPD reporting provides summary results, flagged features, and supporting data for review and pass/fail determination.

Conclusions

- Controlled spike-in strategy enables systematic evaluation of NPD performance in MAM.
- Optimized LOD settings improve sensitivity, while higher thresholds can lead to false negatives.
- The workflow demonstrates high specificity with minimal false positives under optimized conditions.
- NPD preserves accurate fold-change measurements, supporting reliable trend analysis.
- Integrated reporting enables efficient review and decision-making for routine applications.

References

¹Rogers, R.S. et al., Anal. Chem., 2015, 87, 1058–1065.
²Ren, D. et al., mAbs, 2018, 10, 859–870.
³USP <1060>; ICH Q2(R2)